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Effect of *Physalis minima* L. on Interferon-γ and Interleukin-1β Levels in 7,12-Dimethylbenz[a]anthracene Administered Mice

Physalis minima'nın 7,12 Dimetilbenzantrasen Uygulanmış Farelerde Interferon-γ ve İnterlökin 1β Seviyelerine Olan Etkisi

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Abstract

Introduction: 7.12-Dimethylbenz[a]anthracene (DMBA) is a carcinogenic substance that can have immunosuppressive effects. *Physalis minima* L. is one of the herbal medicinal substances that was assumed to modulate immunosuppressive effects in DMBA-induced mice toward interferon γ (IFN- γ) and interleukin-1 β (IL-1 β).

Materials and Methods: Female BALB/c mice were given DMBA. DMBA-induced mice were administered with *Physalis* extract, once a day for two weeks. The level of IFN- γ and IL-1 β were measured using a flow cytometer.

Results: *Physalis minima* L. extract was able to increase (p<0.05) the level of IFN- γ in DMBA-given groups. Meanwhile, *Physalis minima* L. extract could increase the level of IL-1 β in group C+D1 (p<0,05).

Conclusions: Physalis minima L. extract could increase the level of IFN- γ and IL-1 β in DMBA Induced Mice

Keywords: 7.12-Dimethylbenz[a]anthracene, IFN-γ, IL-1β, *Physalis minima* L.

Öz

Giriş: Karsinojenik etkileri olan 7,12-Dimetilbenzantrasen (DMBA) aynı zamanda bağışıklık sistemini de baskılar. Physalis minima'nın bu baskılanmayı DMBA verilmiş farelerde interferon γ (IFN-γ) ve interlökin 1γ (IFN-1β) yolu ile düzenleyebileceği düşünülmektedir.

Gereçler ve Yöntemler: Dişi BALB/c farelere DMBA verildi. Bu DMBA uygulanmış deneklere ayrıca iki hafta boyunca her gün bir kez Physalis ekstresinden verildi. Bu deneklerin IFN-γ ve IFN-1β seviyeleri akan hücre ölçer kullanılarak ölçüldü.

Bulgular: Physalis minima L ekstresi, DMBA verilmiş farelerde IFN-γ ve IFN-1β seviyelerini artırdı (p<0,05). **Sonuçlar:** Phsalis minima ekstresi, DMBA uygulanmış farelerde IFN-γ ve IFN-1β seviyelerini artırmaktadır.

Anahtar Sözcükler: 7-12 Dimetilbenzantrasen, IFN-γ, IL-1β, *Physalis minima* L.

Introduction

7.12-Dimethylbenz[a]anthracene (DMBA) is a carcinogenic substance that was often used to induce cancer in mice or rats. DMBA consists of polycyclic aromatic hydrocarbons (PAHs) which has various structures and toxicity effects.^[1] The specific characteristic of PAHs is consisting of more than one fused aromatic rings.^[2] In the previous study, DMBA had been used to induce mammary cancer in Wistar rats.^[3] It was also used to induce skin tumor formation in C57BL/6 mice.^[4] In addition, DMBA was reported to be able to create an immunosuppressive microenvironment.^[5] The immunosuppressive microenvironment is frequently addressed to immunotoxicity where the immune system is suppressed.^[6]

Continuity of the immunosuppressive microenvironment may enhance the progression of cancer.^[7] An immunosuppressive environment suppresses the level of Interferon- γ (IFN- γ) and Interleukin-1 β (IL-1 β).^[8] IFN- γ and IL-1 β can support anti-tumor immunity.^[9] IFN- γ is a pro-inflammatory cytokine, predominantly produced by T-cell

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©2020 Turkish Journal of Immunology Available online at http://www.turkishimmunology.org lymphocytes.^[10] IFN- γ has a fundamental role in cellmediated immunity that might enhance anti-tumor effects by boosting cytotoxic T-cell lymphocytes.^[11] On the other hand, IFN- γ can enhance the expression of Major Histocompatibility Complex (MHC), which indirectly induce cytotoxic T-cell lymphocytes to recognize tumor cells.^[12] Similar to IFN- γ , IL-1 β is a member of IL-1 family which is a group of pro-inflammatory cytokines. ^[13] Specifically, IL-1 β promotes the activation of Th1 and Th17, which possess anti-tumorigenic effect.^[14]

Physalis minima L. is a member of solanaceae family which is often used as food and a herbal medicine.^[15] *Physalis minima* L. contains a lot of secondary metabolites such as physalins, withaphysalins, and phytosterols.^[16] The study shows that the pharmacological effect of *Physalis minima* L. is a potential source of immunomodulatory and anticancer activity.^[17] The immunomodulatory effects of *Physalis minima* L. is expected to modulate cytokines which has anti-tumorigenic effects.^[17]

The aim of this study is to investigate the effect of *Physalis minima* L. on IFN- γ and IL-1 β level on DMBA administered mice.

Methods

Animal Study and DMBA Administration

In this study, we used female BALB/c mice obtained from LPPT Gadjah Mada University, Yogyakarta. Mice were subcutaneously given with 15 mg/kg BW of DMBA at a dose of 15 mg/kg BW in the mammary gland. The DMBA administration was carried out for 6 weeks, once a week. The mice were divided into 5 groups, consist of N group: healthy mice; K: 15 mg/kg BW DMBA; C+D group: 15 mg/kg BW DMBA + 25 mg/kg BW *Physalis minima* L. extract; C+D2 group: 15 mg/kg BW DMBA + 50 mg/kg BW *Physalis minima* L. extract; C+D3 group: 15 mg/kg BW DMBA + 100 mg/kg BW *Physalis minima* L. extract.

Preparation of Physalis minima L. Extract

Physalis minima L. was obtained from UPT Materia Medica, Batu City, East Java. *Physalis minima* L. was extracted with methanol (w/v 100 mg/mL). The extraction process was done with maceration. *Physalis minima* L. extract was administered daily for 2 weeks.

Antibody Staining and Flow Cytometry

Spleens of the mice were removed, homogenized and centrifuged at 2.500 rpm at 10°C for 5 minutes. The obtained cell suspension was marked with anti-CD4-fluorescein isothiocyanate (FITC), then incubated at 4°C for 20 minutes. The suspension was added 50 μ l cytofix (BioLegend, San Diego, CA) was added then it was washed with 500 μ l washperm (Biolegend, San Diego, CA). The level of cytokines was stained with anti-IFN- γ , and anti-IL-1 β (Biolegend, San Diego, CA) antibodies. To optimize the bonding, antibody-stained cells were incubated at 4°C for 20 minutes, then 400 μ l phospahe buffer saline (PBS) was added. The antibody-stained cells were analyzed using a flow cytometer (BD FACS Calibur, USA).

Data Analysis

Data were analyzed with BD CellQuest ProTM software (USA). Statistical analysis was done by one-way ANOVA with significance threshold of 0,05 HSD Tukey's test was also used.

Results

IFN-7 Levels

IFN-γ levels were shown in Figure 1. The level of IFN-γ level in the Group C was decreased (p<0.05 compared to the Group N. It was decreased from 11.72% to 5.26%. Administration of *Physalis minima* L. extract increased (p<0.05) the level of IFN-γ in the C+D2 group and C+D3 group becoming 8.23% and 9.23%. Meanwhile, the C+D1 group had no statistically significantly different level of IFN-γ (p<0.05) compared to the C group.

IL-1 β Levels

The IL-1 β levels were shown in Figure 2. IL-1 β was decreased in groups C compared to group N (p<0.05). *Physalis minima* L. extract was able to increase IL-1 β in Group C+D1 (p<0.05). Yet, *Physalis minima* L. extract could not increase IL-1 β level in the Group C+D2 and Group C+D3 (p<0.05).

Discussion

The decrease in IFN- γ and IL-1 β in DMBA given mice is interpreted as immunosuppresive impact of DMBA. A number of studies showed that DMBA can suppress the



Figure 1. a, b. Administration of *Physalis minima* L. extract was able to increase the level of IFN- γ , in the C+D2 group and C+D3 group: The level of IFN- γ after was administered with *Physalis minima* L. extract for 2 weeks (**a**). Data were mean \pm SD (n=5) each group (p<0.05) (**b**) (N, healthy mice; C, DMBA-induced mice; C+D1, DMBA-induced mice + 25 mg/kg BW *Physalis minima* L. extract; C+D2, DMBA-induced mice + 50 mg/kg BW *Physalis minima* L. extract; C+D3, DMBA-induced mice + 100 mg/kg BW *Physalis minima* L. extract).

immune system.^[18] It works by reducing the immune and stromal cells inside spleen and bone marrow via apoptosis. ^[19] A recent study showed that there was a close link between immunosuppression and cancer development.^[20] Suppression of immune cells may lead to suppress cellmediated immune responses to eliminate cancer cells. ^[21] Therefore, immunosuppressive effects of DMBA can increase the susceptibility of cancer risk.^[22]

IFN- γ and IL-1 β were reported to be associated with tumor development, although several studies showed different results.^[23] IFN- γ has a role as central mediator of cell-mediated response.^[11] Studies showed that IFN- γ can enhance anti-tumor by increasing motility and killing ability of cytotoxic T-cells.^[11] IFN- γ is also involved in enhancing expression of major histocompatibility complex (MHC), both MHC class-I and MHC class-



Figure 2. a, b. Administration of *Physalis minima* L. extract can increase the level of IFN- γ , in the C+D2 and C+D3 groups: The level of IFN- γ after was administered with *Physalis minima* L. extract for 2 weeks (a). Data were mean \pm SD (n=5) each group (p<0.05) (b) (N, healthy mice; C, DMBA-induced mice; C+D1, DMBA-induced mice + 25 mg/kg BW *Physalis minima* L. extract; C+D2, DMBA-induced mice + 50 mg/kg BW *Physalis minima* L. extract; C+D3, DMBA-induced mice + 100 mg/kg BW *Physalis minima* L. extract).

II.^[24, 25] The up-regulation of MHC class-I is highly important for host response against tumor cells.^[26] Its response assists in cancer cells recognition of cytotoxic T-cells.^[27] Meanwhile, up-regulation of MHC class-II on antigen presenting cells (APCs) is more crucial for CD4⁺ T-cell function by boosting its peptide-specific activation. ^[28] IL-1 β specifically induces the activation of Th-1 cells. ^[29] The previous study showed that IL-1 β can eliminate tumor cells through exogenous injection method.^[30] It exerted its effect by increasing Th-1 mediated response against tumor cells.^[22]

The up-regulation of IFN- γ and IL-1 β levels after being treated proves that *Physalis minima* L. has immunomodulatory effects. Even though, *Physalis minima* L. only could show its effect at a certain dose. Immunomodulatory effects of *Physalis minima* L. may be originated from its phytochemical contents.^[31]

Conclusion

In summary, *Physalis minima* L. increased the levels of IFN- γ and IL-1 β in DMBA-administered mice. at a certain concentration. Thus, the best concentration is yet to be determined.

Ethics Committee Approval: The study has been approved by Brawijaya's University Ethics Commission (No. 1013-KEP-UB. Date: 12.09.2018)

Contribution of authors: We state that the research was carried out by Hary Isnanto, Fatiha Kamilah, Kurnilia Dewi Astuti, Citra Sefrilla Ramadhani, and Muhaimin Rifa'l. Hary Isnanto executed the study conception and drafting of manuscript. Fatiha Kamilah, Kurnilia Dewi Astuti, Citra and Sefrilla Ramadhani performed statistical analysis. Muhaimin Rifa'i performed the critical revision of the manuscript.

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Conflict of interest: There was no conflict of interest linked with this research.

References

- Hawkins, W. E.; Walker, W. W.; Overstreet, R. M.; Lytle, J. S.; Lytle, T. F. Carcinogenic Effects of Some Polycyclic Aromatic Hydrocarbons on the Japanese Medaka and Guppy in Waterborne Exposures. Sci. Total Environ. 1990, 94 (1–2), 155–167. [Crossref]
- Rengarajan T, Rajendran P, Nandakumar N, Lokeshkumar B, Rajendran P, Nishigaki I. Exposure to polycyclic aromatic hydrocarbons with special focus on cancer. Asian Pac J Trop Biomed 2015;5:182–9. [Crossref]
- **3.** Kirubha A, Anburajan M, Venkataraman B, Akila R, Sharath D, Raj B. Evaluation of Mammary Cancer in 7, 12-Dimethylbenz[a] anthracene-Induced Wister Rats by Asymmetrical Temperature Distribution Analysis Using Thermography: A Comparison with Serum CEA Levels and Histopathology. J Biomed Biotechnol 2012;2012:786417. [Crossref]
- 4. Xu C, Huang MT, Shen G, Yuan X, Lin W, Khor TO, et al. Inhibition of 7, 12-Dimethylbenz[a]anthracene-Induced Skin Tumorigenesis in C57BL/6 Mice by Sulforaphane Is Mediated by Nuclear Factor E2-Related Factor 2. Cancer Res 2006;66:8293–6. [Crossref]
- Karimi B, Ashrafi M, Shomali T, Yektaseresht A. Therapeutic effect of simvastatin on DMBA-induced breast cancer in mice. Fundam Clin Pharmacol 2019;33:84–93. [Crossref]
- Germolec D, Luebke R, Rooney A, Shipkowski K, Vandebriel R, van Loveren H. Immunotoxicology: A brief history, current status and strategies for future immunotoxicity assessment. Curr Opin Toxicol 2017;5:55–9. [Crossref]
- Chew V, Toh HC, Abastado JP. Immune Microenvironment in Tumor Progression: Characteristics and Challenges for Therapy. J Oncol 2012;2012:608406. [Crossref]
- Shini S, Huff GR, Shini A, Kaiser P. Understanding stress-induced immunosuppression: Exploration of cytokine and chemokine gene profiles in chicken peripheral leukocytes. Poult Sci 2010;89:841– 51. [Crossref]

- Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Pérez-Gracia JL, et al. Cytokines in clinical cancer immunotherapy. Br J Cancer 2019;120:6–15. [Crossref]
- Ni L, Lu J. Interferon gamma in cancer immunotherapy. Cancer Med 2018;7:4509–16. [Crossref]
- Bhat P, Leggatt G, Waterhouse N, Frazer IH. Interferon-γ derived from cytotoxic lymphocytes directly enhances their motility and cytotoxicity. Cell Death Dis 2017;8:e2836. [Crossref]
- 12. Zhang S, Kohli K, Black RG, Yao L, Spadinger SM, He Q, et al. Systemic Interferon-γ Increases MHC Class I Expression and T-cell Infiltration in Cold Tumors: Results of a Phase 0 Clinical Trial 8. Cancer Immunol Res 2019;7:1237–43. [Crossref]
- Koelman, L.; Pivovarova-Ramich, O.; Pfeiffer, A. F. H.; Grune, T.; Aleksandrova, K. Cytokines for Evaluation of Chronic Inflammatory Status in Ageing Research: Reliability and Phenotypic Characterisation. Immun. Ageing 2019, 16 (1), 11. [Crossref]
- Baker KJ, Houston A, Brint E. IL-1 Family Members in Cancer; Two Sides to Every Story. Front Immunol 2019;10:1197. [Crossref]
- Khan, M. A.; Khan, H.; Khan, S.; Mahmood, T.; Khan, P. M.; Jabar, A. Anti-Inflammatory, Analgesic and Antipyretic Activities of Physalis Minima Linn. J. Enzyme Inhib. Med. Chem. 2009, 24 (3), 632–637. [Crossref]
- 16.Wu, J.; Li, X.; Zhao, J.; Wang, R.; Xia, Z.; Li, X.; Liu, Y.; Xu, Q.; Khan, I. A.; Yang, S. Anti-Inflammatory and Cytotoxic Withanolides from Physalis Minima. Phytochemistry 2018, 155, 164–170. [Crossref]
- 17. Mirzaee F, Hosseini AS, Askian R, Azadbakht M. Therapeutic Activities and Phytochemistry of Physalis Species Based on Traditional and Modern Medicine. Research Journal of Pharmacognosy (RJP) 2019;6:79–96. [Crossref]
- 18. Ladics, G. S.; Kawabata, T. T.; White, K. L. Suppression of the in Vitro Humoral Immune Response of Mouse Splenocytes by 7,12-Dimethylbenz[a]Anthracene Metabolites and Inhibition of Immunosuppression by α-Naphthoflavone. Toxicol. Appl. Pharmacol. 1991, 110 (1), 31–44. [Crossref]
- 19. Teague JE, Ryu H-Y, Kirber M, Sherr DH, Schlezinger JJ, Proximal Events in 7, 12-Dimethylbenz[a]anthracene-Induced, Stromal Cell-Dependent Bone Marrow B Cell Apoptosis: Stromal Cell-B Cell Communication and Apoptosis Signaling. The Journal of Immunology 2010;185:3369–78. [Crossref]
- 20. Santangelo ML, Criscitiello C, Renda A, Federico S, Curigliano G, Dodaro C, et al. Immunosuppression and Multiple Primary Malignancies in Kidney-Transplanted Patients: A Single-Institute Study. BioMed Res Int 2015;2015:183523.
- Abdel-Shafy HI, Mansour MSM. A review on polycyclic aromatic hydrocarbons: Source, environmental impact, effect on human health and remediation. Egyptian J Petroleum 2016;25:107–23. [Crossref]
- **22.** Abba MC, Zhong Y, Lee J, Kil H, Lu Y, Takata Y, et al. DMBA induced mouse mammary tumors display high incidence of activating Pik3caH1047 and loss of function Pten mutations. Oncotarget 2016;7:64289–99. [Crossref]
- 23. Haabeth OA, Lorvik KB, Yagita H, Bogen B, Corthay A. Interleukin-1 is required for cancer eradication mediated by tumor-specific Th1 cells. Oncoimmunology 2016;5:e1039763. [Crossref]

- 24. Zhou, F. Molecular Mechanisms of IFN-Gamma to up-Regulate MHC Class I Antigen Processing and Presentation. Int. Rev. Immunol. 2009, 28 (3–4), 239–260. [Crossref]
- 25. Vardjan, N.; Gabrijel, M.; Potokar, M.; Švajger, U.; Kreft, M.; Jeras, M.; de Pablo, Y.; Faiz, M.; Pekny, M.; Zorec, R. IFN-γ-Induced Increase in the Mobility of MHC Class II Compartments in Astrocytes Depends on Intermediate Filaments. J. Neuroinflammation 2012, 9, 144. [Crossref]
- 26. Garrido, F.; Aptsiauri, N.; Doorduijn, E. M.; Garcia Lora, A. M.; van Hall, T. The Urgent Need to Recover MHC Class I in Cancers for Effective Immunotherapy. Curr. Opin. Immunol. 2016, 39, 44–51. [Crossref]
- 27. Messerschmidt, J. L.; Prendergast, G. C.; Messerschmidt, G. L. How Cancers Escape Immune Destruction and Mechanisms of Action for the New Significantly Active Immune Therapies: Helping Nonimmunologists Decipher Recent Advances. The Oncologist 2016, 21 (2), 233–243. [Crossref]

- 28. Castro F, Cardoso AP, Gonçalves RM, Serre K, Oliveira MJ. Interferon-Gamma at the Crossroads of Tumor Immune Surveillance or Evasion. Front Immunol 2018;9:847. [Crossref]
- 29. Duhen T, Campbell DJ. IL-1β promotes the differentiation of polyfunctional human CCR6+CXCR3+ Th1/17 cells that are specific for pathogenic and commensal microbes. J Immunol 2014;193:120–9. [Crossref]
- 30. Lee, P.-H.; Yamamoto, T. N.; Gurusamy, D.; Sukumar, M.; Yu, Z.; Hu-Li, J.; Kawabe, T.; Gangaplara, A.; Kishton, R. J.; Henning, A. N.; Vodnala, S. K.; Germain, R. N.; Paul, W. E.; Restifo, N. P. Host Conditioning with IL-1β Improves the Antitumor Function of Adoptively Transferred T Cells. J. Exp. Med. 2019, 216 (11), 2619–2634. [Crossref]
- 31. Sunitha K, Nagulu M, Srisailam K. Evaluation of Methanolic Extract of Physalis minima Fruits For Immunomodulatory Activity. Int J Pharmacy Biol Sci 2018;8:394–401. https://ijpbs. com/ijpbsadmin/upload/ijpbs_5bf6d068add0f.pdf