Turk J Hematol 2024;41:128-129

Novel Four-Way t(8;14;15;21)(q22;q22;q15;q22.1) Translocation Variant in Acute Myeloid Leukemia with *RUNX1::RUNX1T1*

RUNX1::RUNX1T1 İçeren Akut Myeloid Lösemide Yeni Dört-Yol t(8;14;15;21) (q22;q22;q15;q22.1) Varyant Translokasyonu

🗈 Noriko Tsuge, 🖻 Fumiya Ogasawara, 🕲 Takumi Kondo, 🕩 Shohei Yoshida, 🕲 Kensuke Kojima

Kochi Medical School, Department of Hematology, Nankoku, Japan

To the Editor,

Acute myeloid leukemia (AML) with t(8;21)(q22;q22.1)/ *RUNX1::RUNX1T1* is a distinct AML entity clinically characterized by extramedullary involvement and favorable prognosis with conventional chemotherapy. Variant t(8;21) translocations involving four chromosomes have been rarely described, and some researchers have suggested that the four-way t(8;21) translocation may predict poor prognosis [1,2,3,4,5,6]. Here, we report a case of AML with t(8;14;15;21)(q22;q22;q15;q22.1)/ *RUNX1::RUNX1T1*.

A 71-year-old woman presented with a 2-week history of shoulder pain. A computed tomography scan showed a posterior mediastinal mass (long-axis diameter: 9 cm) extending to the spinal canal. The hemoglobin level was 11.9 g/dL; white blood cell count was 11.2x10⁹/L with 53.5% blasts, 0.5% myelocytes, 0.0% metamyelocytes, 22.5% neutrophils, 0.5% basophils, 2.5% monocytes, and 20.5% lymphocytes; and platelet count was 187x10⁹/L. A bone marrow aspirate smear showed hypercellular

marrow with 72% myeloperoxidase-positive blasts with Auer rods. A multiplex quantitative real-time polymerase chain reaction panel revealed a chimeric *RUNX1::RUNX1T1* transcript. Leukemia cells expressed CD34, CD33, CD13, CD19, CD56, and HLA-DR and were characterized as 45. X. -X. and t(8:14:15:21) (g22;g22;g15;g22.1) in all 20 metaphases analyzed. Spectral karyotyping with interphase fluorescence in situ hybridization revealed that distal regions of the 8q22, 14q22, 15q15, and 21g22.1 chromosomes were transferred in a cycle, resulting in RUNX1::RUNX1T1 fusion (Figure 1). KIT, NPM1, and FLT3 mutations were not detected. A biopsy specimen of the mediastinal tumor showed proliferation of leukemic blasts. RUNX1::RUNX1T1 AML with extramedullary involvement was diagnosed. Although the AML did not respond to two chemotherapy courses with cytarabine and anthracyclines (daunorubicin and idarubicin) and four courses of treatment with venetoclax combined with azacytidine, the patient achieved complete remission with partial hematological recovery of platelets to 90x10⁹/L after single-agent gemtuzumab ozogamicin (GO) treatment.

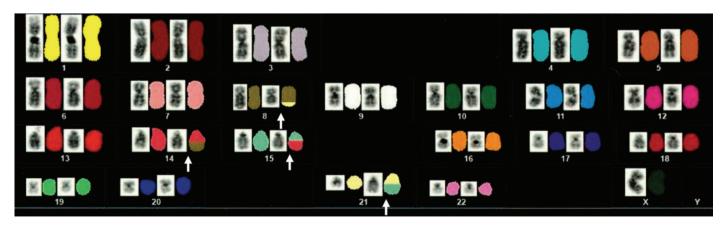


Figure 1. Spectral karyotyping with interphase fluorescence in situ hybridization reveals that distal regions of the 8q22, 14q22, 15q15, and 21q22.1 chromosomes were transferred in a cycle, resulting in t(8;14;15;21)(q22;q22;q15;q22.1). The arrows indicate rearranged chromosomal regions.

Excluding acute promyelocytic leukemia, there have been 8 AML cases presented to date with four-way translocations, including ours [1,2,3,4,5,6,7], among which 7 (88%) had t(8;21) translocations [1,2,3,4,5,6], indicating that four-way translocations are almost always associated with t(8:21) AML. The remaining case was acute megakaryocytic leukemia with t(1;22;17;18)(p13;q13;q22;q12) [7]. Cases of AML with fourway t(8;21) translocations do not share chromosomal regions, excluding 8g22 and 21g22, suggesting that RUNX1::RUNX1T1 fusion plays a central role in the pathogenesis of AML with fourway t(8;21) translocations. From a clinical viewpoint, only 3 of 6 patients with four-way t(8;21) translocations achieved complete remission after conventional chemotherapy with cytarabine and anthracyclines or mitoxantrone [2,3,4,5,6], including our case, and 2 of the 3 patients who achieved complete remission eventually relapsed [2,4], supporting the conclusion that fourway t(8;21) is a poor prognosis factor in RUNX1::RUNX1T1 AML. We used spectral karyotyping to demonstrate that the fourway t(8;21) translocations probably occurred consequent to cyclically ordered chromosomal translocations. The mechanisms remain unknown; however, they might be associated with single-event rearrangement via the simultaneous breakage of several chromosomes followed by mismatched joining. Our patient with refractory AML was successfully treated with CD33-targeting GO. In addition to high CD33 expression, NPM1 mutations, and FLT3 internal tandem duplication, core-binding factor rearrangements have been associated with favorable responses to GO [8]. In our case, the AML cells weakly expressed CD33 and had wild-type NPM1 mutations and FLT3. Regardless of the CD33 expression levels in bulk AML cells, it has been reported that t(8;21) progenitors express CD33 and are sensitive to GO [9,10]. We suggest that GO is a viable treatment option for refractory AML with four-way t(8;21) translocation.

Keywords: Acute myeloid leukemia, Four-way translocation, t(8;21), *RUNX1::RUNX1T1*

Anahtar Sözcükler: Akut myeloid lösemi, Dört-yol translokasyonu, t(8;21), *RUNX1::RUNX1T1*

Ethics

Informed Consent: Written informed consent was obtained from the patient for the publication.

Authorship Contributions

Surgical and Medical Practices: N.T., F.O., T.K., S.Y., K.K.; Concept: T.K., S.Y., K.K.; Design: K.K.; Data Collection or Processing: N.T., F.O., K.K.; Analysis or Interpretation: N.T., F.O., T.K., S.Y., K.K.; Literature Search: N.T., F.O., K.K.; Writing: N.T., F.O., T.K., S.Y., K.K.

Conflict of Interest: No conflict of interest was declared by the authors.

Financial Disclosure: This work was supported by JSPS KAKENHI Grant Number JP21K08396.

References

- de Greef GE, Hagemeijer A, Morgan R, Wijsman J, Hoefsloot LH, Sandberg AA, Sacchi N. Identical fusion transcript associated with different breakpoints in the AML1 gene in simple and variant t(8;21) acute myeloid leukemia. Leukemia 1995;9:282-287.
- Vieira L, Oliveira V, Ambrósio AP, Marques B, Pereira AM, Hagemeijer A, Boavida MG. Translocation (8;17;15;21)(q22;q23;q15;q22) in acute myeloid leukemia (M2). a four-way variant of t(8;21). Cancer Genet Cytogenet 2001;128:104-107.
- Albano F, Specchia G, Anelli L, Liso A, Zagaria A, Santoro A, Mirto S, Liso V, Rocchi M. Submicroscopic deletions in an acute myeloid leukemia case with a four-way t(8;11;16;21). Leuk Res 2005;29:855-858.
- Huang L, Abruzzo LV, Valbuena JR, Medeiros LJ, Lin P. Acute myeloid leukemia associated with variant t(8;21) detected by conventional cytogenetic and molecular studies: a report of four cases and review of the literature. Am J Clin Pathol 2006;125:267-272.
- Park KJ, Park HD, Kim HJ, Yoo KH, Koo HH, Kim SH. A novel four-way t(6;16;21;8)(p21.3;p11.2;q22;q22) in acute myeloid leukemia with *RUNX1/ RUNX1T1* rearrangement. Cancer Genet Cytogenet 2009;192:90-92.
- Isik S, Uskudar Teke H, Gunden G, Erzurumluoglu Gokalp E, Cilingir O, Artan S, Durak Aras B. A new four-way complex translocation variant involving the t(8;5;21;4)(q21;q13,q22,q31) and the relocalization of AML1/ETO fusion gene. Cancer Genet 2021;256-257:1-4.
- Torres L, Lisboa S, Vieira J, Cerveira N, Santos J, Pinheiro M, Correia C, Bizarro S, Almeida M, Teixeira MR. Acute megakaryoblastic leukemia with a four-way variant translocation originating the *RBM15-MKL1* fusion gene. Pediatr Blood Cancer 2011;56:846-849.
- Fenwarth L, Fournier E, Cheok M, Boyer T, Gonzales F, Castaigne S, Boissel N, Lambert J, Dombret H, Preudhomme C, Duployez N. Biomarkers of gemtuzumab ozogamicin response for acute myeloid leukemia treatment. Int J Mol Sci 2020;21:5626.
- 9. Al-Harbi S, Aljurf M, Mohty M, Almohareb F, Ahmed SOA. An update on the molecular pathogenesis and potential therapeutic targeting of AML with t(8;21)(q22;q22.1);*RUNX1-RUNX1T1*. Blood Adv 2020;4:229-238.
- Hills RK, Castaigne S, Appelbaum FR, Delaunay J, Petersdorf S, Othus M, Estey EH, Dombret H, Chevret S, Ifrah N, Cahn JY, Récher C, Chilton L, Moorman AV, Burnett AK. Addition of gemtuzumab ozogamicin to induction chemotherapy in adult patients with acute myeloid leukaemia: a meta-analysis of individual patient data from randomised controlled trials. Lancet Oncol 2014;15:986-996.



Address for Correspondence/Yazışma Adresi: Kensuke Kojima, M.D., Kochi Medical School, Department of
Hematology, Nankoku, JapanReceived/Geliş tarihi: January 25, 2024
Accepted/Kabul tarihi: March 15, 2024Phone : +81-88-888-2920E-mail : k-koji@kochi-u.ac.jp ORCID: orcid.org/0000-0003-4138-0604DOI: 10.4274/tjh.galenos.2024.2024.0038

129