Effect of α-Tocopherol on Lipid Peroxidation Caused by Cisplatin in Rat Kidney

Zahit BOLAMAN*, Mehmet H. KÖSEOĞLU**, Süleyman DEMİR**, Gürhan KADIKÖYLÜ*, Sabri BARUTCA*, Habip ATALAY**, Diler ASLAN**

- * Division of Haematology/Oncology, Department of Internal Medicine, Adnan Menderes University Medical School, Aydın
- ** Department of Biochemistry, Pamukkale University Medical School, Denizli, TURKEY

ABSTRACT

Cisplatin (CDDP) is one of the most commonly used antineoplastic agents in current clinical practice. The major toxicities of CDDP are nonhaematological as nephrotoxicity and ototoxicity. Free oxygen radicals are known to play major role in CDDP-induced acute renal failure in rats. α -tocopherol is one of the well-known antioxidant agents. This study was designed to investigate the role of α -tocopherol pretreatment against CDDP-induced lipid peroxidation in rat kidney. Male Wistar rats were divided into three groups and treated as follows: control (saline intraperitoneally), CDDP (10 mg/kg, intraperitoneally), α -tocopherol (200 mg/kg, plus CDDP, intraperitoneally). Rats were sacrificed on third day of the treatment, and kidney tissues were obtained and analyzed. CDDP-treated rats showed high malondialdehyde (MDA) levels (p< 0.05). In the CDDP plus α -tocopherol group, renal MDA levels were not significantly different from the controls. These data suggest that α -tocopherol may be used to prevent CDDP-induced lipid peroxidation.

Key Words: Cisplatin, α-Tocopherol, Malondialdehyde.

ÖZET

α-Tokoferolün Sıçan Böbreğinde Sisplatinin Yol Açtığı Lipid Peroksidasyonuna Etkisi

Sisplatin (CDDP) günlük klinik pratikte en çok kullanılan anti-neoplastik ajanlardandır. CDDP'nin en önemli toksisiteleri, nefrotoksisite ve ototoksisitedir. Sıçanlarda CDDP'ye bağlı, akut renal yetmezlikte serbest oksijen radikallerinin önemli rol oynadığı bilinmektedir. α-tokoferol iyi bilinen antioksidan ajanlardandır. Bu çalışma α-tokoferolün sıçan böbreğinde CDDP'ye bağlı lipid peroksidasyonu önlemedeki rolünü araştırmak için yapılmıştır. Erkek Wistar sıçanları 3 gruba bölünmüştür. 1. kontrol (periton için salin), 2. CDDP (periton için

10 mg/kg), 3. α-tokoferol (200 mg/kg) + CDDP, periton içi. Tedavinin üçüncü günü sıçan böbrek dokuları incelenmiştir. CDDP verilen sıçanlarda malonildialdehid (MDA) düzeyleri yüksek bulunmuştur (p< 0.05). CDDP artı α-tokoferol alan sıçanlarda ise kontrol grubuna göre fark saptanmamıştır. Bu sonuçlar α-tokoferolün CDDP'ye bağlı lipid peroksidasyonu önlemede kullanılabileceğini göstermektedir.

Anahtar Kelimeler: Sisplatin, α-Tokoferol, Malonildialdehid.

Turk J Haematol 2003;20(1): 35-38

Received: 10.07.2002 **Accepted:** 04.11.2002

INTRODUCTION

Cisplatin (CDDP, cis-diamminedichloroplatinum) is a planar platinum complex consisting of two chloride leaving groups in the cis-position around platinum. CDDP has a broad spectrum of activity against many different solid tumours such as lung, ovary, testis, bladder and head and neck cancers and is also an effective agent in treatment of some haematological malignancies such as refractory lymphomas[1]. The adverse effects of CDDP include nausea and vomiting, nephro-, oto- and, neurotoxicities and myelosupression. The renal damage of CDDP is cumulative to glomeruli and tubules^[2,3]. The exact mechanism of CDDP-induced nephrotoxicity is insufficiently known. It was suggested that the toxic effect of CDDP might be related to free radicals that may cause oxidative damage in the kidney^[4,5].

Biochemical studies indicate that heavy metals can react with free sulphydryl (SH) groups. It was shown that after administration of CDDP, there was a significant decrease in SH groups per gram wet weight in rats^[6,7]. Additionally, CDDP can inhibit the activity of antioxidant enzymes and increase the lipid peroxidase content in renal tissue^[8]. Anti-oxidant therapies may be effective in prevention of the tissue damaging effects of various cytotoxic agents. The concentration of vitamin E in the renal tissue of adults rats was shown to decrease significantly after the administration of CDDP^[9]. It is known that, some antioxidants as vitamin C, α -tocopherol, amifostine and asetylcystein may prevent the damage induced by free oxygen radicals and reduce the incidence of cumulative nephrotoxicity

of CDDP. To our knowledge, there are few studies on the interaction of CDDP and vitamin E in the literature [10-14]. The objective of this study was to investigate the effect of α -tocopherol against CDDP-induced lipid peroxidation in rat kidney tissues.

MATERIAL and METHODS

CDDP (Cisplatinum-Onko) was obtained from TR Onko as 10 kg/10 mL flacons. Vitamin E (Evigen) was obtained from Aksu Farma Ltd. Şti. as 300 mg/2 mL ampules. Male Wistar rats, weighing from 190-220 g, were divided into three groups and treated as follows:

- 1. Controls (saline, 1 cc, intraperitoneally);
- 2. CDDP (10 mg/kg, intraperitoneally);
- 3. CDDP plus α -tocopherol (200 mg/kg, intraperitoneally).

Rats were fed on normal rhythm and standard breeding food and water according to the Institutional Animal Care and Use Committee and National Institute of Health Guidelines for animals care. All were sacrificed on third day of drug administration after 12 hours of fasting (overnight). The kidneys were obtained and washed with cold saline solution and were placed into Petri dishes containing ice-cold isolation medium (125 M KCl, 10 mM Tris, pH 7.4). They were stored at -20°C until analysed. Briefly the tissues were thawed and homogenised with Ultra-Turax in 0.15 M KCl solution. Each mL of the homogenate were mixed with 1.5 mL thiobarbituric acid (TBA), 1.5 mL acetic acid (pH: 3.5) and 0.2 mL sodium dodecyl sulphate. A set of MDA standards was also freshly prepared. After co-

upling, all samples and standards were heated at 100°C for one hour. The samples and standards were cooled on ice and MDA was extracted by adding 5 mL 1-butanol pyridine (15:1) to each sample. Each tube was centrifuged at 200 RPM for 10 minutes to separate the aqueous and organic phases. The absorbency of 1-butanol phase was measured at 532 nm (Shimahatsu VV 1601 Spectrophotometer) and compared with those obtained from MDA standards. The concentration values were calculated from absorption measurements as standard absorption^[15]. Specimen concentration was calculated as (specimen absorption/standard absorption) x standard concentration. TBA was purchased from Sigma Chemical Cooperation.

All results were expressed as the mean \pm the standard error of the mean (SEM). Statistical analyses were performed by Kruskall Wallis and Mann-Whitney U tests. p values of less than 0.05 were considered as significant.

RESULTS

MDA levels of all groups are shown in Figure 1. MDA levels were 2.61 ± 0.66 nmol/g wt in th control group. CDDP increased the MDA levels to 3.87 ± 0.70 nmol/g wt (p< 0.05). So tissue MDA levels were increased significantly in the CDDP group. In the CDDP plus α -tocopherol group, renal MDA levels were not different (2.26 \pm 0.70 nmol/g wt.) as compared to the control levels (p> 0.05).

DISCUSSION

CDDP is a widely used antineoplastic drug with

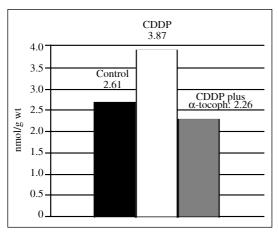


Figure 1. Renal tissue MDA levels.

prominent nephrotoxicity. The exact cellular mechanism of CDDP-induced nephrotoxicity has not been elucidated, but some preclinical studies in rats have demonstrated that lipid peroxidation was involved^[12,16]. Sugihara et al showed that the toxic effect of CDDP might be related to free oxygen radical induced damage^[13,16]. During incubation with ct-DNA in vitro CDDP have generated superoxide anions^[17]. An increment in levels of MDA, a lipid peroxidation product, has been observed in renal cortical slices following in vitro CDDP incubation^[18]. Additionally, the concentration of α-tocopherol may decrease in the renal tissue after administration of CDDP and, pretreatment with vitamin E is likely to be therapeutically useful^[9]. We observed that the tissue MDA levels were increased by CDDP. But in the α-tocopherol plus CDDP group MDA levels were not increased. In our study, renal tissue MDA levels were higher in the CDDP group than CDDP plus α-tocopherol group. The tissue MDA levels in the CDDP plus α-tocopherol group was similar to the control group (Figure 1).

Renal toxicity is one of the most common nonhaematological toxicities that limit the clinical use of CDDP treatments in a variety of cancers^[2]. Many agents have been used to protect the kidney tissues from CDDP-induced free radical damage^[4-7,11-14]. An important observation was that α-tocopherol-treated patients did not have the risk of CDDP toxicity in a former study^[19]. In our study; MDA levels were not increased in kidney tissues of the rats in the CDDP plus αtocopherol group compared to controls. According to our point of view, α-tocopherol may be protective against deleterious effects of CDDP in rat kidney tissues.

In conclusion, CDDP-induced renal toxicity was closely associated with increased lipid peroxidation. This study showed that, the increment of MDA, a lipid peroxidation product related with free oxygen radicals induced kidney damage, could be prevented by α -to-copherol treatment before CDDP. We suggest that the relationship in between CDDP nephrotoxicity and α -to-copherol should be further evaluated by well-designed preclinical and clinical studies.

REFERENCES

- Links M, Lewis C. Chemoprotectants: A review of their clinical pharmacology and therapeutic efficacy. Drugs 1999;57:293-308.
- Loehrer PJ, Einhorn LH. Drugs five years later. Cisplatin. Ann Intern Med 1984;100:704-13.
- Yokozawa T, Liu ZW. The role of ginsenoside-Rd in cisplatin-induced acute renal failure. Ren Fail 2000; 22:115-27.
- Matsushima H, Yonemura K, Ohishi K, Hishida A. The role of oxygen free radicals in cisplatin-induced acute renal failure in rats. J Lab Clin Med 1998;131: 518-26.
- Namikawa K, Hirai K, Tanaka I, et al. Effect of the lipid peroxide reaction caused by repeated cold stress on cisplatin-induced nephrotoxicity. Biol Trace Elem Res 1998;63:213-20.
- Levi J, Jacobs C, Kalman SM, et al. Mechanism of cisplatinum nephrotoxicity: I. Effects of sulfhydryl groups in rat kidneys. J Pharmacol Exp Their 1980; 213:545-50.
- el Daly ES. Protective effect of cysteine and vitamin E, Crocus stativus and Nigella sativa extracts on cisplatininduced toxicity in rats. J Pharm Belg 1998;53: 87-95.
- Sakthisekaran D, Sree S, Sriganth INP. Effect of cisplatin on renal free radical scavengers and lipid-peroxidation studies in rats. Med Sci Res 1993;21: 35-6.
- Appenroth D, Fröb S, Kersten L, et al. Protective effects of vitamin E and C on cisplatin nephrotoxicity in developing rats. Arch Toxicol 1997;71:677-83.
- Sun F, Hayami S, Haruna S, et al. In vivo antioxidative activity of propolis evaluated by the interaction with vitamins C and E and tle level of lipid hydroperoxides in rats.
 J Agric Food Chem 2000;48:1462-5.
- Appenroth D, Winnefeld K. Vitamin E and C in the prevention of metal nephrotoxicity in developing rats. Exp Toxicol Pathol 1998;50:391-6.
- Bolaman Z, Köseoğlu MH, Demir S, et al. Effect of amifostine on lipid peroxidation caused by cisplatin in rat kidney. J Chemother 2001;3:337-9.

- Sugihara K, Gemba M. Modification of cisplatin toxicity by antioxidants. Jpn J Pharmacol 1986;40: 353-5.
- Hannemann J, Duwe J, Baumann K. Iron and ascorbis acid-induced lipid peroxidation in renal microsomes isolated from rats treated with platinum compounds. Cancer Chemother Pharmacol 1991;28:427-33.
- Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 1979;95:351-8.
- Sugihara K, Nakano S, Gemba M. Effect of cisplatin on in vitro production of lipid peroxides in rat kidney cortex. Jpn J Pharmacol 1987;44:71-6.
- Masuda H, Tanaka T, Takahama U. Cisplatin generates superoxide anion by interaction with DNA in cell-free system. Biochem Biophys Res Commun 1994;203:1775-180.
- Nakano S, Gemba M. Potentiation of cisplatin-induced lipid peroxidation in kidney cortical slices by glutathione depletion. Jpn J Pharmacol 1989;50:87-92.
- Weijl NI, Hopman GD, Wipkink-Bakker A, et al. Cisplatin combination chemotherapy induces a fall plasma antioxidants of cancer patients. Ann Oncol 1998;9:1331-7.

Address for Correspondence:

Zahit BOLAMAN, MD

Division of Haematology/Oncology Department of Internal Medicine Adnan Menderes University Medical School Aydın, TURKEY