

## Wound Healing Activity of Arum Maculatum

### Arum Macalatum Bitkisinin Yara İyileştirici Aktivitesi

## Ø Ayse ARZU SAKUL<sup>1</sup> Ø Mehmet Evren OKUR<sup>2</sup> Ø Sule AYLA<sup>3</sup> Ø Benay DAYLAN<sup>4</sup> Ø Ayse Esra KARADAG<sup>5</sup> Ø Sebnem BATUR<sup>6</sup> Ø Ekrem Musa OZDEMIR<sup>7</sup> Ø Ebrar ALTINALAN<sup>1</sup> Ø Mehmet Yalcin GUNAL<sup>8</sup>

<sup>1</sup>Istanbul Medipol University Faculty of Medicine, Department of Medical Pharmacology; Research Institute for Health Sciences and Technologies (SABITA), Istanbul, Turkey

<sup>2</sup>University of Health Sciences Faculty of Pharmacy, Department of Pharmacology, Istanbul, Turkey

<sup>3</sup>Istanbul Medeniyet University Faculty of Medicine, Department of Histology and Embryology, Istanbul, Turkey

<sup>4</sup>Istanbul Medipol University Faculty of Medicine, Department of Histology and Embryology, Istanbul, Turkey

<sup>5</sup>Istanbul Medipol University Faculty of Pharmacy, Department of Pharmacognosy, Istanbul, Turkey

<sup>6</sup>Istanbul University-Cerrahpasa, Cerrahpasa Faculty of Medicine, Department of Pathology, Istanbul, Turkey

<sup>7</sup>Istanbul Medipol University, Research Institute for Health Sciences and Technologies (SABITA); Medical Research Center (MEDITAM), Istanbul, Turkey

<sup>8</sup>Alanya Alaaddin Keykubat University Faculty of Medicine, Department of Physiology, Antalya, Turkey

#### ABSTRACT

**Objective:** In this study, the antioxidant properties of *Arum maculatum* plant were evaluated. This study reported for the first time the wound healing activity of the methanol extract of *A. maculatum* fruits. This study aimed to assess and determine the possible pharmacological activities of *A. maculatum* and evaluate its potential to act as a wound care plant.

**Methods:** The antioxidant and antimicrobial activities of *A. maculatum* were investigated using excisional *in vivo* and *in vitro* wound healing mouse models. A total of 32 Balb-c mice were used, which were equally, divided into four groups: saline control group, control group, *A. maculatum* group, and *Centella asiatica* extract group. Treatment applications were performed topically once per day. Wound area narrowing, wound healing percentage, and epithelialization time were analyzed.

**Results:** A. maculatum application supported the healing process in *in vivo* and *in vitro* wound models. A. maculatum contributed to the healing process by promoting granulation tissue formation, epidermal regeneration, and angiogenesis.

**Conclusions:** Wound healing is a complex and well-organized process that requires communication between cells. The antioxidant and antimicrobial activities of *A. maculatum* extract have been determined by current studies. *A. maculatum* extract may provide significant benefits in promoting the wound healing process.

Keywords: A. maculatum, wound healing, mice, antioxidant, extract

### ÖZ

**Amaç:** Bu araştırmada antioksidan özelliklerini değerlendirmek için *Arum maculatum* bitkisi seçilmiştir. Bildiğimiz kadarıyla *A. maculatum* meyvelerinin metanol özünün yara iyileştirici aktivitesi ilk kez bu çalışmada rapor edilmiştir. Bu çalışma, *A. maculatum*'un olası farmakolojik aktivitelerini belirlemek, değerlendirmek ve bir yara tedavi edici bitki olarak etki gösterme potansiyelini değerlendirmek içindi.

**Yöntemler:** A. maculatum'un antioksidan ve antimikrobiyal aktiviteleri, farelerde eksizyonel *in vivo* ve *in vitro* yara iyileşme modelleri kulanılarak araştırılmıştır. Toplamda 32 Balb-c fare kullanılmış olup salin kontrol grubu, kontrol grubu, A. maculatum uygulanan grup ve *Centella asiatica* özütü uygulanan grup olmak üzere 4 gruba ayrılmıştır. Tedavi uygulamaları günde bir kez topikal olarak gerçekleştirilmiştir. Skar alanı hacminde gerçekleşen değişim, yara iyileşme yüzdesi ve epitelizasyon süresi analiz edilmiştir.

**Bulgular:** A. maculatum uygulaması in vivo ve in vitro yara modelinde iyileşme sürecini desteklemiştir. A. maculatum, granülasyon dokusunu artırarak iyileşme sürecine katkıda bulunmuş, epidermal rejenerasyonu ve anjiyogenezi artırmıştır.

**Sonuçlar:** Yara iyileşmesi, hücreler arası iletişimi gerektiren karmaşık ve iyi organize edilmiş bir süreçtir. Mevcut çalışmalar doğrultusunda antioksidan ve antimikrobiyal aktivitesi belirlenmiş olan *A. maculatum* özü, yara iyileşme sürecinin desteklenmesinde önemli bir fayda sağlayabilir.

Anahtar kelimeler: A. maculatum, yara iyileşmesi, fareler, antioksidan, ekstrakt

Address for Correspondence: A. Arzu Sakul, Istanbul Medipol University Faculty of Medicine, Department of Medical Pharmacology; Research Institute for Health Sciences and Technologies (SABITA), Istanbul, Turkey **E-mail:** aasakul@medipol.edu.tr **ORCID ID:** orcid.org/0000-0002-9354-0000

Received: 07 October 2022 Accepted: 15 December 2022 Online First: 05 January 2023

Cite as: Arzu Sakul A, Okur ME, Ayla S, Daylan B, Karadag AE, Batur S, Ozdemir EM, Altinalan E, Gunal MY. Wound Healing Activity of Arum Maculatum. Medeni Med J 2023;38:8-15

©Copyright 2023 by the Istanbul Medeniyet University / Medeniyet Medical Journal published by Galenos Publishing House. Licenced by Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0)

### INTRODUCTION

As wounds are linked to the increase in loss of life, wound assessment and care are a challenge beyond medical society from ancient times onward<sup>1</sup>. Wounds gradually heal through well-organized processes that require cells to communicate with each other. This process is accompanied by cytokines, growth factors, and extracellular matrix (ECM) elements<sup>2</sup>. ECM proteins should maintain the skin's complex, and their roles are primarily found in connective tissue. Their relationship with skin cells is crucial in maintaining skin homeostasis and regeneration. In the phases of wound healing are as follows: (1) inflammatory phenotype transition that occurs in macrophages during the inflammatory process, (2) new artery development and angiogenesis stimulation by promoting endothelial cell differentiation and cell migration, and (3) granulation tissue, skin-cell, and ECM development, which leads to proliferation and remodeling. The above features can aid treatment plans in treating wounds and skin aging. They also sparked new research and clinical studies<sup>3</sup>.

Arum maculatum L. (Arum) belongs to the Araceae family and known as "yılan pancarı, yılan yastığı" in Turkey. The plant has many traditional uses such as for cancer, constipation, fungal diseases, hemorrhoids, and rheumatism. It is also used for its diaphoretic, sudorific, and expectorant properties. A. maculatum is an acceptable remedy for paralysis<sup>4</sup>. In a previous study about the biological activity of A. maculatum, Alencar et al.<sup>5</sup> investigated lectins' pro-inflammatory activity when isolated. In addition, the insecticidal action of lectins obtained from the tuber of the plant was clarified<sup>6</sup>. Previous studies have also investigated the cytogenic, anti-hemorrhoid, antimutagenic, antifungal, antiaflatoxigenic, anticholinesterase, antimicrobial, and antioxidant activities of several plants<sup>7-9</sup>. The analgesic activity and hepatotoxicity of A. maculatum methanol extract were examined in vivo in rats, and remarkable results were obtained<sup>8,10</sup>. Reactive oxygen species (ROS) can form during the wound healing process, and regular consumption of foods showing antioxidant activity can reduce these harmful effects<sup>11</sup>. Plant extracts have antioxidant effects<sup>12</sup>. In this study, A. maculatum was chosen to determine the plant's antioxidant properties; to our knowledge, during the preparation of this manuscript, this was the only study that examined the activity of A. maculatum methanol extract in wound healing in vivo. This study aimed to identify and establish the potential pharmacological properties of A. maculatum, as well as its potential as a wound care product.

### **MATERIALS and METHODS**

### Materials

In this study, 2,2-diphenyl-1-picrylhydrazyl (DPPH), 2,2'-azino-bis [3-ethylbenzothiazoline-6-sulfonic acid (ABTS)], Trolox, and ascorbic acid were provided by Sigma (Sigma-Aldrich Chemie GmbH, Steinheim, Germany). Only analytical-grade or higher chemicals, unless specified otherwise, were used.

### Preparation of A. maculatum Extract

A. maculatum fruits were collected from Silivri province, Istanbul (Turkey), in May 2019. Ayse Esra Karadag described the plant material. Voucher samples IMEF no. 1099 were deposited at the herbarium in the Department of Pharmacognosy of the Istanbul Medipol University Faculty of Pharmacy, Turkey. The plant sample was cut into small pieces and dried under fresh air. The air-dried plant sample was crushed, macerated for 24 h in methanol, and ground into a powder. The methanol extract was used to prepare gel formulations for further analysis after filtration and evaporation.

### **Antioxidant Activity**

### **DPPH Radical Scavenging Assay**

DPPH• was used to determine the total antioxidant capacity<sup>13</sup>. The reaction compound included a total of 100  $\mu$ M DPPH• in methanol and extract. The absorbance was measured at 517 nm using a UV spectrophotometer (UV-1800, Shimadzu, Japan) at 25 °C±2 °C after 30 min. As a % of radical reduction, the radical scavenging activity (RSA) was determined using the following equation:

DPPH•RSA% = [(Absorbance<sub>control</sub>-Absorbance<sub>test\_sample</sub>)/ Absorbance<sub>control</sub>)] ×100

### **ABTS Radical Scavenging Assay**

The ABTS radical cation decolorization protocol was used to evaluate the antioxidant capacity of *A. maculatum*<sup>14</sup>. To produce ABTS•, a 2.45 mM potassium persulfate and 7 mM aqueous ABTS mixture reacted. Before using it, the mixture was kept at a 25 °C dark room for 16 h. The final mixture was stored in a dark room at a temperature of 25 °C for 16 h. The mixture was added with ethanol, and its absorbance at 25 °C was calculated at 734 nm. The procedure was carried out three times, and Trolox was chosen as the "positive control"<sup>15</sup>. Table 1 presents the results calculated as IC<sub>50</sub>.

ABTS•RSA% = [(Absorbance<sub>control</sub>-Absorbance<sub>test sample</sub>)/ Absorbance<sub>control</sub>]] ×100

Table 1. <i>A. maculatum</i> ABTS and DPPH radical scavenging activities (ICD <sub>50</sub> ± mg/mL).		
	A. maculatum	Reference compounds
	IC <sub>50</sub> ± SEM (mg/mL)	
ABTS·	0.75±0.15	0.013±0.004 (Trolox)
DPPH•	0.52±0.18	0.004±0.001 (Ascorbic acid)
ABTS: 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid), DPPH: 2,2-diphenyl-1-picrylhydrazyl,		

### In Vivo Experiments

### **Research Subjects**

rate was calculated using the formula specified in the literature<sup>17</sup>.

# Balb-c mice weighing 24-28 g were used. The experimental animals were held inside houses in line with standards at 24 °C, had free access to food and water, and exposed to indoor lighting between 0700 and 1900 hours. Ethical approval was granted by the Istanbul Medipol University Animal Experiments Local Ethics Committee (decision no: 91, date: December 19, 2019) before the experiments.

# Protocols of Wound Formation and Experiment Groups

The animals used in the experiment were divided into four (8 animals in each) groups:

Control: Treated with saline (saline control group)

Group 1: Vehicle group (control group)

Group 2: A. maculatum group.

Group 3: *Centella asiatica* extract (CAE) group (reference molecule, Madecassolâ cream)

Base gel, which served as the vehicle, was prepared according to a previous study<sup>16</sup>. To prepare the extract gel (5%, w/w), *A. maculatum* extract was added to the base gel and stirred slowly until a homogeneous gel was formed.

Mice were anesthetized with a mixture of ketamine (80-100 mg/kg) and xylazine (10 mg/kg) administered intraperitoneally. To create two wound tissues, the back region was shaved, and excisional scar tissue was created using a 5-mm biopsy apparatus on the left side of the midline. Wound tissues were localized 1 cm from each other and 1.5 cm from the midline. Topical treatment application to the wound areas was continued for 10 days.

### Macroscopic Assessment

To score wound healing, wound areas were photographed at the beginning (day 0) and end of the application (day 10). Areas of wound surfaces were measured using the Image J software. The wound healing **Histological Assessment** 

The scar tissues were removed at the end of day 10 after the animals were euthanized. Then, 10% neutral formalin was used for fixation. Furthermore, ethanol was used to dehydrate the wound samples, and toluene was used to clear them. The samples were then embedded in paraffin. Thereafter, 5  $\mu$ m-thick sections of the samples were subjected to hematoxylin-eosin (HE) and immunohistochemical staining: vascular endothelial growth factor (VEGF; Santa Cruz sc-7269], collagen (COL1A1; Santa Cruz sc-293182), platelet-derived growth factor (PDGF-A; Santa Cruz sc-9974) on glass slides.

Wound healing was evaluated histologically using the method defined by Galeano et al.<sup>18</sup> and Ayla et al.<sup>16</sup>. In terms of angiogenesis, 1 point indicates one to two vessels/site; 2 points, few neovascularizations (3-4/site); 3 points, newly formed capillary vessels (5-6/site); and 4 points, newly formed and normal looking capillary vessels (>7/site). In the assessment of angiogenesis, four microscopic fields in each subject were evaluated at ×20 magnification.

In terms of epidermal organization, 1 point indicates weak epidermal creation ( $\geq$ 20%); 2 points, incomplete epidermal creation ( $\geq$ 40%); 3 points, moderate epithelial creation ( $\geq$ 60%); and 4 points, complete epidermal organization ( $\geq$ 80%). Granulation tissue thickness was measured as follows: 1 point, weak granulation layer; 2 points, intermediate granulation layer; 3 points, dense granulation layer; and 4 points, very thick layer.

In this study, a quantitative method was used to determine the immunoreactivity of immunohistochemical staining. Five fields were randomly selected, and these fields were evaluated and averaged: 0 points, no staining; 1 point, poor staining; 2 points, moderate staining; and 3 points, strong staining<sup>16,19</sup>.

### **Statistical Analysis**

Tests were conducted using IBM SPSS Statistics for Windows version 20 (IBM Corp., Armonk, NY, USA). Intergroup differences were analyzed using analysis of variance and the least significant differences tests. All values were presented as mean  $\pm$  standard error of the mean, and a p-value <0.05 indicated the significance limit.

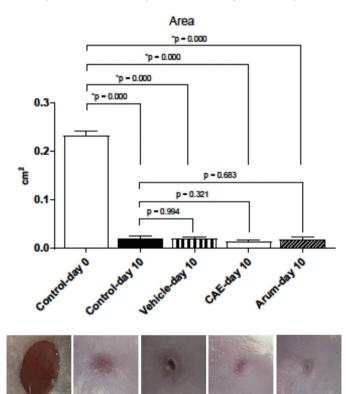
### RESULTS

### **Antioxidant Activity**

Treatment compounds containing antioxidants have been demonstrated to aid in wound healing and protect them from oxidative damage<sup>16</sup>. Table 1 presents the study results of *A. maculatum* extract's antioxidant activity. According to comparisons with standard substances, *A. maculatum* extract was thought to have effective antioxidant activity. As a result of this study, it has been shown that *A. maculatum* extract has an antioxidant effect. This antioxidant capacity was thought to have an important effect on the wound healing process.

### Macroscopic Wound Healing

The wound healing rates and macroscopic wound healing are shown in Figure 1. After day 10, no significant



**Figure 1.** Healing of the wound area and macroscopic wound healing observations for the control group, vehicle group, CAE group, and *A. maculatum* group.

CAE: Centella asiatica extract, A. maculatum: Arum maculatum

wound contraction difference was found between the *A. maculatum* (Arum) group and the control group ( $p \ge 0.05$ ) (Figure 1). However, when compared with day 0, a significant difference in wound contraction was found. Similarly, no significant wound contraction difference was found between the CAE group and the control group after day 10 (p=0.321). However, a significant difference was observed between day 0 and the control group. The outcome was the same for the vehicle group (Figure 1).

### **Histology of Wound Healing**

Histological (H&E) and immunohistochemical (VEGF, PDGF, and collagen) evaluations were appraised separately. The results of the macroscopic evaluation are presented in Figure 1. Histopathologically, histological evaluation (Figure 2) and immunohistochemical staining data, which enable comparison of immunohistochemistry collagen, PDGF, and VEGF wound healing scores between the groups, are shown in Figure 3. Epidermal regeneration, thickness of the granulation tissue, and angiogenesis from HE sections were evaluated and scored separately (Figure 2). Both A. maculatum extract (p<0.0001) and CAE (p<0.0001) were more potent than the control and vehicle groups in terms of vascularization, granulation tissue development, and epidermal tissue integrity (Figure 2). PDGF and VEGF immunohistochemical staining performed in the CAE groups was more potent than those in the control and vehicle groups (p≤0.05) (Figure 3). However, only VEGF staining increased significantly for A. maculatum (p≤0.02) (Figure 3); however, PDGF (p=0.08) and collagen (p=0.8) staining was not significantly different between the A. maculatum group and the control group (Figure 3).

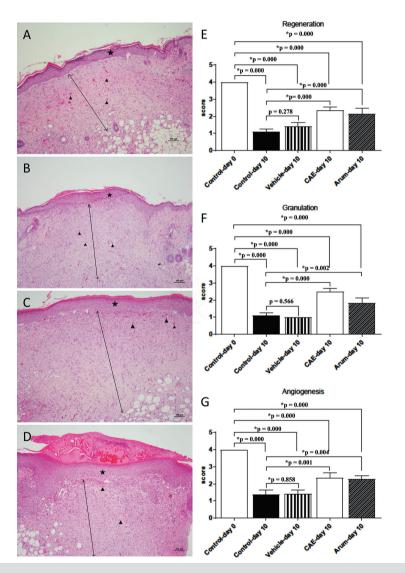
### DISCUSSION

Since wound assessment and treatment is a complex process involving various aspects with significant influence on healing, wound healing is an unfulfilled clinical challenge in healthcare<sup>1</sup>. Wounds occur when tissue integrity is disrupted. Wound healing is frequently studied, and some of its mechanisms still need explanation<sup>20</sup>. According to traditional definitions, hemostasis, inflammation, proliferation, and tissue remodeling are four separate stages of wound healing. The arrangement of these stages in wound healing relies on factors such as cytokines, growth factors, proteases, eicosanoids, kinins, and cellular metabolites<sup>21</sup>, and these may delay wound healing. Previously, all wounds heal with complications such as scars and infection; however, at present, side effects related to wound healing are reduced. Despite this, chronic wounds still bore doctors<sup>22</sup>.

Plants have been traditionally utilized to improve wound healing as topical formulations, and they proved beneficial in wound care and accelerate wound healing without causing distress, pain, and scarring<sup>23</sup>.

As a result of phytochemical screening, *A. maculatum* was found to contain glycosides such as flavonoids, saponins, cyanogenes, alkaloids, polyphenols, 2-heptanone, (E)-caryophyllene, indoles, p-cresol, proanthocyanidins, monoterpenes, two sesquiterpenes, and lectins<sup>10</sup>.

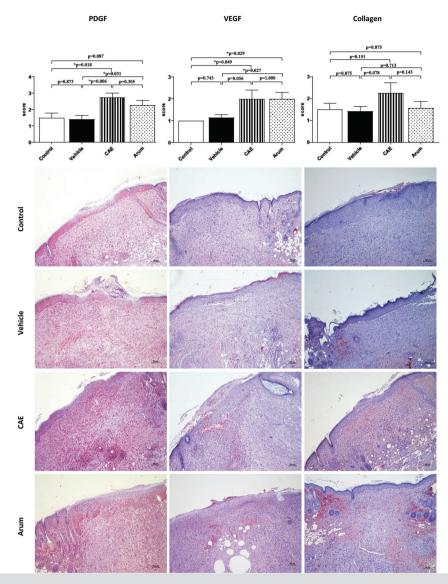
Because of their capacity to scavenge free radicals, singlet oxygen, and superoxide radicals, phenolic compounds are classified as strong antioxidants. The antioxidant potential related to mature collagen fiber growth and enhanced angiogenesis supported by fibroblasts was determined to be responsible for these effects<sup>24</sup>. According to the above studies, *A. maculatum* fruits have a high phenolic profile. Therefore, the wound healing activity can be considered to result from this phenolic profile.



**Figure 2.** Histopathological view of the injured tissues of the control (A), vehicle gel (B), CAE (C), and A. maculatum (Arum) (D) extract gel on day 10 after wound incision (original magnification, ×10). (E-G) Histological scores of epidermal regeneration, granulation tissue thickness, and angiogenesis of the control, vehicle gel, CAE, and A. maculatum (Arum) groups. Arrow, granulation tissue thickness; star, epidermal regeneration; triangle, angiogenesis. Statistically significant when compared with the control group; p<0.05. Values are presented as the mean  $\pm$  SEM. The scale bars represent 100  $\mu$ m.

CAE: Centella asiatica extract, A. maculatum: Arum maculatum





**Figure 3.** Comparison of immunohistochemical collagen, PDGF, and VEGF wound healing scores among the groups. Statistically significant when compared with the control group; p<0.05 (\*). Values are presented as the mean  $\pm$  SEM. Histopathological view of the injured tissues of the untreated (control), vehicle, *A. maculatum* (Arum), and CAE groups onday 10 after wound incision (original magnification, ×10).

CAE: Centella asiatica extract, A. maculatum: Arum maculatum, PDGF: Platelet-derived growth factor, VEGF: Vascular endothelial growth factor

Wounds heal to reduce inflammatory factors and increase tissue formation with a new blood vessel<sup>20</sup>. Studies have used herbal extracts to speed wound healing, and many agents increase cell proliferation, angiogenesis, and collagen production. Therefore, these compounds have been used for these properties<sup>25,26</sup>. Previous studies have also investigated the antimicrobial and antioxidant activities of various plants, such as *A. maculatum*<sup>7</sup>. In fact, *A. maculatum* was used for wound healing purposes. In addition, the use of *A. maculatum* 

to treat hemorrhoids among the general population may be associated with its wound healing effect<sup>27</sup>. However, no studies have shown its effect on wound healing. In this study, topical application of *A. maculatum* was found to improve skin healing. A statistically significant boost in the re-epithelialization, angiogenesis, and granulation tissue thickness was found in the *A. maculatum* group or the CAE group, compared with the control group. Accordingly, scientific data confirmed the effectiveness of this herbal preparation. When the hemostatic plug is formed, platelets produce transforming growth factor beta, PDGF, and fibroblast growth factor, inducing angiogenesis. In this study, *A. maculatum* was observed to increase wound healing, primarily by increasing VEGF. Angiogenesis improves fibroblast activity and feeding wound area. The fibroblast activity contributes to the wound healing process by contributing to the formation of granulation tissue. Fibroblast proliferation and migration are decreased by ROS in wounds, which decreased collagen synthesis<sup>28</sup>. We can attribute this to the absence of a difference in collagen staining between the *A. maculatum* group and the control group.

In this study, *A. maculatum* was observed to promote the healing of skin wounds. Epidermal regeneration and granulation tissue were increased in the *A. maculatum* and CAE groups compared with those in the saline control group. Thus, *A. maculatum* extract could be a new alternative in treating wounds, owing to its feature similar to CAE.

### CONCLUSION

The results of this show that topical treatment of any wound with *A. maculatum* promotes advanced wound contraction and helps in healing experimental skin wounds in mice. In light of the present results, *A. maculatum* extract could contribute significantly to the wound healing process *in vivo* owing to its antioxidant activity. Owing to the great *in vivo* wound healing behavior of *A. maculatum* extract, further investigation on larger animal models or humans could confirm its possible application for wound healing.

### Ethics

**Ethics Committee Approval:** Ethical approval was granted by the Istanbul Medipol University Animal Experiments Local Ethics Committee (decision no: 91, date: December 19, 2019) before the experiments.

Informed Consent: Animal experiment study.

**Peer-review:** Externally and internally peer-reviewed.

### **Author Contributions**

Surgical and Medical Practices: A.A.S., M.E.O., S.A., B.D., A.E.K., S.B., E.M.O., E.A., M.Y.G., Concept: A.A.S., Design: A.A.S., M.E.O., Data Collection and/or Processing: A.A.S., M.E.O., S.A., B.D., A.E.K., S.B., E.M.O., Analysis and/or Interpretation: A.A.S., M.E.O., S.A., M.Y.G., Literature Search: A.A.S., M.E.O., A.E.K., E.A., Writing: A.A.S., S.A. **Conflict of Interest:** The authors have no conflict of interest to declare.

**Financial Disclosure:** The authors declared that this study has received no financial support.

### REFERENCES

- Okur ME, Karantas ID, Şenyiğit Z, Üstündağ Okur N, Siafaka PI. Recent trends on wound management: New therapeutic choices based on polymeric carriers. Asian J Pharm Sci. 2020;15:661-84.
- 2. Pang X, Dong N, Zheng Z. Small Leucine-Rich Proteoglycans in Skin Wound Healing. Front Pharmacol. 2020;10:1649.
- Mazini L, Rochette L, Amine M, Malka G. Regenerative Capacity of Adipose Derived Stem Cells (ADSCs), Comparison with Mesenchymal Stem Cells (MSCs). Int J Mol Sci. 2019;20:2523.
- Alachkar A, Jaddouh A, Elsheikh MS, Bilia AR, Vincieri FF. Traditional medicine in Syria: folk medicine in Aleppo governorate. Nat Prod Commun. 2011;6:79-84.
- Alencar VB, Alencar NM, Assreuy AM, Mota ML, Brito GA, Aragão KS, Bittencourt FS, Pinto VP, Debray H, Ribeiro RA, Cavada BS. Pro-inflammatory effect of Arum maculatum lectin and role of resident cells. Int J Biochem Cell Biol. 2005;37:1805-14.
- Majumder P, Mondal HA, Das S. Insecticidal activity of Arum maculatum tuber lectin and its binding to the glycosylated insect gut receptors. J Agric Food Chem. 2005;53:6725-9.
- Erbil N, Arslan M, Murathan ZT. Antioxidant, Antimicrobial, and Antimutagenic Effects and Biochemical Contents of Arum Maculatum L. That Is a Medical Plant from Turkish Flora. Fresen Environ Bull. 2018;27:8709-14.
- Farahmandfar R, Esmaeilzadeh Kenari R, Asnaashari M, Shahrampour D, Bakhshandeh T. Bioactive compounds, antioxidant and antimicrobial activities of Arum maculatum leaves extracts as affected by various solvents and extraction methods. Food Sci Nutr. 2019;7:465-75.
- Kurt BZ, Gazioğlu I, Sevgi E, Sönmez F. Anticholinesterase, Antioxidant, Antiaflatoxigenic Activities of Ten Edible Wild Plants from Ordu Area, Turkey. Iran J Pharm Res. 2018;17:1047-56.
- Nabeel M, Abderrahman S, Papini A. Cytogenetic Effect of Arum maculatum Extract on the Bone Marrow Cells of Mice. Caryologia. 2014;61:383-7.
- Farhoosh R, Johnny S, Asnaashari M, Molaahmadibahraseman N, Sharif A. Structure-antioxidant activity relationships of o-hydroxyl, o-methoxy, and alkyl ester derivatives of p-hydroxybenzoic acid. Food Chem. 2016;194:128-34.
- Bonoli-Carbognin M, Cerretani L, Bendini A, Almajano MP, Gordon MH. Bovine serum albumin produces a synergistic increase in the antioxidant activity of virgin olive oil phenolic compounds in oilin-water emulsions. J Agric Food Chem. 2008;56:7076-81.
- 13. Blois MS. Antioxidant Determinations by the Use of a Stable Free Radical. Nature. 1958;181:1199-200.
- Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radic Biol Med. 1999;26:1231-7.
- Okur ME, Ayla Ş, Çiçek Polat D, Günal MY, Yoltaş A, Biçeroğlu Ö. Novel insight into wound healing properties of methanol extract of Capparis ovata Desf. var. palaestina Zohary fruits. J Pharm Pharmacol. 2018;70:1401-13.

- Ayla S, Okur ME, Gunal MY, et al. Wound healing effects of methanol extract of Laurocerasus officinalis roem. Biotech Histochem. 2019;94:180-8.
- Günal MY, Okçu Heper A, Zaloglu N. The Effects of Topical Carvacrol Application on Wound Healing Process in Male Rats. Pharmacognosy Journal. 2014;6:10-4.
- Galeano M, Altavilla D, Bitto A, et al. Recombinant human erythropoietin improves angiogenesis and wound healing in experimental burn wounds. Crit Care Med. 2006;34:1139-46.
- Tüzün F, Gencpınar P, Ozbal S, Dilek M, Ergur BU, Duman N, Ozkan H, Kumral A. Neuroprotective effect of neotrofin in a neonatal rat model of periventricular leukomalacia. Neurosci Lett. 2012;520:6-10.
- Ayla Ş, Günal MY, Sayın Şakul AA, et al. Effects of Prunus spinosa L. fruits on experimental wound healing. Medeniyet Medical Journal. 2017;32:152-8.
- Henry G, Garner WL. Inflammatory mediators in wound healing. Surg Clin North Am. 2003;83:483-507.
- Mustoe TA, O'Shaughnessy K, Kloeters O. Chronic wound pathogenesis and current treatment strategies: a unifying hypothesis. Plast Reconstr Surg. 2006;117(7 Suppl):35S-41.

- Cetin EO, Yesil-Celiktas O, Cavusoglu T, Demirel-Sezer E, Akdemir O, Uyanikgil Y. Incision wound healing activity of pine bark extract containing topical formulations: a study with histopathological and biochemical analyses in albino rats. Pharmazie. 2013;68:75-80.
- Lodhi S, Singhai AK. Wound healing effect of flavonoid rich fraction and luteolin isolated from Martynia annua Linn. on streptozotocin induced diabetic rats. Asian Pac J Trop Med. 2013;6:253-9.
- Kawanabe T, Kawakami T, Yatomi Y, Shimada S, Soma Y. Sphingosine 1-phosphate accelerates wound healing in diabetic mice. J Dermatol Sci. 2007;48:53-60.
- Somboonwong J, Kankaisre M, Tantisira B, Tantisira MH. Wound healing activities of different extracts of Centella asiatica in incision and burn wound models: an experimental animal study. BMC Complement Altern Med. 2012;12:103.
- 27. Ivancheva S, Stantcheva B. Ethnobotanical inventory of medicinal plants in Bulgaria. J Ethnopharmacol. 2000;69:165-72.
- Zeng Z, Zhu BH. Arnebin-1 promotes the angiogenesis of human umbilical vein endothelial cells and accelerates the wound healing process in diabetic rats. J Ethnopharmacol. 2014;154:653-62.