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Association of Interleukin-2-330T/G and Interleukin-10-1082A/G Genetic Polymorphisms with B-Cell Non-Hodgkin Lymphoma in a **Cohort of Egyptians**

Bir Mısırlı Hasta Kohortunda İnterlökin-2-330T/G ve İnterlökin-10-1082A/G Genetik

Polimorfizmlerinin B-Hücreli Hodgkin Dışı Lenfoma ile İlişkisi

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Abstract

Objective: Polymorphisms in the interleukin (IL)-2 and IL-10 genes are known to be associated with susceptibility to different immunedysregulated disorders and cancers such as non-Hodgkin lymphoma (NHL). To explore the possible association between IL-2-330T/G and IL-10-1082A/G single-nucleotide polymorphisms and the susceptibility to B-cell NHL (B-NHL) in Egyptians, we conducted a case-control study.

Materials and Methods: Genotyping of the studied genetic variations was done for 100 B-NHL patients as well as 100 age- and sex-matched healthy controls.

Results: The IL-2 variant allele occurred at a significantly higher rate in patients than controls and was associated with susceptibility to B-NHL [odds ratio (OR): 1.91, 95% confidence interval (CI): 1.28-2.85]. It was also associated with advanced performance status score. IL-2 polymorphism conferred an almost threefold increased risk of diffuse large B-cell lymphoma (OR: 2.64, 95% CI: 1.35-5.15) and a fourfold increased risk of indolent subtypes (OR: 4.34, 95% CI: 1.20-15.7). The distribution of IL-10-1082A/G genotypes in our patients was close to that of the controls. Co-inheritance of the variant genotypes of IL-2 and the common genotype of IL-10 conferred an almost sixfold increased risk (OR: 5.75, 95% CI: 1.39-23.72), while co-inheritance of the variant genotypes of IL-2 and IL-10 conferred fivefold increased risk of B-NHL (OR: 5.43, 95% CI: 1.44-20.45). The variant genotypes of IL-2-330T/G and IL-10-1082A/G had no effect on the disease-free survival of B-NHL patients.

Conclusion: The present study highlights the possible involvement of the IL-2-330T/G genetic polymorphism in the susceptibility to B-NHL in Egypt, especially indolent subtypes. Moreover, IL-10-1082A/G is not a molecular susceptibility marker for B-NHL in Egyptians.

Keywords: Interleukin-2-330T/G, rs2069762, Interleukin-10-1082A/G, rs1800896, B-cell non-Hodgkin lymphoma, Egypt



Öz

Amaç: İnterlökin (IL)-2 ve IL-10 genlerindeki polimorfizmlerin değişik immün bozukluklar ve Hodgkin dışı lenfoma (HDL) gibi kanserlere duyarlılık ile ilişkili olduğu bilinmektedir. Mısırlılardaki IL-2-330T/G ve IL-10-1082A/G tek nükleotid polimorfizmleri ile B hücreli HDL've (B-HDL) duyarlılık arasındaki olası ilişkinin araştırılması için bir olgukontrol çalışması yapılmıştır.

Gereç ve Yöntemler: Bahsedilen genetik varyasyonlar için, 100 B-HDL hastası ve yaş ile cinsiyet uyumlu 100 sağlıklı kontrole genotipleme yapıldı.

Bulgular: IL-2 varyant alleli hastalarda kontrollere göre anlamlı olarak daha yüksekti ve B-HDL duyarlılığı ile ilişkili bulundu [olasılık oranı (00): 1,91, %95 güven aralığı (GA): 1,28-2,85). Ayrıca bunun ileri performans skoru ile de ilişkili olduğu görüldü. IL-2 polimorfizminin diffüz büyük B hücreli lenfoma için yaklaşık üç kat (00: 2,64; %95 GA: 1,35-5,15) ve yavas sevirli (indolan) alt tiplerde dört kat artıs doğurmaktaydı (00: 4,34, %95 GA: 1,20-15,7). IL-10-1082A/G genotiplerinin dağılımı hastalar ve kontrollerde benzerdi. IL-2 varyant genotipleri ile IL-10'un sık rastlanan genotiplerinin es kalıtımı yaklasık altı kat artmış risk (00: 5,75, %95 GA: 1,39-23,72) yaratmaktayken, IL-2 ve IL-10 varyant genotiplerinin eş kalıtımı B-HDL riskinde beş kat artışa (00: 5,43, %95 GA: 1,44-20,45) neden olmaktaydı. IL-2-330T/G ve IL-10-1082A/G variant genotiplerinin B-HDL hastalarında hastalıksız sağkalım üzerine etkisi yoktu.

Sonuc: Bu çalışma Mısır'da, IL-2-330T/G genetik polimorfizmlerinin özellikle yavaş seyirli B-HDL'ye yatkınlık ile olası ilişkisini vurgulamaktadır. Ayrıca Mısırlılarda IL-10-1082A/G, B-HDL için duyarlı bir moleküler belirteç değildir.

Anahtar Sözcükler: İnterlökin-2-330T/G, rs2069762, İnterlökin-10-1082A/G, rs1800896, B-hücreli Hodgkin dışı lenfoma, Mısır

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Introduction

Despite the fact that there are some proven non-Hodgkin lymphoma (NHL) risk factors, the etiology of NHL still warrants extensive investigations [1]. Interleukin-2 (IL-2) has multiple opposing functions in the immune system. It plays a master role in T-cell growth and activation and in natural killer cell-mediated immune responses [2]. It has been reported to have antitumor effects through its contribution in the development of regulatory T cells, as well as expansion and apoptosis among activated T cells [3]. It is postulated that low production of IL-2 can suppress the antitumor response via the antibody-dependent cellular cytotoxicity (ADCC) seen in NHL patients, thus increasing the susceptibility to develop NHL [4].

IL-10 has both immunosuppressive and antiangiogenic functions. It thus has tumor-promoting as well as tumor-suppressing properties [5]. It may protect malignant cells through the inhibition of cytotoxic T lymphocyte-mediated tumor-specific cell lysis. Thus, IL-10 has an important role in carcinogenesis and it is postulated that it affects cancer risk, specifically for NHL [6]. The IL-10 promotor region may influence its expression and consequently alter susceptibility to NHL and disease outcome. It has been hypothesized that decreased production of IL-10 may increase the risk of NHL by less effectively downregulating the production of proinflammatory cytokines [7]. Accordingly, genetic factors that downregulate IL-10 production may provide a proinflammatory medium that favors lymphomagenesis [8]. However, other studies have hypothesized that IL-10, which is a B-cell stimulatory cytokine, could promote lymphomagenesis [9]. Therefore, these conflicting findings suggest that dysregulation in IL-10 in general could be a pivotal factor in NHL development. The aim of the current work was to study the possible role of IL-2-330T/G (rs2069762) and IL-10-1082A/G (rs1800896) singlenucleotide polymorphisms (SNPs) as genetic risk factors for B-cell NHL (B-NHL) in a group of Egyptian patients.

Materials and Methods

Study Population

This case-control study included 100 adult Egyptian B-NHL patients recruited from the Department of Medical Oncology, National Cancer Institute (NCI), Cairo University. These comprised either de novo cases or patients attending the NCI for follow-up. There were 54 males and 46 females. Their ages ranged between 20 and 83 years with a mean age of 52.7 years. One hundred unrelated age- and sex-matched volunteers were included in the study as a control group. The research protocol was approved by the Research Ethics Committee of the Kasr Al Ainy Faculty of Medicine, Cairo University. From all participants, informed consent was obtained in writing, and all procedures

were in accordance with the 1964 Helsinki Declaration. Diagnosis and subtyping of B-NHL was performed according to the World Health Organization classification of 2008. Patients were subjected to thorough clinical examinations, as well as laboratory investigations and radiological work-up for proper clinical assessment. The demographic and clinical features of the B-NHL patients are presented in Table 1.

Genotyping of IL-2-330T/G (rs2069762) and IL-10-1082A/G (rs1800896)

DNA extraction from peripheral blood leukocytes was done with the GeneJET Whole Blood Genomic DNA Purification Mini Kit (Fermentas Life Sciences, Canada) according to the manufacturer's instructions. Samples were stored in the elution buffer at -20 °C until use.

Detection of the IL-2-330T/G (rs2069762) SNP was performed with the polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) technique according to Cavet et al. [10]. The primer set used was as follows: forward, 5'-TAT TCA CAT GTT CAG TGT AGT TCT-3'; and reverse, 5'-AGA CTG ACT GAA TGG ATG TAG GTG-3'. Amplification was performed in a thermocycler (PerkinElmer 9700; PerkinElmer, USA) using the following program: 94 °C (5 min); then 30 cycles of 94 °C (1 min), 48° C (1 min), and 72 °C (1 min); and a final extension step for 8 min at 72 °C. This SNP abolishes the restriction site that can be recognized by the *Mael* restriction enzyme; accordingly, the T allele was restricted into two bands of 124 and 64 bp, while the G allele remained a 188-bp band.

Genotyping of the IL-10-1082A/G (rs1800896) SNP was performed by allele-specific PCR (ARMS) technique [11]. The following primers were used: F-5'-AGCAACACTCCTCGTCGCAAC, with either B1-5'-CCTATCCCTACTTCCCCC (G allele) or B2-5'-CCTATCCCTACTTCCCCT (A allele). The thermocycler program applied was 95 °C (10 min); then 30 cycles of 94 °C (30 s), 60 °C (1 min), and 72 °C (1 min); and a final extension step for 7 min at 72 °C. The AA genotype was identified by a single 153-bp band in tube B2, while the homozygous variant (GG) showed a 153-bp band in tube B1. The heterozygous variant (AG) was identified by a 153-bp band in both tubes. To validate our results, re-genotyping of 40 samples with respect to case-control status was performed. The results were interpreted blindly and found to be 100% concordant.

Treatment Regimen and Response to Therapy

All patients received the standard protocol treatment for NHL at the NCI of Cairo University. Diffuse large B-cell lymphoma (DLBCL) patients were treated according to stage and bulkiness. Non-bulky (<10 cm) stage I-II cases including extranodal presentations received 4 cycles of R-CHOP/21 days [rituximab at

Table 1. Demographic and clinical data of B-cell non-Hodgkin lymphoma patients at presentation and their response to therapy.

Item		B-NHL patients		
	NA I	(n=100)		
	Male	54/100		
Sex	Female	46/100		
B-symptoms: Feve loss	er, night sweats, weight	25/100		
Lymphadenopath	у	81/100		
	Cervical	61/100		
	Axillary	42/100		
	Inguinal	38/100		
Groups of lymph nodes involved	Abdominal	33/100		
noues involved	Para-aortic	24/100		
	Submandibular	20/100		
	Mesenteric	9/100		
Extranodal involv				
<2	79/100			
≥2		21/100		
Splenomegaly		40/100		
Hepatomegaly		38/100		
Clinical stage				
	23/100			
III & IV	77/100			
PS		-		
Score <2		61/100		
Score ≥2		39/100		
IPI risk group				
Low		29/100		
Intermediate/High		71/100		
IPI risk groups for	DLBCL subtype (n=78)			
Low/Intermediate Io	w (1, 2)	44/78		
Intermediate high/H	igh (3, 4)	34/78		
Histological aggre	essiveness			
Indolent		21/100		
Aggressive		79/100		
Regimen of treati	nent			
Chemotherapy		78/100		
Chemotherapy & rad	diotherapy	17/100		
No treatment		5/100		
Response to treatme	ent			
CR	59/100			
Non-CR	27/100			
PR		17/100		
PD		4/100		
SD		6/100		
Unavailable	14/100			

PS: Performance status, IPI: International Prognostic Index, CR: complete remission, PR: partial remission, PD: progressive disease, SD: stable disease, B-NHL: B-cell non-Hodgkin lymphoma.

375 mg/m², cyclophosphamide at 750 mg/m², doxorubicin at 50 mg/m², vincristine at 2 mg total dose, and prednisone at 100 mg for 5 days, followed by involved field radiotherapy (IFRT)]. Stage III or IV patients received 6-8 cycles of R-CHOP guided by the patient's response by positron emission tomography–computed tomography, which was done after 4 cycles. Patients with initial bulky disease received IFRT after their chemotherapy cycles. Follicular lymphoma of stage I and II was treated with IFRT only, while stages III and IV were treated if patients met the Groupe d'Etude des Lymphomes Folliculaires criteria for initiation of treatment. Mantle cell lymphoma patients were treated with R-CHOP alternated with R-DHAP. Therapeutic responses were assessed according to Oken et al. [12].

Statistical Analysis

Data management and analysis were performed using SPSS 21. Data were explored for normality using the Kolmogorov-Smirnov test and the Shapiro-Wilk test. Comparisons between groups for parametric numeric variables were done using the Student t-test, while for non-parametric numeric variables, comparisons were done by the Mann-Whitney U test. Chi-square or Fisher exact tests were used for comparing categorical data. For risk estimation, the odds ratio (OR) and 95% confidence interval (CI) were calculated. The Kaplan-Meier method was used to assess disease-free survival (DFS). Differences between survival curves were evaluated for statistical significance with the log-rank test. All p-values are two-sided and p<0.05 was considered significant.

Results

The genotypic and allelic frequencies of the IL-2-330T/G and IL-10-1082A/G SNPs in B-NHL patients and controls are presented in Tables 2 and 3. The genotypic distribution of the studied SNPs was in agreement with Hardy-Weinberg equilibrium (p>0.05).

The IL-2-330T/G variant genotypes (TG and GG) are associated with B-NHL risk, and the risk was higher for the indolent subtypes. Statistical comparison revealed that a performance status score of ≥2 was more common in patients harboring the variant genotypes (Supplementary Tables 1 and 2). The distribution of the variant genotypes of IL-10-1082A/G (AG and GG) did not differ between B-NHL patients and controls. Extranodal involvement of ≥2 sites was statistically more common in patients having the common genotype (Supplementary Tables 3 and 4). Combined genotype analysis showed that B-NHL risk increased almost sixfold in those having the variant genotypes of IL-2-330T/G and the common genotype of IL-10-1082A/G (AA), while co-inheritance of the variant genotypes of both SNPs was associated with fivefold increased risk of B-NHL (OR: 5.43, 95% CI: 1.44-20.45).

Table 2. Distribution of interleukin-2-330T/G and interleukin-10-1082A/G genotypes in B-cell non-Hodgkin lymphoma patients and controls.

Genotypes		Controls, n (%)	B-NHL patients, n (%)	OR	95% CI	p-value
	π	42 (42%)	20 (20%)	(1) Ref.		
	TG	26 (26%)	38 (38%)	3.07	1.48-6.37	0.003
IL-2-330T/G	GG	32 (32%)	42 (42%)	2.76	1.36-5.57	0.005
	TG & GG	58 (58%)	80 (80%)	2.90	1.54-5.44	0.001
	T allele	0.55	0.39	1.01	1 20 2 05	-0.001
	G allele	0.45	0.61	1.91	1.28-2.85	<0.001
	AA	28 (28%)	26 (26%)	(1) Ref.		
	GA	59 (59%)	51 (51%)	1.91	0.80-4.52	0.144
IL-10-1082A/G	GG	13 (13%)	23 (23%)	0.93	0.49-1.79	0.830
	GA & GG	72 (72%)	74 (74%)	1.11	0.59-2.07	0.750
	A allele	0.575	0.515	0.70	0.52.1.10	0.000
	G allele	0.425	0.485	0.79	0.53-1.16	0.228
Combined	TT/AA	3 (3%)	12 (12%)	(1) Ref.		
genotypes	TT/GA &/or GG	17 (17%)	30 (30%)	2.27	0.56-9.17	0.251
analysis, IL-2/IL-10	TG &/or GG / AA	23 (23%)	16 (16%)	5.75	1.39-23.72	0.016
	TG &/or GG / GA &/or GG	57 (57%)	42 (42%)	5.43	1.44-20.45	0.012
OR: Odds ratio, 95%	CI: 95% confidence interval, Ref.: refe	rence, B-NHL: B-cell non-Ho	odgkin lymphoma, IL: interleukin.			

Table 3. Distribution of interleukin-2-330T/G and interleukin-10-1082A/G genotypes in indolent and aggressive subtypes of B-cell non-Hodgkin lymphoma patients and controls.

Genotypes		Controls (n=100)	Indolent B-NHL (n=21)	OR (95% CI)	p-value	Aggressive B-NHL (n=79)	OR (95% CI)	p-value
IL-2-330 T/G	П	42 (42%)	3 (14.3%)	4.34 (1.2-15.71)	0.017	17 (21.5%)	2.64 (1.35-5.15)	0.004
1/0	TG & GG	58 (58%)	18 (85.7%)	(1.2-15.71)		62 (78.5%)	(1.35-5.15)	
IL-10-1082	AA	28 (28%)	3 (14.3%)	2.33	0.191	23 (29.1%)	0.95	0.870
A/G	GA & GG	72 (72%)	18 (85.7%)	(0.64-8.54)		56 (70.9%)	(0.49-1.82)	
OR: Odds ratio, 95%	CI: 95% confiden	ce interval, B-NHL: B	B-cell non-Hodgkin lymp	homa, IL: interleukin.				

Regarding the potential role of these SNPs as molecular prognostic markers, the 3-year and 5-year DFS rates were estimated. The 3-year DFS rate for the variant genotypes (GG or TG) of IL-2-330T/G was 65.4% versus 69.2% for the common genotype (TT), while the 5-year DFS rate for the variant genotypes (GG or TG) was 45.3% versus 69.2% for the common genotype (TT) with no statistically significant difference (p=0.211). The 3-year DFS rate for the variant genotypes (GG or AG) of IL-10-1082A/G was 60.7% versus 79.5% for the common genotype (AA), while the 5-year DFS rate for the variant genotypes (GG or AG) was 49.1% versus 39.8% for the common genotype (AA), which was statistically insignificant (p=0.205). Other potential prognostic

factors, such as the patients' age at diagnosis, sex, clinical stage, performance status, International Prognostic Index score, extranodal involvement, and histopathological subtypes, did not affect the DFS of our B-NHL patients (Supplementary Table 5).

Discussion

The relationship between the IL-2-330T/G SNP and NHL remains ambiguous. Some studies showed that the variant (G) allele correlates with decreased IL-2 production in vivo [13]. It has been suggested that reduced IL-2 levels may downregulate the antitumor response through ADCC and thus increase the risk of NHL [4]. In the present study, 38% of B-NHL patients had the

heterozygous genotype (TG), while 42% had the homozygous genotype (GG). These frequencies differed from those reported by Song et al. [4], being 56.2% and 12.7% for the TG and GG genotypes in Chinese NHL patients. This might be due to ethnicity. In the study presented here, the frequency of the variant genotypes was significantly higher in patients than controls and was associated with increased risk of B-NHL among Egyptians. This is in agreement with the study of Song et al. [4] involving Chinese patients.

The IL-2-330T/G polymorphism was associated with advanced performance status score. Otherwise, there was no association between the IL-2-330T/G SNP and sex, presenting symptoms, or other clinical and laboratory features, as well as response to therapy. Song et al. [4] could not find any association between the IL-2-330T/G SNP and clinical features in the Chinese patients in their study. Based on the clinical behavior of the disease, our patients were stratified into cases of indolent and aggressive lymphomas. IL-2-330T/G polymorphic genotypes were found to confer threefold increased risk of DLBCL, and the increase in risk for indolent B-NHL was fourfold.

Being an anti-inflammatory cytokine, the main functions of IL-10 are suppression of cytokine synthesis in Th1 cells as well as downregulation of cytotoxic and cell-mediated inflammatory responses [14]. It acts as an autocrine growth factor that upregulates BCL-2 expression in some cases of B-cell neoplasms [15]. High IL-10 levels were shown to be associated with poor outcomes and shorter survival in B-NHL patients [16,17].

Genetic polymorphisms in the promotor area of the IL-10 gene have been reported to influence IL-10 levels. IL-10-1082 common (A) and variant (G) alleles respectively correlate with low and high IL-10 expression levels [18]. Several studies have investigated the association of IL-10 gene polymorphisms and NHL susceptibility, reporting conflicting results. In the current study, 74% of B-NHL patients harbored this genetic variation, with 51% being heterozygous (AG) and 23% homozygous (GG). These frequencies agree with those previously reported in Australian patients, being 51% and 29% for AG and GG variant genotypes, respectively [19]. Similarly, Lan et al. [20] found the AG and GG genotypes in 52% and 23% of their female American B-NHL patients, and these frequencies were close to those of their controls. Extranodal involvement (i.e. the involvement of ≥2 extranodal sites) was more prominent in patients having the common genotype. Otherwise, there were no statistical differences between patients harboring the common or the variant genotypes. Lech-Maranda et al. [24] found that DLBCL patients harboring the variant genotypes had slightly higher complete remission (CR) rates. They stated that patients with

elevated cytokine levels had significantly lower CR rates.

IL-10-1082A/G variant genotypes (AG and GG) were not associated with susceptibility to either indolent or aggressive B-NHL subtypes. Similar results were reported by Talaat et al. [21], who concluded that IL-10-1082A/G polymorphic genotypes could not be considered as a genetic risk factor for DLBCL in Egyptians. Moreover, the studies of Kube et al. [22] and Berglund et al. [23] revealed that the IL-10-1082A/G SNP was not associated with susceptibility to aggressive B-NHL in German or Swedish populations, respectively. Contrary to our results, Purdue et al. [19] found that the frequency of the variant genotypes conferred increased risk of DLBCL. Lech-Maranda et al. [24] reported a similar frequency of the variant genotypes in France, which was statistically significant when compared to controls. They considered the IL-10-1082A/G SNP as a genetic risk factor for DLBCL in the French population. Lan et al. [20] stated that the GG homozygous variant genotype was significantly associated with an increased risk for DLBCL in female Americans. However, Cunningham et al. [25] reported that the low-producing IL-10-1082 AA genotype was significantly higher in patients with aggressive lymphoma compared to controls.

Combined genotype analysis showed that B-NHL risk was increased when IL-2-330T/G variant genotypes were co-inherited with either common or variant genotypes of IL-10-1082A/G. Accordingly, we assume that B-NHL risk can be attributed to the IL-2 rather than the IL-10 SNP.

Regarding DFS, none of the potentially known prognostic factors affected the DFS of B-NHL patients. Furthermore, the polymorphic genotypes of either IL-2-330T/G or IL-10-1082A/G had no effect on the 3- and 5-year DFS rates of these patients.

Study Limitations

The relatively small sample size of this study is a limitation of the present work. Larger sample size is recommended to validate our results regarding the role of the studied SNPs as molecular risk factors for B-NHL and to clarify their impact on therapeutic response and disease course. Furthermore, IL-2 and IL-10 levels should have been examined to conclude the association between the examined variations and NHL.

Conclusion

The current study highlights the possible involvement of the IL-2-330T/G SNP in susceptibility to B-NHL. Moreover, IL-10-1082A/G is not a molecular susceptibility marker for B-NHL in Egyptians.

Ethics

Ethics Committee Approval: The research protocol was approved by the Research Ethics Committee of the Departments of Clinical Pathology and Medical Oncology, Cairo University,

and all procedures were performed in accordance with the 1964 Helsinki Declaration.

Informed Consent: Informed written consent was obtained from all participants prior to enrollment in the study.

Supplementary Table 1. Comparison between B-cell non-Hodgkin lymphoma patients having wild genotype and polymorphic genotypes of interleukin-2-330T/G regarding their clinical data.

Item		IL-2 wild genotype (n=20)	IL-2 polymorphic genotypes (n=80)	n volue		
No (%)		No (%)	(11-00)	p-value		
Sex						
Male		12/20 (60%)	42/80 (52.5%)			
Female		8/20 (40%)	38/80 (47.5%)	0.547		
B-sympton		6/20 (30%)	19/80 (23.8%)	0.564		
Lymphader		17/20 (85%)	64/80 (80%)	0.610		
	Cervical	14/20 (70%)	47/80 (58.8%)	0.356		
	Axillary	8/20 (40%)	34/80 (42.5%)	0.939		
Groups	Inguinal	8/20 (40%)	30/80 (37.5%)	0.937		
of lymph nodes	Submandibular	6/20 (30%)	14/80 (17.5%)	0.211		
involved	Abdominal	8/20 (40%)	25/80 (31.3%)	0.457		
	Mesenteric	4/20 (20%)	5/80 (6.3%)	0.055		
	Para-aortic	8/20 (40%)	16/80 (20%)	0.061		
Extranodal	involvement					
<2		18/20 (90%)	61/80 (76.3%)	0.230		
≥2		2/20 (10%)	19/80 (23.7%)			
Splenomeg	aly	8/20 (40%)	32/80 (40%)	1.0		
Hepatomeg	galy	6/20 (30%)	32/80 (40%)	0.453		
Clinical sta	ge					
I & II		8/20 (40%)	15/80 (18.8%)	0.071		
III & IV		12/20 (60%)	65/80 (81.2%)			
PS						
Score <2		16/20 (80%)	45/80 (56.3%)	0.05*		
Score ≥2		4/20 (20%)	35/80 (43.7%)			
IPI risk gro	ир					
Low		9/20 (45%)	20/80 (25%)	0.078		
Intermedia	te/high	11/20 (55%)	60/80 (75%)			
_	ups for DLBCL subtype					
Low/Interm	ediate low (1, 2)	12/17 (70.6%)	32/61 (52.5%)	0.183		
Intermedia	te high/high (3, 4)	5/17 (29.4%)	29/61 (47.5%)	0.100		
Treatment	outcome					
CR		12/20 (60%)	47/80 (58.8%)			
Non-CR (PR, PD, SD)		5/20 (25%)	22/80 (27.5%)	0.971		
Unavailable		3/20 (15%)	11/80 (13.7%)			
Pathology		3/20 (15%)	18/80 (22.5%)			
Indolent		17/20 (85%)	62/80 (77.5%)	0.461		
Aggressive		, (55.75)	, (

^{*}p-value <0.05 = significant, PS: Performance status, IPI: International Prognostic Index, CR: complete remission, PR: partial remission, PD: progressive disease, SD: stable disease, IL: interleukin, DLBCL: Diffuse large B-cell lymphoma.

Supplementary Table 2. Comparison between B-cell non-Hodgkin lymphoma patients with wild genotype and those with polymorphic genotypes of interleukin-2-330T/G regarding their hematological data.

Item		IL-2 wild genotype			IL-2 polymorphic genotypes			p-value
		Range	Median	Mean ± SD	Range	Median	Mean ± SD	p-value
	Hb, g/dL	9.60-16.70	12.80	12.59±1.83	5-17	11.85	11.68±2.34	0.092
Hemogram	TLC, x10 ³ /cm ³	1.50-50	8.55	10.64±9.87	1.90-80	7.55	8.89±8.73	0.274
	Plts, x10 ³ /cm ³	133-491	300.50	281.25±113.37	14-675	278.50	295.40±138.87	0.829
LDH, IU/L		130-3531	239	499.45±751.66	135-3664	359	521.09±626.45	0.196
Hb: Hemoglobin, Tl	LC: total leukocyte count, LD	H: lactate dehyd	rogenase, Plts: platel	ets, IL: interleukin.				

Supplementary Table 3. Comparison between B-cell non-Hodgkin lymphoma patients with wild genotype and those with polymorphic genotypes of interleukin-10-1082A/G regarding their clinical data.

		IL-10 wild genotype	IL-10 polymorphic genotypes	
Item		(n=26)	(n=74)	p-value
		No. (%)	No. (%)	
Sex				
Male		11/26 (42.3%)	43/74 (58.1%)	
Female		15/26 (57.7%)	31/74 (41.9%)	0.164
Temate		10/20 (07.7.70)	01/71 (11.070)	
B-symptoms		7/26 (26.9%)	18/74 (24.3%)	0.792
Lymphadenopathy	,	22/26 (84.6%)	59/74 (79.7%)	0.585
	Cervical	15/26 (57.7%)	46/74 (62.2%)	0.688
	Axillary	13/26 (50%)	29/74 (39.2%)	0.337
	Inquinal	13/26 (50%)	25/74 (33.8%)	0.143
Groups of lymph	Submandibular	3/26 (11.5%)	17/74 (23%)	0.210
nodes involved	Abdominal	10/26 (38.5%)	23/74 (31.1%)	0.491
	Mesenteric	1/26 (3.9%)	8/74 (10.8%)	0.439
	Para-aortic	4/26 (15.4%)	20/74 (27%)	0.232
Extranodal involvem		1,720 (1011.10)	25,7.1 (27.76)	0.202
<2	ciic	16/26 (61.5%) 63/74 (85.1%)		
 ≥2		10/26 (38.5%)	11/74 (14.9%)	0.011*
Splenomegaly		10/26 (38.5%)	30/74 (40.5%)	0.852
Hepatomegaly		10/26 (38.5%)	28/74 (37.8%)	0.955
Clinical stage				
I & II		4/26 (15.4%)	19/74 (25.7%)	0.000
III & IV		22/26 (84.6%)	55/74 (74.3%)	0.283
PS				
Score <2		14/26 (53.9%)		
Score ≥2		12/26 (46.1%)	27/74 (36.5%)	0.385
IPI risk group				
Low		4/26 (15.4%)	25/74 (33.8%)	0.075
Intermediate/High		22/26 (84.6%)	49/74 (66.2%)	0.07.0
IPI risk groups for D	LBCL subtype			
Low/Intermediate lov		12/23 (52.2%)	32/55 (58.2%)	0.626
Intermediate high/High	gn (3, 4)	11/23 (47.8%)	23/55 (41.8%)	
Treatment outcome		14/26 (52.00)	45/74 (60,00%)	
CR Non-CR (PR, PD, SD)		14/26 (53.8%) 8/26 (30.8%)	45/74 (60.8%) 19/74 (25.7%)	0.822
Unavailable		8/26 (30.8%) 4/26 (15.4%)	19/74 (25.7%)	0.822
Pathology		4/20 (13.4%)	10/74 (13.340)	
Indolent		3/26 (11.5%)	18/74 (24.3%)	
Aggressive		23/26 (88.7%)	56/74 (75.7%)	0.169

*p-value <0.05=significant.

PS: Performance status, IPI: International Prognostic Index, CR: complete remission, PR: partial remission, PD: progressive disease, SD: stable disease, DLBCL: Diffuse large B-cell, IL: interleukin.

Supplementary Table 4. Comparison between B-cell non-Hodgkin lymphoma patients with wild genotype and those with polymorphic genotypes of interleukin-10-1082A/G regarding their hematological data.

Item		IL-10 wild genotype			IL-10 polyr			
		Range	Median	Mean ± SD	Range	Median	Mean ± SD	p-value
	Hb, g/dL	5-16.70	12	11.47±2.36	6.50-17	12	12±2.24	0.366
Hemogram	TLC, x10 ³ /cm ³	3.20-80	7.30	10.18±14.46	1.50-50	8	8.91±6.04	0.368
Ticmogram	Plts, x10 ³ /cm ³	23-675	294	287.96±160.88	14-644	278.50	294.19±124.04	0.771
LDH, IU/L		155-2922	350	539.42±582.06	130-3664	297.50	508.79±674.97	0.346
Hb: Hemoglobin.	TLC: total leukocyte	count. LDH: lactate de	hvdrogenase, Plts: r	olatelets. IL: interleukin. SD:	standard deviat	ion.		

Supplementary Table 5. Disease-free survival of B-cell non-Hodgkin lymphoma patients.								
Factors		Number of cases	Number of relapses	3 years	5 years	Median	p-value	
All		100	22	61.8	49.6	56.5		
A = 0	<60	71	16	68.5	56.2	130.6	0.318	
Age	≥60	29	6	57.1	28.6	56.5	0.318	
Cau	Male	54	12	61.6	35.2	130.6	0.000	
Sex	Female	46	10	67.2	67.2	47.4	0.989	
Ctomo	1 & 11	23	4	71.1	71.1	-	0.000	
Stage	III & IV	77	18	65.1	46.8	56.5	0.868	
IDI	Intermediate/High	71	17	63.3	49.7	56.5	0.903	
IPI	Low	29	5	76.7	38.4	47.4		
IPI for DLBCL	Interm. high/High	34	8	49.8	33.2	31.8	0.156	
	Low/Interm/low	44	6	77.0	57.7	-		
B-symp.	No	75	16	68.7	61.8	130.6	0.689	
	Yes	25	6	60.9	22.9	36.8		
DC	<2	61	11	78.2	57.9	230.0	0.153	
PS	≥2	39	11	48.7	36.5	31.8		
C 1	No	60	12	71.9	46.2	56.5	1	
Spleen	Yes	40	10	57.4	47.8	36.8	0.840	
Lt	No	62	12	70.7	49.5	56.5	0.541	
Liver	Yes	38	10	59.3	49.4	36.8	0.541	
Costum and all	<2	79	18	66.0	46.4	56.5	0.000	
Extranodal	≥2	21	4	64.9	64.9	-	0.932	
111.	Abnormal	57	15	56.1	48.1	36.8	0.170	
Hb	Normal	43	7	76.3	66.8	-	0.178	
TLC	<11,000	79	16	66.9	50.2	-	0.422	
TLC	≥11,000	21	6	84.4	36.8	36.8	0.433	
DI4-	≤150	17	3	85.7	64.3	130.6	0.222	
Plts	>150	83	19	61.9	45.8	56.5	0.333	

Supplementary Ta	ble 5. Continue.						
LDH	Elevated	61	16	64.6	60.0	130.6	0.010
	Normal	39	6	77.0	38.5	47.4	0.616
Pathology	Aggressive	79	15	64.6	48.4	56.5	0.000
	Indolent	21	7	68.0	56.7	230.8	0.889
IL-2	Π	20	2	69.2	69.2	-	0.211
IL-Z	TG & GG	80	20	65.4	45.3	56.5	0.211
IL-10	AA	26	4	79.5	39.8	-	0.205
	AG & GG	74	18	60.7	49.1	47.4	0.203
Hb: Hemoglobin, TLC: tot	al leukocyte count, LDH: lactate deh	vdrogenase. Plts: plate	lets. II : interleukin.	IPI: International P	rognostic Index.		•

Authorship Contributions

Surgical and Medical Practices: O.M.R.K.; Concept: H.A.R., O.M.R.K.; Design: M.M.K., H.A.A.R. O.M.R.K.; Data Collection or Processing: H.M.M.; Analysis or Interpretation: M.M.K., H.A.R.; Literature Search: M.M.K., H.M.M.; Writing: M.M.K., H.M.M., H.A.R., O.M.R.K.

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References

- Wróbel T, Mazur G, Dzietczenia J, Gebura K, Kuliczkowski K, Bogunia-Kubik K. VEGF and bFGF gene polymorphisms in patients with non-Hodgkin's lymphoma. Biomed Res Int 2013;2013:159813.
- Gerber SA, Sorensen EW, Sedlacek AL, Lim JY, Skrombolas D, Frelinger JG, Lord EM. Local expression of interleukin-2 by B16 melanoma cells results in decreased tumour growth and long-term tumour dormancy. Immunology 2013;138:280-292.
- D'Souza WN, Lefrançois L. IL-2 is not required for the initiation of CD8 T cell cycling but sustains expansion. J Immunol 2003;171:5727-5735.
- Song H, Chen L, Cha Z, Bai J. Interleukin 2 gene polymorphisms are associated with non-Hodgkin lymphoma. DNA Cell Biol 2012; 31:1279-1284.
- Howell WM, Rose-Zerilli MJ. Cytokine gene polymorphisms, cancer susceptibility, and prognosis. J Nutr 2007;137(1 Suppl):194–199.
- Lech-Maranda E, Bienvenu J, Michallet AS, Houot R, Robak T, Coiffier B, Salles G. Elevated IL-10 plasma levels correlate with poor prognosis in diffuse large B-cell lymphoma. Eur Cytokine Netw 2006;17:60-66.
- 7. Rothman N, Skibola CF, Wang SS, Morgan G, Lan Q, Smith MT, Spinelli JJ, Willett E, De Sanjose S, Cocco P, Berndt SI, Brennan P, Brooks-Wilson A, Wacholder S, Becker N, Hartge P, Zheng T, Roman E, Holly EA, Boffetta P, Armstrong B, Cozen W, Linet M, Bosch FX, Ennas MG, Holford TR, Gallagher RP, Rollinson S, Bracci PM, Cerhan JR, Whitby D, Moore PS, Leaderer B, Lai A, Spink C, Davis S, Bosch R, Scarpa A, Zhang Y, Severson RK, Yeager M, Chanock S, Nieters A. Genetic vvariation in TNF and IL10 and risk of non-Hodgkin lymphoma: a report from the InterLymph Consortium. Lancet Oncol 2006;7:27-38.
- Spink CF, Keen LJ, Mensah FK, Law GR, Bidwell JL, Morgan GJ. Association between non-Hodgkin lymphoma and haplotypes in the TNF region. Br J Haematol 2006;133:293–300.
- Mittal RD, Manchanda PK. Association of interleukin (IL)-4 intron-3 and IL-6 -174 G/C gene polymorphism with susceptibility to end-stage renal disease. Immunogenetics 2007;59:159-165.

- Cavet J, Middleton PG, Segall M, Noreen H, Davies SM, Dickinson AM. Recipient tumor necrosis factor-alpha and interleukin-10 gene polymorphisms associate with early mortality and acute graft-versus-host disease severity in HLA-matched sibling bone marrow transplants. Blood 1991;94:3941-3946.
- Breen EC, Boscardin WJ, Detels R, Jacobson LP, Smith MW, O'Brien SJ, Chmiel JS, Rinaldo CR, Lai S, Martínez-Maza O. Non-Hodgkin's B cell lymphoma in persons with acquired immunodeficiency syndrome is associated with increased serum levels of IL10, or the IL10 promoter -592 C/C genotype. Clin Immunol 2003;109:119-129.
- Oken MM, Creech RH, Tormey DC, Horton J, Davis TE, McFadden ET, Carbone PP. Toxicity and response criteria of the Eastern Cooperative Oncology Group. Am J Clin Oncol 1982;5:649–655.
- Matesanz F, Fedetz M, Leyva L, Delgado C, Fernández O, Alcina A. Effects of the multiple sclerosis associated -330 promoter polymorphism in IL2 allelic expression. J Neuroimmunol 2004;148:212-217.
- Wu MS, Huang SP, Chang YT, Shun CT, Chang MC, Lin MT, Wang HP, Lin JT. Tumor necrosis factor-alpha and interleukin-10 promoter polymorphisms in Epstein-Barr virus-associated gastric carcinoma. J Infect Dis 2002;185:106-110
- Rousset F, Garcia E, Defrance T, Péronne C, Vezzio N, Hsu DH, Kastelein R, Moore KW, Banchereau J. Interleukin 10 is a potent growth and differentiation factor for activated human B lymphocytes. Proc Natl Acad Sci U S A 1992;89:1890-1893.
- Blay JY, Burdin N, Rousset F, Lenoir G, Biron P, Philip T, Banchereau J, Favrot MC. Serum interleukin-10 in non-Hodgkin's lymphoma: a prognostic factor. Blood 1993;82:2169-2174.
- Fayad L, Keating MJ, Reuben JM, O'Brien S, Lee BN, Lerner S, Kurzrock R. Interleukin-6 and interleukin-10 levels in chronic lymphocytic leukemia: correlation with phenotypic characteristics and outcome. Blood 2001:97:256-263.
- Hulkkonen J, Pertovaara M, Antonen J, Lahdenpohja N, Pasternack A, Hurme M. Genetic association between interleukin-10 promoter region polymorphisms and primary Sjögren's syndrome. Arthritis Rheum 2001;44:176-179.
- Purdue MP, Lan Q, Kricker A, Grulich AE, Vajdic CM, Turner J, Whitby D, Chanock S, Rothman N, Armstrong BK. Polymorphisms in immune function genes and risk of non-Hodgkin lymphoma: findings from the New South Wales non-Hodgkin Lymphoma Study. Carcinogenesis 2007;28:704–712.
- Lan Q, Zheng T, Rothman N, Zhang Y, Wang SS, Shen M, Berndt SI, Zahm SH, Holford TR, Leaderer B, Yeager M, Welch R, Boyle P, Zhang B, Zou K, Zhu Y, Chanock S. Cytokine polymorphisms in the Th1/Th2 pathway and susceptibility to non-Hodgkin lymphoma. Blood 2006;107:4101–4108.
- Talaat RM, Abdel-Aziza AM, El-Maadawya EA, Abdel-Baryb N. Interleukin 10 gene promoter polymorphism and risk of diffuse large B cell lymphoma (DLBCL). Egyptian Journal of Medical Human Genetics 2014;15:7-13.
- 22. Kube D, Hua TD, von Bonin F, Schoof N, Zeynalova S, Klöss M, Gocht D, Potthoff B, Tzvetkov M, Brockmöller J, Löffler M, Pfreundschuh M, Trümper

- L. Effect of interleukin-10 gene polymorphisms on clinical outcome of patients with aggressive non-Hodgkin's lymphoma: an exploratory study. Clin Cancer Res 2008;14:3777-3784.
- 23. Berglund M, Thunberg U, Roos G, Rosenquist R, Enblad G. The interleukin-10 gene promoter polymorphism (-1082) does not correlate with clinical outcome in diffuse large B-cell lymphoma. Blood 2005;105:4894-4895.
- 24. Lech-Maranda E, Baseggio L, Bienvenu J, Charlot C, Berger F, Rigal D, Warzocha K, Coiffier B, Salles G. Interleukin-10 gene promoter polymorphisms influence the clinical outcome of diffuse large B-cell lymphoma. Blood 2004;103:3529-3534.
- Cunningham LM, Chapman C, Dunstan R, Bell MC, Joske DJ. Polymorphisms in the interleukin 10 gene promoter are associated with susceptibility to aggressive non-Hodgkin's lymphoma. Leuk Lymphoma 2003;44:251–255.