

CYTOTOXICITY EVALUATION of SOME PHENOLIC COMPOUNDS in V79 CELLS

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SUMMARY

Phenolic compounds play an important role on the growth and reproduction of plants and also exhibit a wide range of pharmacological activities such as anti-allergic, anti-atherogenic, anti-inflammatory and anticancer. But the data about toxicity of many phenolics are lacking and limited. The aim of this study was to evaluate the cytotoxic effects of five different commonly used phenolic compounds (galangin, limonene, naringin, puerarin and ursolic acid) which are widely used because of their health protecting effects by neutral red uptake (NRU) assay in Chinese hamster fibroblast cell line (V79). It is found that all of the studied phenolics decreased the cell viability of V79 cells in a dose dependent manner. The IC₅₀ values of galangin, limonene, naringin, puerarin and ursolic acid were found to be 104,36 µM, 10574 µM, 1976,4 µM, 51,71 µM and 92,94 µM, respectively.

Key Words: Galangin, limonene, naringin, puerarin, ursolic acid, neutral red uptake assay

BAZI FENOLİK BİLEŞİKLERİN SİTOTOKSİSİTESİNİN V79 HÜCRELERİNDE DEĞERLENDİRİLMESİ

ÖZET

Fenolik bileşikler bitkilerin büyüme ve gelişmesinde önemli rol oynarlar ve ayrıca antiallerjik, antiarterojenik, antienflamatuvar ve antikanser etkiler gibi çok sayıda farmakolojik aktivite gösterirler. Ancak pek çok fenolik maddenin toksisitesine ilişkin veri sınırlı ve az sayıdadır. Bu çalışmanın amacı, sağlığı koruyucu etkilerinden dolayı yaygın olarak kullanılan beş farklı fenolik bileşiğin (galangin, limonen, naringin, puerarin ve ursolik asit) Çin hamster fibroblast hücrelerinde (V79) sitotoksik etkilerinin nötral kırmızı alım (NKA) yöntemiyle değerlendirilmesidir. Çalışılan tüm fenolik bileşiklerin V79 hücrelerinin canlılığını doz artışıyla birlikte azalttığı bulunmuştur. Galangin, limonen, naringin, puerarin ve ursolik asit için IC₅₀ değerleri sırasıyla 104,36 µM, 10574 µM, 1976,4 µM, 51,71 µM and 92,94 µM olarak bulunmuştur.

Anahtar Kelimeler: Galangin, limonen, naringin, puerarin, ursolik asit, nötral kırmızı alım yöntemi

INTRODUCTION

Phenolic compounds are secondary metabolites of the derivatives of the pentose phosphate, shikimate, and phenylpropanoid pathways in plants [1]. These compounds play an important role on the growth and reproduction in plants [1] and afford protection against ultraviolet radiation, pathogens, and herbivores [2]. Phenolic compounds also exhibit a wide range of pharmacological activities such as anti-allergic, anti-atherogenic, anti-inflammatory, anti-microbial, antioxidant, anti-thrombotic, anticancer and cardioprotective [1]. Due to the cytotoxicity profile of many phenolic compounds, it is suggested that these chemicals can inhibit the survival of cancer cells. But the data about the cytotoxicity of these compounds in healthy cells are limited.

Galangin (3,5,7-trihydroxyflavone), belongs to a class of phenolic compounds known as flavonols [3]. It is present at high concentrations in propolis and in an Indian root, *Alpinia officinarum*, which is a common spice in Asia [4]. Galangin has been suggested to have biological activities including antimutagenic, antioxidant, anti-inflammatory, antiviral and anticarcinogenic [5, 6].

Limonene (p-mentha-1,8-diene), is the major component of oils obtained from orange, grape fruit and lemon. This monocyclic monoterpene has shown chemopreventive and therapeutic activity against a wide variety of experimental tumors [7]. Limonene has variety of uses such as industrial degreaser, household cleaning agent, food and cosmetic flavoring and pesticide [8].

Naringin (4',5,7-trihydroxy flavanone 7-rhamnoglucoside) is a flavanone also present in grape fruit [9], which exhibits diverse biological and pharmacological properties including anti-inflammatory, antioxidant, anticarcinogenic and lipid-lowering activities [10].

Puerarin (daidzein-8-C-glucoside) is the major isoflavone derived from the root of

Pueraria lobata (kudzu root) [11]. It is used in alternative medicine because of its estrogenic and antioxidant properties. In experimental models it is also suggested to be used in the prevention and treatment of cardiovascular diseases, diabetes, cancer and osteoporosis [12].

Ursolic acid (3 β -hydroxy-urs-12-en-28-oic acid) is a pentacyclic triterpenoid obtained from plants. It has long been used in Chinese medicine as anti-inflammatory, anti-arthritic, cytostatic and anti-proliferative, hepatoprotective agent [13].

The aim of this study was to evaluate the cytotoxic effects of these commonly used phenolic compounds (galangin, limonene, naringin, puerarin and ursolic acid) by neutral red uptake (NRU) assay in Chinese hamster fibroblast cell line (V79).

MATERIALS and METHODS

Chemicals

The chemicals used in the experiments were purchased from the following suppliers: fetal calf serum (FCS), trypsin-EDTA, penicillin-streptomycin, from Biological Industries (Kibbutz Beit-Haemek, Israel), minimum essential medium (MEM), dimethyl sulfoxide (DMSO), Triton X-100, phosphate buffered saline (PBS), ethanol, neutral red (NR), galangin, naringin, ursolic acid from Sigma (St Louis, USA), limonene and puerarin from Fluka (St. Gallen, Switzerland).

Cell Culture

Cells were seeded in 75 cm² flasks in 20 ml MEM supplemented with 10% FCS and 1% penicillin-streptomycin and then grown for 1 day in an incubator at 37°C in a humidified atmosphere supplemented with 5% CO₂.

Determination of Cytotoxicity by NRU Assay

The cytotoxicity of phenolic compounds was performed in V79 cell line by NRU assay following the protocols described by Virgilio et al. (2004) [14] and Saquib et al. (2012) [15].

Following disaggregation of cells with trypsin/EDTA and resuspension of cells in the medium, a total of 10^5 cells/well were plated in 96 well tissue-culture plates. After 24 h incubation, the different concentrations of galangin, limonene, naringin, puerarin and ursolic acid in medium were added. The cells were incubated for 18 h (1.5 cell cycle) at 37°C in 5% CO₂, then the medium was aspirated. The cells were washed twice with PBS and incubated for an additional 3 h in the medium supplemented with NR (50 µg/ml). After the medium was discarded, the cells were rinsed five times with warm PBS (37°C) to remove the nonincorporated excess dye and 200 µl of “fixation solution” (50% ethanol, 1% acetic acid, and 49% distilled water) was added to each well to fix the cells and bring NR into solution. The plates were shaken for 20 min, and the absorbance of the solution in each well was measured in a microplate reader at 540 nm and compared with the wells containing untreated cells. Results were expressed as the mean percentage of cell growth inhibition from three independent experiments. Cell viability was plotted as the percent of control (assuming data obtained from the absence of phenolic compounds as 100 %). IC₅₀ values represent the concentrations that reduced the mean absorbance of 50% of those in the untreated cells.

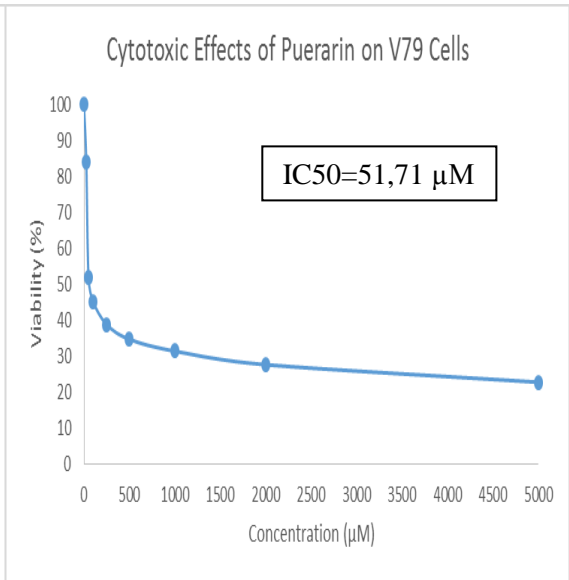
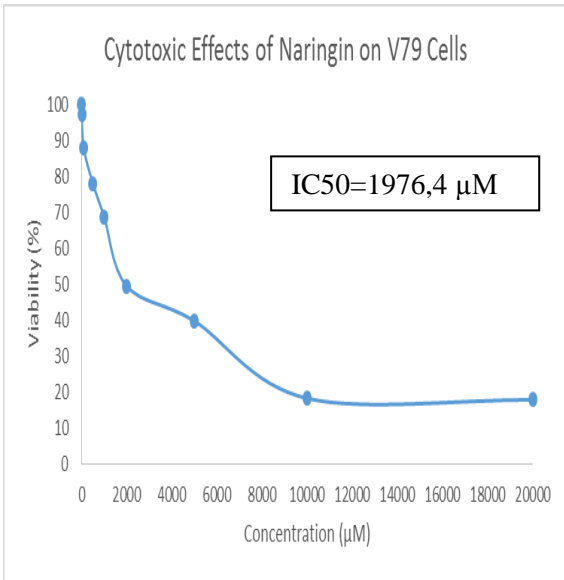
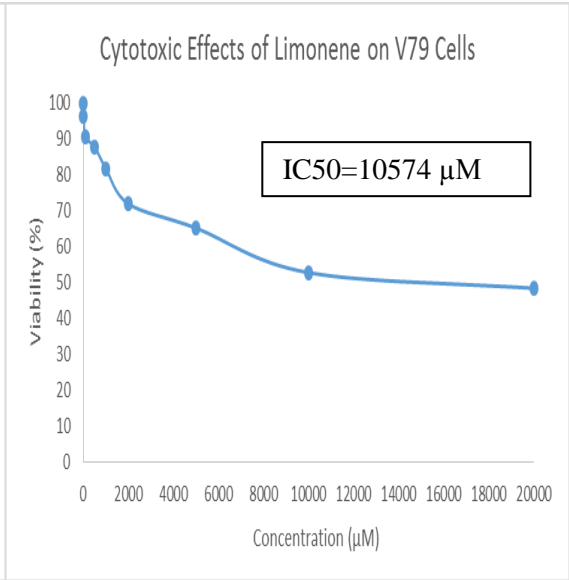
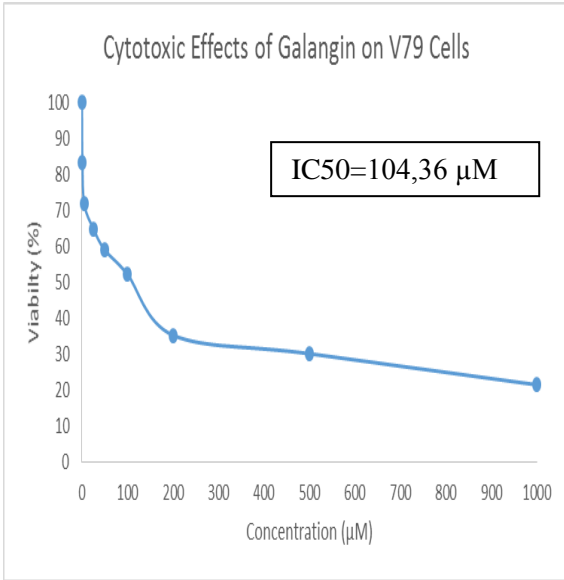
RESULTS

A concentration dependent decrease was seen in the survival of cells exposed to galangin, limonene, naringin, puerarin and ursolic acid (Figure 1). IC₅₀ values of galangin, limonene, naringin, puerarin and ursolic acid in V79 cells were found to be 104,36 µM, 10574 µM, 1976,4 µM, 51,71 µM and 92,94 µM respectively.

DISCUSSION

Several studies have showed that diets rich in fruits and vegetables can be associated with a markedly decreased risk of chronic diseases. This has been attributed to high levels of antioxidant contents of these foods [1]. Natural products are widely used as dietary supplements because of their potential antioxidant properties. On the other hand, it is suggested that at low doses phenolic compounds have antioxidant properties, but at high doses they can exert pro-oxidant effects[16].

The word “cytotoxicity” has a broad and sometimes vague meaning [17]. With regard to in vitro cell culture systems, a test substance is considered to be cytotoxic if it interferes with attachment of cells, significantly alters morphology, adversely affects the rate of cell growth, or causes cells to die [18]. There has been much attention devoted to cytotoxicity studies as a first step in evaluating the toxicity of test substances. This is especially valid in connection with screening biological activity of plant extracts and active compounds isolated from plants[19].



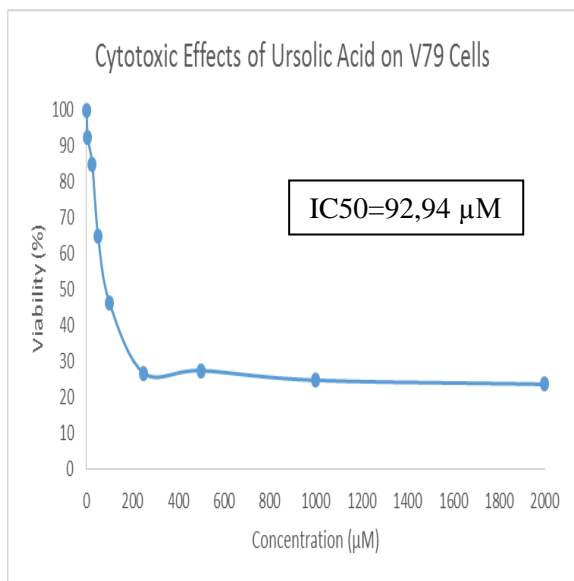


Figure 1. Cytotoxic Effects of Galangin, Limonene, Naringin, Puerarin and Ursolic Acid in V79 Cells

V79 cells, a well-defined cell line, are widely used in toxicity studies [20]. These cells are available for cytotoxicity studies. Because they do not have p450 enzyme system and they can keep basal cell functions in normal cell culture conditions [21].

Galangin, a member of the flavonol class of flavonoid, used in many countries for the management of various diseases, including airway affections, viral infections [5], and found to have antioxidant [22], anticancer [23] and antibacterial [24] activities. The data about the cytotoxicity of galangin is limited. In a previous study, gastric cancer (SNU-484) cells were incubated with galangin for 24 hours then cell viability was evaluated by MTT assay. The cell viability was decreased with the increasing concentrations of galangin and the IC₅₀ value of galangin was found to be 100 µM in these cell lines [25]. The cytotoxic effects of galangin were increased depending on the doses in liver cancer (HepG2 and PCL/RPF/5) cells and cell cycles of three cell lines were blocked by galangin [26]. In our study, we found that cell viability of V79 cells was decreased with the increasing doses of galangin and IC₅₀ value of galangin was found to be 104,36 µM. It is found that galangin has no toxic effects under 100 µM concentration in different cells.

Limonene, a monocyclic monoterpene, is the major constituent of *Citrus* fruits is widely used as a flavor and fragrance additive because of its pleasant citrus fragrance. Limonene is listed in the Code of Federal Regulation as generally recognized as safe (GRAS) for flavoring agent [27]. This monocyclic monoterpene has shown to be have chemopreventive and therapeutic activity against a wide variety of diseases [7]. The cytotoxic effects of *Citrus sinensis* essential oil was evaluated on human adenocarcinoma (SW480 and HT-29) cells. The cell viability was decreased at 6.25-500 ppm concentration with 24, 48 and 72 h incubations [28]. In a different cytotoxicity study with limonene, the cytotoxic effects were increased in a dose dependent manner in human lung adenocarcinoma (A549) and HepG2 cell lines and IC₅₀ values were found to be 586 µM and 889 µM, respectively. In our study, in healthy V79 cells the IC₅₀ value of limonene was found to be 10574 µM.

Studies about naringin have focused on its health preventive and antioxidant effects [29]. Our study is the first study about the cytotoxicity of naringin. In our study, IC₅₀ value of naringin was found to be 1976,4 µM. It is found that naringin has no toxic effects under 2000 µM concentration in V79 cell line.

Puerarin has also been suggested to have health promoting effects against

cardiovascular and gynecological diseases, osteoporosis and diabetic nephropathy [30]. There is limited data about the cytotoxicity of puerarin. In a study, it is claimed that puerarin has blocked proliferation of HT-29 cells in a dose and time dependent manner [31]. In our study, we found that cell viability of V79 cells was decreased in a dose dependent manner and IC50 value of puerarin was found to be 51,71 μ M.

Ursolic acid is naturally found in several medicinal herbs including rosemary, thyme, oregano and lavender. It possesses several biological activities such as anti-inflammatory, antiangiogenic, antiviral, antioxidant, antihypertensive, antihyperlipidemic and antihyperglycemic [32, 33]. In a study with colon cancer (CaCo-2) cells, ursolic acid decreased the cell viability at the higher concentrations in MTT assay [34]. In our study, we found that cell viability of V79 cells was decreased with the increasing doses of ursolic acid and IC50 value of ursolic acid was found to be 92,94 μ M.

In conclusion, in this study the cytotoxic effects of galangin, limonene, naringin, puerarin and ursolic acid were examined. All of the studied phenolics were decreased the cell viability of V79 cells with increasing dose in NRU assay. Further investigation such as using more cell lines and different reliable cytotoxicity assays and incubations with various concentrations at many time points should be performed to confirm beneficial and toxic effects of phenolics.

References

1. Balasundram N, Sundram K, Samman S. Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chem*99(1), 191-203, 2006.
2. Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *J Nutr Biochem*13(10), 572-584, 2002.
3. Jaiswal JV, Wadegaonkar PA, Hajare SW, The bioflavonoid galangin suppresses the growth of ehrlich ascites carcinoma in Swiss Albino mice: a molecular insight. *Appl Biochem Biotech*167(5), 1325-1339, 2012.
4. Bestwick CS, Milne L. Influence of galangin on HL-60 cell proliferation and survival. *Cancer Lett*243(1), 80-89, 2006.
5. Capasso R, Mascolo N. Inhibitory effect of the plant flavonoid galangin on rat vas deferens in vitro. *Life Sci*72(26), 2993-3001, 2003.
6. Capasso R, Tavares IA. Effect of the flavonoid galangin on urinary bladder rat contractility in-vitro. *J Pharm Pharmacol*54(8), 1147-1150, 2002.
7. Del Toro-Arreola S, Flores-Tolares E, Torres-Lozano C, Del Toro-Arreola A, Tostado-Pelayo K, Guadalupe Ramirez-Duenas M, Daneri-Navarro A. Effect of D-limonene on immune response in BALB/c mice with lymphoma. *Int Immunopharmacol*5(5), 829-38, 2005.
8. Ferrarini SR, Duarte MO, da Rosa RG, Rolim V, Eifler-Lima VL, von Poser G, Ribeiro VLS. Acaricidal activity of limonene, limonene oxide and β -amino alcohol derivatives on *Rhipicephalus (Boophilus) microplus*. *Vet Parasitol*157(1), 149-153, 2008.
9. Amudha K, Pari L. Beneficial role of naringin, a flavanoid on nickel induced nephrotoxicity in rats. *Chem-Biological Interact*193(1), 57-64, 2011.
10. Gaur V, Aggarwal A, Kumar A. Protective effect of naringin against ischemic reperfusion cerebral injury: possible neurobehavioral, biochemical and cellular alterations in rat brain. *Eur J Pharmacol*616(1), 147-154, 2009.

11. Gao Q, Yang B, Ye Z, Wang J, Bruce IC, Xia Q. Opening the calcium-activated potassium channel participates in the cardioprotective effect of puerarin. *Eur J Pharmacol* 574(2), 179-184, 2007.
12. Hou S-Z, Su Z-R, Chen S-X, Ye M-R, Huang S, Liu L, Zhou H, La X-P. Role of the interaction between puerarin and the erythrocyte membrane in puerarin-induced hemolysis. *Chem-Biological Interact* 192(3), 184-192, 2011.
13. Martin-Aragon S, Heras BI, Sanchez-Reus MI, Benedi J. Pharmacological modification of endogenous antioxidant enzymes by ursolic acid on tetrachloride-induced liver damage in rats and primary cultures of rat hepatocytes. *Exp Toxicol Pathol* 53(2-3), 199-206, 2001.
14. Virgilio ALD, Iwami K, Watjen W, Kahl R, Degen GH. Genotoxicity of the isoflavones genistein, daidzein and equol in V79 cells. *Toxicol Lett* 151(1), 151-162, 2004.
15. Saquib Q, Al-Khedhairy AA, Siddiqui MA, Abou-Tarboush FM, Azam A, Musarrat J. Titanium dioxide nanoparticles induced cytotoxicity, oxidative stress and DNA damage in human amnion epithelial (WISH) cells. *Toxicol In Vitro* 26(2), 351-361, 2012.
16. Wong WS, McLean AE. Effects of phenolic antioxidants and flavonoids on DNA synthesis in rat liver, spleen, and testis in vitro. *Toxicology* 139(3), 243-253, 1999.
17. Niles AL, Moravec RA, Riss TL. Update on in vitro cytotoxicity assays for drug development. *Expert Opin Drug Discov* 3, 655-669, 2008.
18. Horvath S. Cytotoxicity of drugs and diverse chemical agents to cell cultures. *Toxicology*, 16(1), 59-66, 1980.
19. McGaw LJ, Elgorashi EE, Eloff JN, 8 - Cytotoxicity of African Medicinal Plants Against Normal Animal and Human Cells. In *Toxicological Survey of African Medicinal Plants*, Ed: V. Kuete, pp. 181-233, Elsevier, Netherlands.
20. Cingi MR, De Angelis I, Fortunati E, Reggiani D, Bianchi V, Tiozzo R, Tucco N. Choice and standardization of test protocols in cytotoxicology: a multicentre approach. *Toxicol In Vitro*, 5(2), 119-125, 1991.
21. Rodriguez JA, Haun M. Cytotoxicity of trans-dehydrocrotonin from *Croton cajucara* on V79 cells and rat hepatocytes. *Planta Med*, 65(6), 522-526, 1999.
22. Choi JK, Kim SH. Inhibitory effect of galangin on atopic dermatitis-like skin lesions. *Food Chem Toxicol* 68, 135-41, 2014.
23. Ciolino H, Yeh G. The flavonoid galangin is an inhibitor of CYP1A1 activity and an agonist/antagonist of the aryl hydrocarbon receptor. *Brit J Cancer* 79(9-10), 1340, 1999.
24. Cushnie T, Lamb A. Assessment of the antibacterial activity of galangin against 4-quinolone resistant strains of *Staphylococcus aureus*. *Phytomedicine* 13(3), 187-191, 2006.
25. Kim D, Jeon YK, Nam MJ. Galangin induces apoptosis in gastric cancer cells via regulation of ubiquitin carboxy-terminal hydrolase isozyme L1 and glutathione S-transferase P. *Food Chem Toxicol* 50(3), 684-688, 2012.
26. Zhang H, Li N, Wu J, Su L, Chen X, Lin B, Luo H. Galangin inhibits proliferation of HepG2 cells by activating AMPK via increasing the AMP/TAN ratio in a LKB1-independent manner. *Eur J Pharmacol* 718(1-3), 235-244, 2013.
27. Sun J. D-Limonene: safety and clinical applications. *Altern Med Rev* 12(3), 2007.

28. Chidambara Murthy KN, Jayaprakasha GK, Patil BS. D-limonene rich volatile oil from blood oranges inhibits angiogenesis, metastasis and cell death in human colon cancer cells. *Life Sci*91(11), 429-439, 2012.
29. Kumar P, Kumar A. Protective effect of hesperidin and naringin against 3-nitropropionic acid induced Huntington's like symptoms in rats: possible role of nitric oxide. *Behav Brain Res*206(1), 38-46, 2010.
30. Fu-Liang X, Xiao-Hui S, Lu G, Xiang-Liang Y, Hui-Bi X. Puerarin protects rat pancreatic islets from damage by hydrogen peroxide. *Eur J Pharmacol*529(1), 1-7, 2006.
31. Yu Z, Li W. Induction of apoptosis by puerarin in colon cancer HT-29 cells. *Cancer Lett*238(1), 53-60, 2006.
32. Messner B, Zeller I, Ploner C, Frotschnig S, Ringer T, Steinacher-Nigisch A, Ritsch A, Laufer G, Huck C, Bernhard D. Ursolic acid causes DNA-damage, p53-mediated, mitochondria- and caspase-dependent human endothelial cell apoptosis, and accelerates atherosclerotic plaque formation in vivo. *Atherosclerosis* 219(2), 402-8, 2011.
33. Ovesna Z, Kozics K, Slaménova D. Protective effects of ursolic acid and oleanolic acid in leukemic cells. *Mutat Res*600(1-2), 131-7, 2006.
34. Ramos AA, Pereira-Wilson C, Collins AR. Protective effects of ursolic acid and luteolin against oxidative DNA damage include enhancement of DNA repair in Caco-2 cells. *Mutat Res*692(1), 6-11, 2010.