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# **Predictive Values of Early Suppression of Tumorigenicity 2 for** Acute GVHD and Transplant-related Complications after Allogeneic **Stem Cell Transplantation: Prospective Observational Study**

Allojenik Kök Hücre Transplantasyonu Sonrası Akut GVHD ve Transplantasyonla İlişkili Komplikasyonlar İçin Tümörjenisite 2'nin Erken Baskılanmasının Prediktif Değeri: Prospektif Gözlemsel Çalışma

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## Abstract

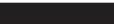
Objective: A soluble form of suppression of tumorigenicity 2 (sST2) has emerged as a biomarker for acute graft-versus-host disease (GVHD) and non-relapse mortality (NRM). We prospectively monitored sST2 levels during the early phase of hematopoietic stem cell transplantation (HSCT) and evaluated the clinical association with transplant-related complications including acute GVHD.

Materials and Methods: Thirty-two adult Japanese patients who received a first allogeneic HSCT were enrolled in this study. Levels of sST2 were measured at fixed time points (pre-conditioning, day 0, day 14, day 21, and day 28).

Results: The median age was 50.5 years (range=16-66). With a median follow-up of 21.5 months (range=0.9-35.4), 9 patients developed grade II-IV acute GVHD. Median sST2 levels on the day of HSCT were higher than baseline and reached the maximum value (92.7 ng/mL; range=0-419.7) on day 21 after HSCT. The optimal cutoff value of sST2 on day 14 for predicting grade II-IV acute GVHD was determined as 100 ng/mL by ROC analysis. The cumulative incidence of acute GVHD was 56.7% and 16.5% in the high- and low-sST2 groups, respectively (p<0.01). Multivariate analyses showed that high sST2 levels at day 14 were associated with a higher incidence of acute GVHD (hazard ratio=9.35, 95% confidence interval=2.92-30.0, p<0.01). The cumulative incidence of NRM was increased in the highsST2 group (33% vs 0%, p<0.01), but all the patients died of non-GVHD complications. Among 6 patients in the high-sST2 group without grade II-IV GVHD, 5 patients developed veno-occlusive disease (VOD) and one also had thrombotic microangiopathy (TMA).

Conclusion: The early assessment of sST2 after HSCT yielded predictive values for the onset of acute GVHD and other transplant-related complications including VOD and TMA.

Keywords: Suppression of tumorigenicity 2, Graft-versus-host disease, Biomarker, Hematopoietic stem cell transplantation



Amac: Akut graft-konakcı hastalığı (GVHD) ve relaps dısı mortalite (NRM) için bir biyobelirteç olarak, tümör oluşumu 2'nin baskılanmasının (sST2) cözünür bir formu ortaya konulmuştur. Hematopoietik kök hücre transplantasyonunun (HSCT) erken safhasında sST2 düzeylerini prospektif olarak izledik ve akut GVHD dahil transplantla ilişkili komplikasyonlarla klinik ilişkisini değerlendirdik.

Öz

Gereç ve Yöntemler: Bu çalışmaya ilk allojenik HSCT'si olan 32 yetişkin Japon hasta alındı. sST2 seviyeleri sabit zaman noktalarında (hazırlık rejimi öncesi, 0. gün, 14. gün, 21. gün ve 28. gün) ölçüldü.

Bulgular: Ortanca yaş 50,5 idi (aralık=16-66). Ortalama 21,5 aylık takip süresi (aralık= 0,9-35,4) ile 9 hastada Grade II-IV akut GVHD gelişti. Nakil gününde ortanca sST2 seviyeleri bazal değerlerden daha yüksekti ve HSCT'den sonraki 21. günde maksimum değere (92,7 ng/mL; aralık=0-419,7) ulaştı. Grade II-IV akut GVHD'yi tahmin etmek için 14. günde sST2'nin optimal eşik değeri, ROC analizi ile 100 ng/mL olarak belirlenmiştir. Akut GVHD'nin kümülatif insidansı yüksek ve düşük sST2 gruplarında sırasıyla %56,7 ve %16,5 idi (p<0,01). Çok değişkenli analizler 14. günde yüksek sST2 düzeylerinin daha yüksek akut GVHD insidansı ile ilişkili olduğunu göstermiştir (tehlike oranı=9, 35, %95 güven aralığı=2,92-30,0, p<0,01). NRM'nin kümülatif insidansı yüksek sST2 grubunda artmıştır (%33'e karşı %0, p<0,01), ancak tüm hastalar GVHD dışı komplikasyonlardan öldü. Yüksek sST2 grubunda grade II-IV GVHD olmayan 6 hastadan 5'inde veno-oklüzif hastalık (VOD) gelişti ve birinde trombotik mikroanjiyopati (TMA) vardı.

**Sonuç:** HSCT sonrası sST2'nin erken değerlendirmesi, akut GVHD'nin başlangıcı ve VOD ve TMA dahil transplantla ilişkili diğer komplikasyonlar için prediktif değerler vermiştir.

Anahtar Sözcükler: Tümör oluşumu 2'nin baskılanması, Akut graft-konakçı hastalığı, Biyomarker, Hematopoietik kök hücre transplantasyonu

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# Introduction

Acute graft-versus-host disease (GVHD) is one of the major complications after allogeneic hematopoietic stem cell transplantation (HSCT) and remains the leading cause of nonrelapse mortality (NRM) [1,2]. Identifying useful biomarkers for predicting onset or severity of acute GVHD at the early phase of HSCT may help in the development of a more individualized treatment strategy for GVHD. Recent studies show several plasma biomarkers that correlate with acute GVHD: suppression of tumorigenicity 2 (ST2) [3,4], interleukin (IL)-2 receptor  $\alpha$ [5], tumor necrosis factor receptor 1 (TNFR1) [5], hepatocyte growth factor [5], IL-8 [5], and IL-6 [6] for systemic GVHD; elafin for skin GVHD [7]; and regenerating islet-derived  $3-\alpha$ [8] and T-cell immunoglobulin mucin-3 for gastrointestinal GVHD [8,9,10,11,12]. Among these biomarkers, ST2 has emerged as a promising biomarker for onset or steroid-resistant acute GVHD and NRM [3,4,12,13,14]. In addition, recent studies have demonstrated that high ST2 levels are associated with the development of thrombotic microangiopathy (TMA) and venoocclusive disease (VOD) after HSCT [15,16].

ST2 is a membrane receptor expressed on several immune cell types that belong to the IL-1 receptor family. ST2 induces T helper type 2 (Th2) and Treg immune responses, which play important roles in GVHD [17]. A soluble form of ST2 (sST2) is a decoy receptor for IL-33, which blocks the IL-33/ST2 pathway and drives Th2 cells toward a Th1 cell phenotype, and it is thought to be important in the pathophysiology of GVHD [18,19]. In murine models, the ST2/IL-33 axis has been reported as a potential therapeutic target for GVHD [20,21].

Despite the accumulating evidence that higher sST2 levels are associated with increased GVHD risk and subsequent mortality, a clinical role of sST2 during the early phase of transplantation has not been fully elucidated. We performed a multicenter, prospective, observational study monitoring the serial changes in sST2 levels before and early after HSCT with the aim of determining the diagnostic and prognostic values for acute GVHD, other transplant-related complications, and mortality.

# **Materials and Methods**

# **Patients and Transplant Procedure**

Patients who received first allogeneic HSCT for hematological diseases were enrolled consecutively between February 2014 and July 2015 at Yokohama City University Hospital or Yokohama City University Medical Center. The selection of donor source and conditioning regimen was based on patients' hematological diagnosis, donor availability, and patients' clinical status.

Conditioning regimens were classified into myeloablative conditioning (MAC) and reduced-intensity conditioning (RIC) according to the definitions previously reported [22]. The

pre-transplant risk category was defined as standard or high according to the diagnosis and the disease stage at the time of transplantation, as described previously [23].

Written informed consent was obtained from all the patients enrolled in this study before the start of the trial. This study was approved by the institutional review boards of our university hospital and medical center and was conducted in accordance with the Declaration of Helsinki. All authors vouched for the accuracy and completeness of the reported data, analyses, and adherence to the study protocol.

# **Peripheral Blood Samples**

Samples were collected prospectively before conditioning, on the day of HSCT (day 0), and on days 14, 21, and 28 after HSCT. Serum sST2 and IL-33 levels were measured by ELISA (Human ST2/IL-1 R4 ELISA Kit and Human IL-33 ELISA Kit; R&D Systems, Minneapolis, MN, USA). Absorbance was measured using a microplate reader (Powerscan HT, DS Pharma Biomedical, Osaka, Japan).

# **Transplant-related Complications**

Acute GVHD was diagnosed clinically with histological confirmation when available. The classification of acute GVHD was based on the diagnostic criteria of the 1994 Consensus Conference on Acute GVHD Grading [24]. GVHD prophylaxis consisted of cyclosporine or tacrolimus with short-term methotrexate. Anti-thymocyte globulin was administered for HLA-serological mismatched transplant based on each institution's criteria.

Other transplant-related complications were diagnosed as follows: TMA was diagnosed according to the Blood and Marrow Clinical Trials Network and European Group for Blood and Marrow Transplantation guidelines [25,26]. VOD was diagnosed based on the Baltimore and Seattle criteria [27,28].

## Statistical Analysis

Fisher's exact test and the Mann-Whitney U test were used to assess the categorical and continuous variables, respectively. The receiver operating characteristics (ROC) curve from logistic regression models with the area under the curve (AUC) was used to present the correlation between sST2 and other biomarkers. The Pearson test was used to determine the correlation between sST2 and other biomarkers. The Kaplan-Meier method was used to assess overall survival (OS) using the log-rank test. For multivariate analysis, the Cox proportional hazards method was used to assess the OS. The Gray test and Fine-Gray test were used to assess the cumulative incidence of GVHD and NRM. The competing risks were GVHD and death prior to GVHD. Values of p<0.05 were considered statistically significant. All analyses were performed using EZR version 1.36

statistical software, which is a graphical user interface for R version 3.4.1 [29].

## Results

## **Patient Characteristics**

The clinical characteristics of 32 patients are summarized in Table 1. The transplant procedure was deemed heterogeneous based on the donor source, conditioning regimen, and GVHD prophylaxis.

# **Expression Patterns of sST2 Following Transplantation**

First, we evaluated the expression patterns of sST2 by serial sampling from individuals at a fixed time point before and early after HSCT. Different sST2 expression patterns were observed in each individual after the conditioning therapy (Figure 1A). Compared with median sST2 levels before conditioning (median=25.9 ng/mL; range=0-42.7 ng/mL), sST2 levels on day 0 were remarkably elevated in most of the patients (median=51.4 ng/mL; range=0-227.9 ng/mL). The sST2 levels reached the maximum value on day 21 after HSCT (median=92.7 ng/mL; range=0-419.7 ng/mL) (Figure 1B).

# Effects of Conditioning Damages and Inflammatory Conditions on sST2 Levels

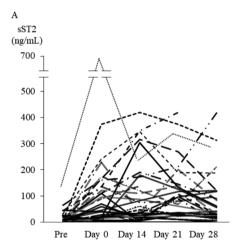
As sST2 expression levels may depend on conditioning intensity [4], the sST2 levels in patients who underwent MAC and RIC were compared. The sST2 levels in patients who underwent MAC were higher than those in patients who underwent RIC on days 0, 14, 21, and 28 without statistical difference between the two groups (Figure 2). To determine whether sST2 was affected by various inflammatory conditions after HSCT, the correlations between sST2 and representative inflammatory markers, serum ferritin and C-reactive protein (CRP), were estimated (Table 2). sST2 levels were strongly correlated with ferritin and CRP levels in all samples (r=0.456 and 0.615, respectively). Specifically, sST2 was well correlated with CRP at days 0, 14, and 21 (r=0.717, 0.630, and 0.628, respectively). Furthermore, levels of serum IL-33, the ligand of ST2, were under the detection limits in most of the patients, resulting in no correlation of IL-33 with sST2 levels (data not shown).

#### Association of sST2 Levels with Onset of Acute GVHD

The clinical courses of all patients are summarized in Figure 3. With a median follow-up of 21.5 months (range=0.9-35.4) after HSCT, 14 patients (43%) developed some grade of acute GVHD [median days to onset=39 days (range=9-84); median days to maximum grade=44 (range=15-94)]. The maximum grade of GVHD was grade I in five patients, grade II in three, and grade III in six; none developed grade IV. Nine patients had involvement of the gastrointestinal tract (stage 1 in two patients, stage 2 in two, stage 3 in four, and stage 4 in one), and ten patients had

skin involvement (stage 1 in one patient, stage 2 in five, and stage 3 in four). None developed liver GVHD.

When comparing the sST2 levels in patients with and without GVHD, median sST2 levels on day 14 after HSCT were relatively higher in patients with GVHD, but with no significant difference (Figure 4). Based on ROC curve analyses for predicting the onset of acute GVHD, the sST2 levels on day 14 showed the best AUC (0.66), with 66.7% sensitivity and 73.9% specificity (Figure S1, A). The optimal cut-off point of sST2 was 100 ng/mL based on the ROC analysis. We focused on day 14 sST2 levels as the earliest time point for predicting acute GVHD. There was no significant difference in patient characteristics between the high-sST2 group (>100 ng/mL) and the low-sST2 group (≤100 ng/mL) (Table 1). The cumulative incidence of grades II-IV GVHD was significantly higher in the high-sST2 group (56.7%) than that in the low-sST2 group (16.5%) (p<0.01) (Figure 5A).



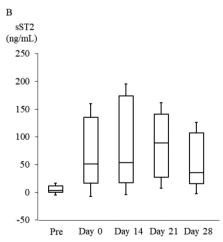


Figure 1. Expression patterns of sST2 following transplantation. A) Expression patterns of sST2 before conditioning (Pre) and on days 0, 14, 21, and 28 after transplantation in individual patients. B) Expression patterns of median sST2 values before conditioning (Pre) and on days 0, 14, 21, and 28 after transplantation. Box plots indicate medians, interquartiles, and ranges of sST2 levels.

sST2: Suppression of tumorigenicity 2.

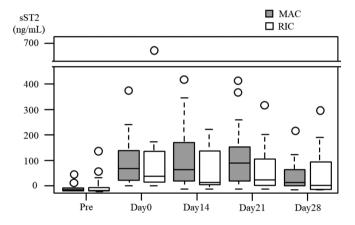
Characteristics	Total, n	sST2 low, n	sST2 high, n	р
Age, median (range), years	50.5 (16-66)	49 (16-64)	51.5 (26-66)	0.60
Sex, male/female	20/12	11/9	9/3	0.45
Performance status	20/12	11/3	3/3	0.59
0	16	11	5	0.33
<u>0</u> 1	13	8	5	
2	2	1	1	
Z Unknown	1	0	1	
Diagnosis	1	0	I	0.39
Acute myeloid leukemia	18	13	5	0.33
Acute lymphoid leukemia	4	2	2	
Myelodysplastic syndrome	4	1	3	
Malignant lymphoma	4	2	2	
Multiple myeloma				
Aplastic anemia	1	1	0	
Disease risk <sup>†</sup>	1	1	0	0.70
Disease risk <sup>1</sup> Standard	22	15	0	0.70
	23	15	8	
High	9	5	4	0.22
Donor type Unrelated donor	24	14	10	0.33
	24	14	10	
Related donor	8	6	2	0.47
Donor source	24	10	0	0.47
Bone marrow	24	16	8	
Peripheral blood	4	3	1	
Cord blood	4	1	3	0.70
HLA <sup>‡</sup>	0.1	1.4	7	0.70
Matched	21	14	7	
Mismatched	11	6	5	
Conditioning				1
MAC	21	13	8	
RIC	11	7	4	0.10
HCT-CI	1.0			0.42
0	18	11	7	
1-2	8	5	3	
≥3	6	4	2	
ATG used				0.34
Yes	7	3	4	
No	25	17	8	
GVHD prophylaxis				0.37
FK506 + sMTX	26	15	11	
CyA + sMTX	6	5	1	

sST2: Suppression of tumorigenicity, HLA: Human leukocyte antigen, MAC: Myeloablative conditioning, RIC: Reduced-intensity conditioning, HCT-CI: Hematopoietic cell transplantation comorbidity index, ATG: Anti-thymocyte globulin, GVHD: Graft-versus-host disease, FK506: Tacrolimus, sMTX: Short-term methotrexate, CyA: Cyclosporine. †Disease risk: "standard risk" indicates complete remission, partial response, untreated MDS, and aplastic anemia. "High risk" indicates all others [23].

\*HLA serological match or mismatch (6-loci).

Multivariate analyses showed that high sST2 was associated with higher incidence of acute GVHD (hazard ratio=9.35, 95% confidence interval=2.92-30.0, p<0.01) (Table 3). sST2 levels at other time points of days 0, 21, and 28 were not associated with acute GVHD (data not shown). Although post-transplant CRP and ferritin levels were well correlated with sST2 (Table 2), these markers did not influence the cumulative incidence of acute GVHD (cut-off values were calculated by ROC analysis=CRP 6.0 mg/dL and ferritin 1700 mg/dL, respectively) (Figure 5B and Figure S1, B). These data suggest that the day 14 sST2 levels are supportive findings for predicting the onset of acute GVHD.

Among the nine patients who had grade II-IV GVHD, six patients were part of the high-sST2 group (Figure 6). The patients in the



**Figure 2.** Comparison of sST2 expression patterns based on conditioning intensity. Median sST2 levels in patients who received MAC and RIC following transplantation. Box plots indicate medians, interquartiles, and ranges of sST2 levels.

RIC: Reduced-intensity conditioning, MAC: Myeloablative conditioning, sST2: Suppression of tumorigenicity 2.

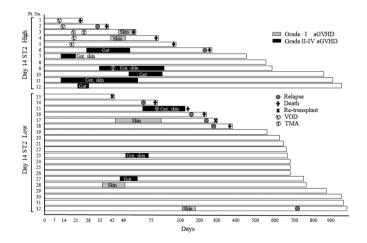


Figure 3. Clinical courses following HSCT in individual patients. aGVHD: Acute graft-versus-host disease, Pt. No.: Patient number, VOD: Veno-occlusive disease, TMA: Thrombotic microangiopathy, ST2: Suppression of tumorigenicity 2, HSCT: Hematopoietic stem cell transplantation.

high-sST2 group developed acute GVHD relatively early after HSCT (median onset of GVHD=22 days; range=9-50), and sST2 levels were elevated several days before the onset of acute GVHD. Four patients (cases 6, 7, 11, and 12) developed acute GVHD before day 28, and the sST2 levels were the highest just before the development of GVHD. On the other hand, low-sST2 patients did not develop GVHD in the early phase of HSCT (median onset of GVHD=48 days; range=42-55).

# Association of sST2 Levels with GVHD Severity and Target Organ Involvement

The association of day 14 sST2 levels with GVHD severity was examined, but the sST2 levels were not associated with the grade of GVHD (data not shown). The cumulative incidence of gastrointestinal GVHD was significantly increased in the high-sST2 group (50% vs. 15%, p=0.03). However, there was no significant association between sST2 levels and skin GVHD.

# Association of sST2 Levels with Other Transplant-related Complications

Finally, we evaluated the association of sST2 levels with other transplant-related complications and mortality (Figure 3). The 1-year NRM and OS after HSCT was 12.5% and 65.6%, respectively. The cumulative incidence of 1-year NRM was significantly increased in the high-sST2 group on day 14 (33% vs. 0%, p<0.01), while there was no significant difference in 1-year OS by univariate analysis (high sST2=50.0% vs. low sST2=84.2%; p=0.11). Severe non-GVHD complications were observed in 10 patients, including TMA (n=2), VOD (n=5), graft failure (n=1), engraftment syndrome (n=4), sepsis (n=3), and

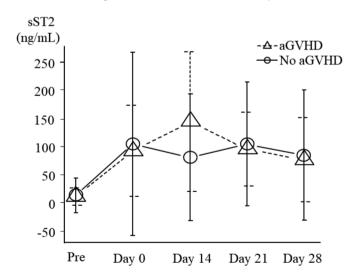


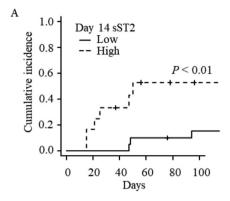
Figure 4. Comparison of sST2 expression patterns in patients with and without acute GVHD. Mean sST2 levels in patients who developed grade II-IV acute GVHD or not following transplantation. Bars indicate the mean  $\pm$  SEM of sST2 levels.

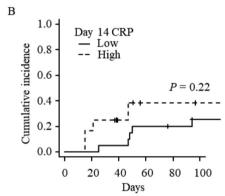
aGVHD: Acute graft-versus-host disease, sST2: Suppression of tumorigenicity 2, GVHD: Graft-versus-host disease.

Table 2.	Correlation between s	uppression of tur	norigenicity 2 and	other inflammato	ory markers.		
		Pre	Day 0	Day 14	Day 21	Day 28	Total
sST2	Median, ng/mL (range)	25.9 (0-42.7)	51.4 (0-227.9)	53.7 (0-419.7)	92.7 (0-419.7)	35.9 (0-419.3)	
CRP	Median, mg/dL (range)	0.16 (0-10.5)	1.5 (0-11.8)	4.8 (0.1-24.7)	1.7 (0.1-25.7)	0.6 (0-16.5)	
	r	0.303	0.717	0.630	0.628	0.476	0.615
	р	0.092	<0.001	<0.001	<0.001	0.011	<0.001
Ferritin	Median, ng/dL (range)	986 (100-2675)	1384 (257-7287)	1943 (432-5231)	2733 (859-9086)	2652 (896-9795)	
	r	0.463	0.304	0.322	0.467	0.284	0.456
	р	0.009	0.116	0.078	0.009	0.135	<0.001
sST2: Suppr	ression of tumorigenicity 2, CRP	: C-reactive protein.					

	Univariate	ultivariate analyses for acute graft-versus-ho		Multivariate	
Variable	n	Day 100 aGVHD	р	Hazard ratio (95% CI)	р
	"	% (95% CI)			
Age	'	<u>'</u>		'	
<55 years	23	22.8 (7.9-42.9)		Reference	
≥55 years	9	49.2 (11.6-79.2)	0.29	3.48 (0.50-24.5)	0.21
Performance status				-	,
0	16	31.2 (0.1-0.5)			
1	13	27.0 (0.1-0.6)			
2	2	50.0 (0.0-1.0)			
Unknown	1	NA	0.85		
Disease risk				,	,
Standard	23	30.4 (0.1-0.5)			
High	9	30.6 (0.0-0.7)	0.71		
HLA	,	,			'
Matched	21	31.4 (12.2-52.9)			
Mismatched	11	30.1 (5.8-60.3)	0.98		
Conditioning					
MAC	21	26.0 (8.8-47.3)		Reference	
RIC	11	39.4 (10.4-68.1)	0.53	1.94 (0.19-19.7)	0.57
нст-сі					•
0	18	29.8 (0.1-0.5)			
1-2	8	37.5 (0.1-0.7)			
≥3	6	16.7 (0.0-0.6)	0.79		
Donor source					•
Bone marrow	24	30.3 (13-49.6)		Reference	
Peripheral blood	4	25.0 (0.3-71.4)		0.37 (0.05-2.74)	0.33
Cord blood	4	NA	0.98	0.13 (0.02-0.81)	0.03
sST2, day 14					
Low (≤100 ng/mL)	20	16.5 (3.8-37.1)		Reference	
High (>100 ng/mL)	12	56.7 (19.2-82.2)	<0.01	9.35 (2.92-30.0)	<0.01

pneumonia (n=1), which often overlapped. Four patients in the high-sST2 group died of transplant-related complications including sepsis, graft failure, TMA, and pneumonia (cases 1, 3, 4, and 5) without developing grade II-IV GVHD. The two patients





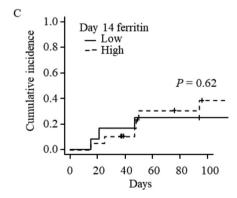


Figure 5. High sST2 on day 14 is correlated with subsequent GVHD development. A) Cumulative incidence of grade II-IV acute GVHD by day 100 in patients with high sST2 (>100 ng/mL) and low sST2 ( $\leq$ 100 ng/mL) levels on day 14 after transplantation. B) Cumulative incidence of grade II-IV acute GVHD by day 100 in patients with high-CRP (>6.0 mg/dL) and low-CRP ( $\leq$ 6.0 mg/dL) levels on day 14 after transplantation. C) Cumulative incidence of grade II-IV acute GVHD by day 100 in patients with high ferritin (>1700 mg/dL) and low ferritin ( $\leq$ 1700 mg/dL) levels on day 14 after transplantation. The CRP and ferritin cut-off values were calculated by ROC curve analysis for predicting the onset of acute GVHD (ROC curves are available in Figure S1).

sST2: Suppression of tumorigenicity 2, CRP: C-reactive protein, GVHD: Graft-versus-host disease.

who had TMA (cases 3 and 9) and five who had VOD (cases 1–5) were included in the high-ST2 group. All patients who died had severe overlapping complications including VOD.

# **Discussion**

Previous studies have shown that ST2 may predict the onset of acute GVHD, but no detailed analysis regarding the serial monitoring of sST2 following HSCT has been reported thus far. In this study, we performed a prospective evaluation of sST2 expression patterns by serial monitoring before and early after HSCT and identified that early assessment of sST2 after HSCT can yield predictive values for the onset of acute GVHD in addition to other severe transplant-related complications, such as TMA and VOD.

Although high sST2 levels on day 28 are associated with an increased risk of acute GVHD in cord blood transplantation [2], the optimal timing to measure sST2 remains unclear. We conducted serial sST2 measurements at fixed time points during HSCT and found that high sST2 levels around day 14 had the most significant association for predicting GVHD development. The expression patterns of sST2 in individual patients showed that sST2 is relatively higher during exacerbation of acute GVHD. As shown in Figure 6, the sST2 levels in patients with early-phase GVHD were higher than those with late-phase GVHD, suggesting that sST2 is clinically useful in predicting the early phase of GVHD. As acute GVHD often develops in the early

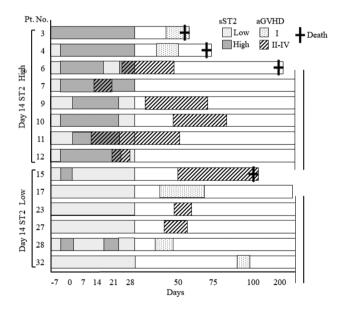


Figure 6. Association of early-phase sST2 levels with subsequent development of acute GVHD. Patient numbers correspond with those in Figure 3. Clinical course of 14 patients who developed grade I-IV acute GVHD. Dotted bars (grade I GVHD) or striped bars (grade II-IV GVHD) indicate the duration of acute GVHD clinical symptoms. Cut-off point of sST2 is 100 ng/mL.

aGVHD: Acute graft-versus-host disease, Pt. No.: Patient number, GVHD: Graft-versus-host disease, sST2: Suppression of tumorigenicity 2.

phase of HSCT, it is advantageous to establish biomarkers that can predict the onset of GVHD at earlier time points.

A soluble form of ST2 is released from endothelial cells, epithelial cells, and fibroblasts in response to inflammatory stimuli [30,31]. A previous study has shown that sST2 levels are associated with conditioning intensity; sST2 levels were two to four times higher after the MAC regimen than the RIC regimen [4]. The present study showed that sST2 levels after MAC were relatively higher than those after RIC. To reduce the effect of conditioning intensity on sST2 levels, we also examined the predictive values of the day 14/day 0 sST2 ratio for acute GVHD, but we did not detect any relationship with the development of GVHD (data not shown). Furthermore, we examined the association between sST2 and targets of GVHD involvement (skin, gastrointestinal tract, and liver) and found that most of the patients in the high-sST2 group developed gastrointestinal GVHD, which is thought to occur due to endothelial damage. These findings imply that the release of sST2 during GVHD exacerbation partly depends on the degree of endothelial injury occurring after conditioning therapy.

On the other hand, examining the association of sST2 with other complications and mortality is also important. Vander Lugt et al. [4] showed that high sST2 levels were associated with NRM within 6 months after HSCT. Moreover, other studies showed that high sST2 levels had a similar association with increased risk of NRM [3,12,13]. In this study, although it was difficult to evaluate this issue since only four patients developed NRM, they all showed high sST2 levels on day 14. Notably, they died of sepsis, graft failure, TMA, and pneumonia, without any GVHD, and all of them developed VOD. Recent studies demonstrated that high sST2 levels were significantly associated with TMA [15] and VOD [16], both of which are characterized by endothelial cell injury. In accordance with these observations, two patients who developed TMA and five who developed VOD showed high sST2 levels on day 14, even in our small cohort. In addition, the correlation of sST2 with CRP and ferritin suggests that sST2 is released under a variety of inflammatory conditions. Several studies have shown that pro-inflammatory cytokines and their receptors are potential GVHD biomarkers, but some factors other than GVHD contribute to these cytokines' release (e.g., TNF- $\alpha$ , TNFR1, IL-6) [5,6,32]. Therefore, caution should be taken when diagnosing patients who have high sST2 levels as sST2 is not a specific biomarker for acute GVHD.

## **Study Limitations**

The limitations of this study include the small number of patients and the heterogeneous patient populations and transplant procedures, such as conditioning regimen, donor source, and disease status. Even though we conducted detailed analyses of

sST2 expression patterns in individual patients, it was difficult to exclude the multiple factors that can cause an increase in sST2 (e.g., cardiac overload or infection).

# Conclusion

We revealed that sST2 levels increased not only in patients with acute GVHD but also in those with other life-threatening complications, such as TMA, VOD, and severe infection, as identified during patient monitoring. Although these complications often overlap with each other in the clinical settings for HSCT, gastrointestinal GVHD, TMA, and VOD, all of which are linked to endothelial injury, may be key complications related to high sST2 release. The early assessment of sST2 after HSCT may be a predictive indicator for acute GVHD and other transplant-related complications. Further studies with larger sample sizes and serial monitoring are needed to clarify the clinical value of sST2.

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## **Ethics**

**Ethics Committee Approval:** This study was approved by the Institutional Review Board of Yokohama City University.

Informed Consent: It was obtained.

## **Authorship Contributions**

Study Design: T.M., T.T.; Data Collection: A.M., T.M., T.T., T.A., M.K., S.K., Y.I., H.T., Y.N., A.N., W.Y., K. Mo., M.H., K. Ma., S.F.; Analysis: A.M.; Writing: A.M., T.M.; Approval: H.N.

**Conflict of Interest:** The authors declare no conflicts of interest.

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## References

- Morishima Y, Kawase T, Malkki M, Morishima S, Spellman S, Kashiwase K, Kato S, Cesbron A, Tiercy JM, Senitzer D, Velardi A, Petersdorf EW; International Histocompatibility Working Group in Hematopoietic Cell Transplantation. Significance of ethnicity in the risk of acute graft-versushost disease and leukemia relapse after unrelated donor hematopoietic stem cell transplantation. Biol Blood Marrow Transplant 2013;19:1197-1203.
- Ferrara JL, Levine JE, Reddy P, Holler E. Graft-versus-host disease. Lancet 2009;373:1550-1561.
- Ponce DM, Hilden P, Mumaw C, Devlin SM, Lubin M, Giralt S, Goldberg JD, Hanash A, Hsu K, Jenq R, Perales MA, Sauter C, van den Brink MR, Young JW, Brentjens R, Kernan NA, Prockop SE, O'Reilly RJ, Scaradavou A, Paczesny S, Barker JN. High day 28 ST2 levels predict for acute graft-versus-host disease and transplant-related mortality after cord blood transplantation. Blood 2015;125:199-205.

- Vander Lugt MT, Braun TM, Hanash S, Ritz J, Ho VT, Antin JH, Zhang Q, Wong CH, Wang H, Chin A, Gomez A, Harris AC, Levine JE, Choi SW, Couriel D, Reddy P, Ferrara JL, Paczesny S. ST2 as a marker for risk of therapy-resistant graft-versus-host disease and death. N Engl J Med 2013;369:529-539.
- Paczesny S, Krijanovski OI, Braun TM, Choi SW, Clouthier SG, Kuick R, Misek DE, Cooke KR, Kitko CL, Weyand A, Bickley D, Jones D, Whitfield J, Reddy P, Levine JE, Hanash SM, Ferrara JL. A biomarker panel for acute graft-versushost disease. Blood 2009;113:273–278.
- Kennedy GA, Varelias A, Vuckovic S, Le Texier L, Gartlan KH, Zhang P, Thomas G, Anderson L, Boyle G, Cloonan N, Leach J, Sturgeon E, Avery J, Olver SD, Lor M, Misra AK, Hutchins C, Morton AJ, Durrant ST, Subramoniapillai E, Butler JP, Curley Cl, MacDonald KP, Tey SK, Hill GR. Addition of interleukin-6 inhibition with tocilizumab to standard graft-versus-host disease prophylaxis after allogeneic stem-cell transplantation: a phase 1/2 trial. Lancet Oncol 2014;15:1451-1459.
- Paczesny S, Braun TM, Levine JE, Hogan J, Crawford J, Coffing B, Olsen S, Choi SW, Wang H, Faca V, Pitteri S, Zhang Q, Chin A, Kitko C, Mineishi S, Yanik G, Peres E, Hanauer D, Wang Y, Reddy P, Hanash S, Ferrara JL. Elafin is a biomarker of graft-versus-host disease of the skin. Sci Transl Med 2010;2:13ra12.
- Ferrara JL, Harris AC, Greenson JK, Braun TM, Holler E, Teshima T, Levine JE, Choi SW, Huber E, Landfried K, Akashi K, Vander Lugt M, Reddy P, Chin A, Zhang Q, Hanash S, Paczesny S. Regenerating islet-derived 3-alpha is a biomarker of gastrointestinal graft-versus-host disease. Blood 2011;118:6702-6708.
- Harris AC, Ferrara JL, Braun TM, Holler E, Teshima T, Levine JE, Choi SW, Landfried K, Akashi K, Vander Lugt M, Couriel DR, Reddy P, Paczesny S. Plasma biomarkers of lower gastrointestinal and liver acute GVHD. Blood 2012;119:2960–2963.
- Levine JE, Logan BR, Wu J, Alousi AM, Bolanos-Meade J, Ferrara JL, Ho VT, Weisdorf DJ, Paczesny S. Acute graft-versus-host disease biomarkers measured during therapy can predict treatment outcomes: a Blood and Marrow Transplant Clinical Trials Network study. Blood 2012;119:3854-3860.
- Hansen JA, Hanash SM, Tabellini L, Baik C, Lawler RL, Grogan BM, Storer B, Chin A, Johnson M, Wong CH, Zhang Q, Martin PJ, McDonald GB. A novel soluble form of Tim-3 associated with severe graft-versus-host disease. Biol Blood Marrow Transplant 2013;19:1323-1330.
- McDonald GB, Tabellini L, Storer BE, Lawler RL, Martin PJ, Hansen JA. Plasma biomarkers of acute GVHD and nonrelapse mortality: predictive value of measurements before GVHD onset and treatment. Blood 2015;126:113-120.
- Abu Zaid M, Wu J, Wu C, Logan BR, Yu J, Cutler C, Antin JH, Paczesny S, Choi SW. Plasma biomarkers of risk for death in a multicenter phase 3 trial with uniform transplant characteristics post-allogeneic HCT. Blood 2017;129:162-170.
- Yu J, Storer BE, Kushekhar K, Abu Zaid M, Zhang Q, Gafken PR, Ogata Y, Martin PJ, Flowers ME, Hansen JA, Arora M, Cutler C, Jagasia M, Pidala J, Hamilton BK, Chen GL, Pusic I, Lee SJ, Paczesny S. Biomarker panel for chronic graft-versus-host disease. J Clin Oncol 2016;34:2583–2590.
- Rotz SJ, Dandoy CE, Davies SM. ST2 and endothelial injury as a link between GVHD and microangiopathy. N Engl J Med 2017;376:1189-1190.
- Akil A, Zhang Q, Mumaw CL, Raiker N, Yu J, Velez de Mendizabal N, Haneline LS, Robertson KA, Skiles J, Diaz-Ricart M, Carreras E, Renbarger J, Hanash S, Bies RR, Paczesny S. Biomarkers for diagnosis and prognosis of sinusoidal obstruction syndrome after hematopoietic cell transplantation. Biol Blood Marrow Transplant 2015;21:1739-1745.
- Griesenauer B, Paczesny S. The ST2/IL-33 axis in immune cells during inflammatory diseases. Front Immunol 2017;8:475.
- Schmitz J, Owyang A, Oldham E, Song Y, Murphy E, McClanahan TK, Zurawski G, Moshrefi M, Qin J, Li X, Gorman DM, Bazan JF, Kastelein RA. IL-

- 33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. Immunity 2005;23:479-490.
- 19. Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. Immunity 2013;39:1003-1018.
- Reichenbach DK, Schwarze V, Matta BM, Tkachev V, Lieberknecht E, Liu Q, Koehn BH, Pfeifer D, Taylor PA, Prinz G, Dierbach H, Stickel N, Beck Y, Warncke M, Junt T, Schmitt-Graeff A, Nakae S, Follo M, Wertheimer T, Schwab L, Devlin J, Watkins SC, Duyster J, Ferrara JL, Turnquist HR, Zeiser R, Blazar BR. The IL-33/ST2 axis augments effector T-cell responses during acute GVHD. Blood 2015;125:3183-3192.
- 21. Matta BM, Reichenbach DK, Zhang X, Mathews L, Koehn BH, Dwyer GK, Lott JM, Uhl FM, Pfeifer D, Feser CJ, Smith MJ, Liu Q, Zeiser R, Blazar BR, Turnquist HR. Peri-alloHCT IL-33 administration expands recipient T-regulatory cells that protect mice against acute GVHD. Blood 2016;128:427-439.
- Bacigalupo A, Ballen K, Rizzo D, Giralt S, Lazarus H, Ho V, Apperley J, Slavin S, Pasquini M, Sandmaier BM, Barrett J, Blaise D, Lowski R, Horowitz M. Defining the intensity of conditioning regimens: working definitions. Biol Blood Marrow Transplant 2009;15:1628-1633.
- 23. Marks R, Potthoff K, Hahn J, Ihorst G, Bertz H, Spyridonidis A, Holler E, Finke JM. Reduced-toxicity conditioning with fludarabine, BCNU, and melphalan in allogeneic hematopoietic cell transplantation: particular activity against advanced hematologic malignancies. Blood 2008;112:415-425.
- Przepiorka D, Weisdorf D, Martin P, Klingemann HG, Beatty P, Hows J, Thomas ED. 1994 Consensus Conference on Acute GVHD Grading. Bone Marrow Transplant 1995;15:825–828.
- Ruutu T, Barosi G, Benjamin RJ, Clark RE, George JN, Gratwohl A, Holler E, lacobelli M, Kentouche K, Lammle B, Moake JL, Richardson P, Socie G, Zeigler Z, Niederwieser D, Barbui T, European Group for B, Marrow T, European L. Diagnostic criteria for hematopoietic stem cell transplant-associated microangiopathy: results of a consensus process by an International Working Group. Haematologica 2007;92:95-100.
- Ho VT, Cutler C, Carter S, Martin P, Adams R, Horowitz M, Ferrara J, Soiffer R, Giralt S. Blood and marrow transplant clinical trials network toxicity committee consensus summary: thrombotic microangiopathy after hematopoietic stem cell transplantation. Biol Blood Marrow Transplant 2005;11:571-575.
- McDonald GB, Hinds MS, Fisher LD, Schoch HG, Wolford JL, Banaji M, Hardin BJ, Shulman HM, Clift RA. Veno-occlusive disease of the liver and multiorgan failure after bone marrow transplantation: a cohort study of 355 patients. Ann Intern Med 1993;118:255-267.
- Jones RJ, Lee KS, Beschorner WE, Vogel VG, Grochow LB, Braine HG, Vogelsang GB, Sensenbrenner LL, Santos GW, Saral R. Venoocclusive disease of the liver following bone marrow transplantation. Transplantation 1987;44:778–783.
- Kanda Y. Investigation of the freely available easy-to-use software 'EZR' for medical statistics. Bone Marrow Transplant 2013;48:452-458.
- Kumar S, Tzimas MN, Griswold DE, Young PR. Expression of ST2, an interleukin-1 receptor homologue, is induced by proinflammatory stimuli. Biochem Biophys Res Commun 1997;235:474-478.
- Rossler U, Thomassen E, Hultner L, Baier S, Danescu J, Werenskiold AK. Secreted and membrane-bound isoforms of T1, an orphan receptor related to IL-1-binding proteins, are differently expressed in vivo. Dev Biol 1995;168:86-97.
- Schots R, Kaufman L, Van Riet I, Ben Othman T, De Waele M, Van Camp B, Demanet C. Proinflammatory cytokines and their role in the development of major transplant-related complications in the early phase after allogeneic bone marrow transplantation. Leukemia 2003;17:1150-1156.

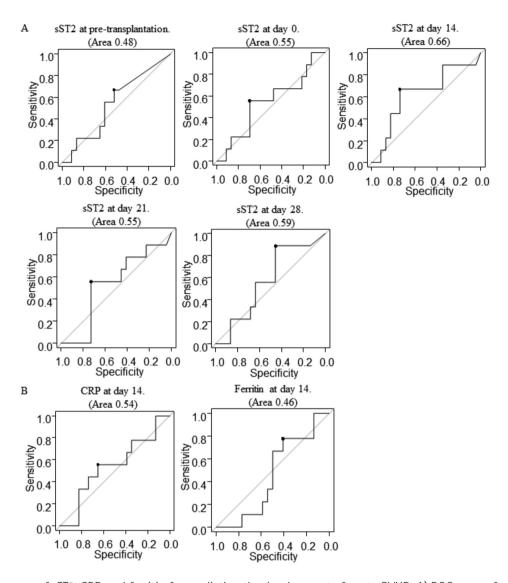


Figure S1. ROC curves of sST2, CRP, and ferritin for predicting the development of acute GVHD. A) ROC curves for sST2 with area under the curve (AUC) before conditioning and on days 0, 14, 21, and 28 after transplantation. B) ROC curves for CRP and ferritin with AUC on day 14.

sST2: Suppression of tumorigenicity 2, CRP: C-reactive protein, GVHD: Graft-versus-host disease, AUC: Area under the curve, ROC: Receiver operating characteristics.