

## DISTRIBUTION AND SEASONAL OCCURRENCE OF AQUATIC PHYCOMYCETES IN WATER AND SUBMERGED MUD IN EL-IBRAHIMIA CANAL (UPPER EGYPT)

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*SUMMARY: From 33 tested grains and seeds, sesame, hemp seeds, barley and maize grains proved to be best baits for aquatic phycomycetes in El-Ibrahimia Canal. The richest periods in aquatic fungi were of low or moderate temperature months and the poorest were summer months. Organic matter contents of water and mud samples showed its highest values in winter months where pH values did not perform any regular seasonal variation. Achlya was the most prevalent genus in water samples (23 months) while Dictyuchus and Pythium were the most common genera in submerged mud (each 24 months) Apodachlya, Ganopodya, Calyptralegnia, Leptomitus and Woronina emerged from water samples only while Aqualindrella, Blastocladia, Blastocladiella, Leptolegnia and Nowakowskia emerged from submerged mud. Achlya dubia, A. proliferoides, A. debaryana, A. carolinina, A. polyandra, A. combrica, A. oligacantha, A. apiculata, A. hypogna, A. radiosa, Sap-rolegnia megasperma, S. anisospora, E. parasitica, S. diclina, S. furcata, S. trufosa, S. uliginosa, S. litoralis, Allomyces javanicus, Isoachlya monilifera, I. eccentrica and Pilobolus kelinii emerged from surface water only. On other side, Pythium echinulatum, P. intermedium, Phytophthora cinchonae, Brevilegnia unisperma var. Delica, Isoachlya unispora and Pilobolus nanus emerged from submerged mud. Saprolegnia disappeared completely during summer months in both surface water and submerged mud samples.*

*Key Words: Phycomycetes, achlya, pythium.*

### INTRODUCTION

The distribution and seasonal fluctuations of aquatic zoospore fungi in relation to environmental conditions as well as to the various geographical regions of the world have been intensively investigated by many authors, e.g. Lund (36) in Denmark, Johnson (28) in Scandinavia, Forbes (21), Waterhouse (51, 52), Perrot (42) and Willo-ughby *et al.* (55) in the United Kingdom, Johnson *et al.* (27) in ICE Land, Jacobson (25) in Germany, Zebrowska (58) in Poland, Coker (6), and Klick and Tiffany (33) in the USA, Nolan (41) in Canada, Karling (29) and Milanez (37, 38) in Brazil, Carronco *et al.* (4) in Mexico, Rossy (46) in Puerto Rico, Karling (30) and Elliot (15) in New Zealand, Youatt (56) in Australia, Knox and Peterson (34) in Antarctica, Naumov (40) in the Soviet Union, Okena (1978) in Japan, Chein (5) in Taiwane, Yung and Stenton (57) in Hong Kong, Chaudri *et al.* (1947), Dayal and Tandon (7) and Misra (39) in India, Rattan *et al.* (19) in Nigeria and El-Hissy (8-11), El-Hissy *et al.* (13), El-

Hissy and El-Nagdy (12), El-Nagdy (16-17), Abd-Allah (1) and El-Hissy and Khallil (14), in Egypt.

### MATERIALS AND METHODS

Ibrahimia canal is the biggest irrigation canal in Egypt. It arises from the River Nile at Assiut and extends to about 360 kilometers up to El-Giza. Its width is about 160 meters which decreases as the canal extends and the water depth is about 6 meters which also decreases as the canal extends.

A preliminary experiment was conducted to choose the best baiting substances for the recovery of aquatic fungi. Thirty-three grains and seeds were tested as baits. These were, *Hordeum vulgare*, *Oryza sativa*, *Sorghum virgatum*, *Sporobolus airoides*, *Triticum vulgare*, *Zea mays*, *Asphodelus sp.*, *Pancreatium sp.*, *Cicer arietinum*, *Lupinus termis*, *Lens esculenta*, *Lagonchichium sp.*, *Phaseolus vulgaris*, *Trifolium alexandrinum*, *Vicia faba*, *Ceratonia siliqua*, *Capsicum sp.*, *Hyposcyamus muticus*, *Eruca sativa*, *Raphanus sativus*, *Pimpinella anisum*, *Citrus sinensis*, *Sesame indicum*, *Nigella sativa*, *Arthrocnemon glaucum*, *Ricinus communis*, *Calitrops sp.*, *Zygophyllum album*, *Cannabis sativa*, *Hibiscus esculentus*, *Gossypium barbadense*, *Carthamus tinctorius* and *Hellianthus annuus*.

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Surface water samples were collected monthly during the period from December 1981 to November 1983 and brought directly to the laboratory in sterile clean conical flasks. The organic matter content and total soluble salts of the water samples were determined. PH values and temperature were also recorded. For the recovery of aquatic fungi from the collected water samples, aliquots of canal water (about 150 ml each) were introduced in 16 sterile Petri-dishes (15 cm in Diameter). Halves of sterilized hemp., sesame seeds and barley, maize grains, which proved to be the best baits, were introduced into the above to be the best baits, were introduced into the above treated Petri-dishes (4 plates for each bait). These dishes were then left overnight at room temperature. Then the colonized baits were transferred into sterile Petri-dishes containing sterile filtered canal water to which crystalline penicillin (2000 units/liter; Roberts 1963) was added, to depress bacterial growth. The dishes were then incubated at  $22\pm 2^{\circ}\text{C}$  for 4-6 weeks during which the dishes were weekly examined. The recovered aquatic fungi were purified on glucose-peptone (Gp) agar medium (54).

During the same period and from the same site where the water samples were collected monthly, submerged mud were also collected in clean plastic bags and were immediately brought to the laboratory, from which the aquatic fungi were recovered:

50 gm of mud were introduced in a sterile 2 L conical flask and were raised to 1500 ml by adding sterile distilled water. Technical flask was then shaken gently for about 5 minutes. Ten Petri-dishes (15 cm in diameter) were plated each with 100 ml of the supernatant mud wash water. Sterilized baits (sesame-, hemp seeds and barley-, maize grains) were introduced into dishes which were then treated exactly as described before to isolate and identify aquatic phycomycetes. The purified aquatic fungal genera and species were identified according to the following references (6, 20, 26, 31, 44, 47-49, 51, 52, 59).

## RESULTS AND DISCUSSION

### Water samples

The results in Table 1 show that the richest periods in aquatic fungal genera and species were December 1981-April 1982, November-December 1982, January-April 1983 and October-November 1983. These periods represent low or moderate temperature months ( $14-20^{\circ}\text{C}$ ). The poorest periods were from May-September (1982, 1983) which are almost summer months with relatively high temperature ( $21-27^{\circ}\text{C}$ ). These results are in accordance with those obtained by many investigators, Forbes (21, 22) studied the aquatic fungi in many ponds in the United Kingdom, reported a marked periodic variation in abundance. This variation generally consists of a gradual increase in the number of records up to a maximum abundance, sometimes during the winter, and then a corresponding decrease until the species may apparently disappear altogether in the summer. Dayal and Tandon (7), Roberts (1963), El-

Nagdy (16), El-Hissy *et al.* (13), Misra (39) and Khallil (32), reported that the low or moderate temperature periods are favorable for aquatic fungal population.

Fifty-two species which belong to twenty genera were collected during this experiment. The genera namely, *Achlya*, *Dictyuchus*, *Pythium* and *Saprolegnia* were of high frequency of occurrence and emerged in 23, 23, 23 and 15 out of 24 months respectively. *Phytophthora* and *Pythiopsis* were of moderate occurrence (9 and 6 months) respectively. The remaining genera were of low or rare occurrence (1-5 months).

*Achlya* was the leading organism and contributed the broadest spectrum of species (12 identified species) in addition to an unidentified species. The richest months in addition to an unidentified species. The richest months in *Achlya* species were June, 1982, June, August and November 1983 yielding 3 species each in addition to unidentified one. The unidentified species, *A. racemora*, *A. dubia*, *A. proliferoides* and *A. debaryana* were the most common species and emerged in (19, 7, 5, 5 and 4 months) respectively. This result was in agreement with that obtained by Chauduri *et al.* (1947) in India, who reported that *Achlya* was the most common genus throughout the year. Klick and Tiffany (33) reported that occurrence of *Achlya* showed its peak not only in the spring and fall but also a third peak in mid-summer. Generally, in the present work, it was observed that centric or sub-centric oospore species of *Achlya* predominated in low or moderate temperature months and those with eccentric oospores were prevalent in relatively higher temperature months. Similar results were obtained by Forbes (21, 22), and Rattan *et al.* (44).

*Dictyuchus* was also of high occurrence (23 months) and contributed five species. The richest months in *Dictyuchus* were February and March, 1982, 1983 (4 species each). Rattan *et al.* (44) in Iraq, mentioned that the species of *Dictyuchus* occur throughout the year but seem to grow best at moderate temperature ( $19-27^{\circ}\text{C}$ ) in spring and autumn. Misra (39) in India, found that *Dictyuchus* species predominated only in winter months.

*Pythium* was represented by *P. undulatum*, *P. thalassium* and unidentified species, which emerging in (5, 2 and 23 months) respectively. This is in accordance with the results obtained by Höhnk and Bock (23) and Waterhouse (50) who reported that *Pythium* has appeared nearly in all months. Moreover, El-Nagdy (16,17), El-Hissy *et al.* (13) and Khallil (32) reported that *Pythium* did not show any regular seasonal trend. On the other hand, El-Sharouny (18) working on *Pythium* in Egyptian soils, reported that *Pythium* was recovered in relatively high counts in cold months and was almost absent in summer months.





### Submerged mud

Thirty-seven species which belong to twenty aquatic fungal genera were recovered from monthly collected submerged mud samples. The richest periods in aquatic fungi were December 1981-May 1982, October-December 1982, January-March 1983 and October-December 1983 which represent moderate or low temperature month months (14-18°C). The poorest periods were from June-September 1982 and April-August 1983 which are almost summer months (21-27°C).

*Dictyuchus*, *Pythium* and *Phytophthora* were of high frequency of occurrence and emerged in (24, 24 and 16 months out of 24) respectively. The richest months in *Dictyuchus* were February and March 1982 and March 1983 (4, 5 and 4 species respectively). *Pythium* was also encountered throughout the experimental period and contributed four species only. *Phytophthora*, *Saprolegnia*, *Achlya*, *Brevilegnia*, *Aqualindrella*, *Pythiopsis*, *Rhizophydium*, *Olpidium*, *Nowakowskia* and *Pilobolus* emerged in 16, 10, 10, 9, 9, 8, 11, 7, 4 and 6 months respectively. Variable results concerning distribution and periodicity of aquatic phycomycetes in various soil and mud types all over the world were obtained by many investigators. In this respect, Rao and Venkateswarlu (43) in their study concerning microbial ecology of the soil of Indian desert, reported that no significant decline in the population of microorganisms was observed during summer, in spite of the high surface soil temperature which may sometimes reach 50°C.

Willoughby (53) in the United Kingdom, concluded that the moisture content of soil samples represents the main important factor in determining the occurrence of aquatic fungal flora in these samples. Lund (36) in Denmark, reported that the majority of aquatic fungi were isolated from the moist soil samples. El-Hissy (10) in Egypt found that the water content of the soil samples is an effective factor on the occurrence of aquatic fungi in soil.

It was interesting to isolate *Pilobolus*, which is known to be coprophilous, from water and submerged mud. It is probable that its spores were ejected from terrestrial mycelium to water where they could colonize the experimental baits and therefore it was recorded in this experiment.

In comparison between aquatic phycomycetes which recovered from surface water and submerged mud samples of the canal, it was found that; 1. *Achlya* was the most prevalent genus in water samples (23 months) while *Dictyuchus* and *Pythium* were predominated in submerged mud (24 months each). 2. *Apodachlya*, *Calyptrolegnia*, *Gonapodya*, *Leptomitus* and *Woronina* emerged from water samples only while *Aqualindrella*, *Blastocladiella*, *Blastoclaadia*, *Leptolegnia* and *Nowakowskia* emerged from submerged mud. 3. *Achlya dubia*, *A. proliferoides*, *A. debaryana*, *A. caroliniana*, *A. polyandra*, *A.*

*cambrica*, *A. oligacantha*, *A. apiculata*, *A. hypogyna*, *A. radiosa*, *Saprolegnia megasperma*, *S. anissospora*, *S. parasitica*, *S. diclina*, *S. furcata*, *S. turfosa*, *S. uliginosa*, *S. littoralis*, *Allomyces javanicus*, *Isoachlya monilifera*, *I. eccentrico* and *Pilobolus kleinii* emerged from surface water and disappeared in mud samples. 4. On the other side, *Pythium echinulatum*, *P. intermedium*, *Phytophthora cinchonae*, *Brevilegnia unisperma var. delica*, *Isoachlya unispora* and *Pilobolus nanus* recovered from mud samples only.

### REFERENCES

1. Abd-Elaah GA : Studies on aquatic fungi from Egyptian soil. M Sc Thesis, Department of Botany Faculty of Science, Sohag. Assiut University, Egypt, p 113, 1986.
2. Alabi RO : Studies of some tropical aquatic phycomycetes found around Ibadan. M Sc dissertation, University of Ibadan, 1967.
3. Alabi RO : Seasonal periodicity of Saprolegniaceae at Ibadan, Nigeria. Trans Br Mycol Soc, 56:337-341, 1971.
4. Carranco PD, Hernandez A, Rivera P, Rosas I : Soil and aquatic fungi in a waste-Stabilization pond system of the state of Mexico. Air Soil Pollut, 23:249-256, 1984.
5. Chein CY : Studies on Taiwanese aquatic fungi. I. Blastocladia and Allomyces. Trans Mycol Soc, 12:178-185, 1974.
6. Coker WC : The Saprolegniaceae, with notes on other water molds. Univ of North Carolina Press, Chozel Hill, North Carolina, p 201, 63 pls, 1923.
7. Dayal R, Tandon RN : Ecological studies of some aquatic Phycomycetes. Hydrobiologia, 20:121-127, 1962.
8. El-Hissy FT : Freshwater fungi in Egypt. Egypt J Bot, 17:187-189, 1974.
9. El-Hissy FT : Seasonal fluctuations of freshwater fungi in River Nile. The First Scientific Conference of Egyptian Graduate Abroad, London, 1979.
10. El-Hissy FT : On the aquatic fungi of Egyptian soil. Bul Fac Sci Assiut Univ, 8:99-107, 1979.
11. El-Hissy FT : Aquatic and terrestrial fungi from the surfaces and the casts of earth-worms in Egypt. Bul Fac Assiut Univ, 1979.
12. El-Hissy FT, El-Nagdy MA : Aquatic phycomycetes on the mud of the River Nile, Assiut, Egypt. Sydowia, 36:118-124, 1983.
13. El-Hissy FT, Moubasher AH, El-Nagdy MA : Seasonal fluctuations of freshwater fungi in River Nile, Egypt. Zeitschrift for Allgemeine Mikrobiologie, 22:521-527, 1982.
14. El-Hissy FT, Khallil AM : Studies on aquatic fungi in Delta region, Egypt. Zentralbl Mikrobiol, 144:421-432, 1989.
15. Elliot RF : Morphological variation in New Zealand Saprolegniaceae, 2. Saprolegnia terrestris Cookson and S australis sp nov. NZ J Bot, 6:94-105, 1968.
16. El-Nagdy MA : Studies on freshwater fungi in River Nile, M Sc Thesis, Dept of Botany, Fac of Science, Assiut Univ, 1981.
17. El-Nagdy MA : Studies on freshwater fungi in Upper Egypt. Ph D Thesis, Dept of Botany, Fac of Science, Assiut Univ, 1985.
18. El-Sharouny HM : Studies on root-infecting fungi with special reference to Pythium. Ph D Thesis, Dept of Botany, Fac of Science, Assiut Univ, 1980.

19. Fajola AD, Slasadura SD, Ogbonna CI : Some aquatic phycomycetes from riverine soils in Ibadan. *Nigeria Nova Hedwigia*, 29:905-911, 1978.
20. Fitzpatrick HM : *The lower fungi phycomycetes*. McGraw-Hill Book Company, INC. New York and London, p 331, 1930.
21. Forbes ET : Observations on some British water moulds (*Saprolegniales* and *Blastocladales*). *Trans Br Mycol Soc*, 19:221-239, 1935.
22. Forbes ET : Water moulds of the Manchester District. *Mem Proc Manchester Lit Phil Soc*, 79:1-11, 1935.
23. Höhnk W, Bock KJ : Ein Beitrag zur Ökologie der Saprophytischen Waspilze, *Veroeffent Inst Meeresforsch Bremerhaven*, 3:9-26, 1954.
24. Ismail SLA, Rattan SS, Muhsin TM : Aquatic Fungi of Iraq. species of *Saprolegnia*. *Hydrobiologia*, 65:83-93, 1979.
25. Jacobsen KR : *Thraustochytriaceae in ausgewählten Sanelwatten in horizontaler und vertikaler verteilung*. Diplomarbeit Pflanzenphysiol Instute der universität Göttingen, p 1385, 1981.
26. Jonhson TW Jr : The genus *Achlya*. Morphology and taxonomy. University of Michigan Ann Arbor, p 180, 1956.
27. Johnson TW Jr, Rogers AL, Beneke ES : Aquatic fungi of Iceland: comparative morphology of *Achlya radiosa*, *A pseudoradiosa* and *A stellata*. *Mycologia*, 67:108-199, 1975.
28. Jonhson TW Jr : Aquatic fungi of Scandinavia some species of *Aphanomyces*. *Bot Notiser*, 129:351-366, 1977.
29. Karling JS : Brazilian chytrids. VI. *Rhopatophlyctis* and *Chytromyces*, two new chitinophilic operaculate genera. *Amer J Bot*, 32:362-369, 1945.
30. Karling JS : Some zoosporic fungi of New Zealand. *Sydowia* 19:213-226, 1965.
31. Karling JS : *Chytridiomycetarum Iconographia Vaduz*. J Cramer, p 414, 1977.
32. Khallil MA : Field and laboratory studies on aquatic phycomycetes of Delta region. Ph D Thesis, Dept of Botany, Fac of Science, Assiut Univ, Egypt, 1987.
33. Klich MA, Tiffany LH : Distribution and seasonal occurrence of aquatic *Saprolegniaceae* in northwest Iowa. *Mycologia*, 77:373-380, 1985.
34. Knox JS, Paterson RA : The occurrence and distribution of some aquatic phycomycetes on Ross Island and the valleys of Victoria Land, Antarctica. *Mycologia*, 65:373-387, 1973.
35. Lund A : Studies on Danish freshwater phycomycetes and notes on their occurrence particularly relative to the hydrogen ion concentration of the water. *Mem Acad Roy Sci Denmark Sect Sci*, 6:1-97, 1934.
36. Lund A : Occurrence of *Saprolegniaceae* in Danish Soils. *Nova Hedwigia*, 39:377-395, 1978.
37. Milanez AI : *Achlya brasiliensis*. A new species from Brasil. *Rickia*, 2:183-189, 1965.
38. Milanez AI : Aquatic fungi of the cerrado region of Seapaulo state, I. First results. *Richia*, 3:97-109, 1968.
39. Misra JK : Occurrence, distribution and seasonality of aquatic fungi as affected by chemical factors in six alkaline ponds of India. *Hydrobiologia*, 97:185-191, 1982.
40. Naumov NA : Flora gribov leningradskoi oblasti vy pusk 1. *Arkhimiseti: Fkiomitseti*. (Flora of the fungi of the leningrad region. Part 1. *Archimycetes and phycomycetes*). Moscow and Leningrad Academy of Sci USSR, 1954.
41. Nolan RA : Physiological comparison of a new found land and a North Carolina isolate of *Saprolegnia australis*. *Mycologia*, 71:1136-1149, 1979.
42. Perrot PE : The ecology of some aquatic phycomycetes. *Trans Br Mycol Soc*, 43:19-30, 1979.
43. Rao AV, Venkateswarlu B : Microbial ecology of the soils of India desert. *Agric Ecosystems and Environment*, 10:361-389, 1983.
44. Rattan SS, Muhsin JM, Ismail ALS: Aquatic fungi of Iraq: species of *Dictyuchus* and *Calyptralegnie*. *Sydowia*, 31:112-121, 1978.
45. Rattan SS, Muhsin JM, Ismail ALS : Notes on the occurrence and seasonal periodicity of *saprolegniaceae* in shatt Al-Arab, Iraq. *Kavaka*, 8:41-46, 1980.
46. Rossy VC : Some water molds from Puerto Rico. *J Elisha Mitchell Sci Soc*, 72:129-137, 1970.
47. Scott WW : A monograph of genus *Aphnamyces*. *Virginia Agr Expt Sta Tech Bull*, 151:1-95, 1961.
48. Seymour RL : The genus *saprolegnia*. *Nove Hedwigia, Beiheft*, 19:1-124, 1970.
49. Sparrow FK : *Aquatic phycomycetes*. Uni of Michigan Press Ann Arbor, Second Ed, 25:1187, 1960.
50. Waterhouse GM : Some water moulds of the Hogsmill River collected from 1937 to 1939. *Trans Br Mycol Soc*, 25:315-325, 1942.
51. Waterhouse GM : Key to *Pythium pringsheim*. In *Mycol Inst Kew, Surrey, England*, pp 1-15, 1986.
52. Waterhouse GM : The genus *Pythium pringsheim*. Pages: 1-71. *Mycol pap*. 110, commonw Mycol Inst, Kew Surrey, England, 1968.
53. Willoughby LG : The ecology of some lower fungi at Esthwaite water. *Trans Br Mycol Soc*, 44:305-332,
54. Willoughby LG, Pickering AD : Viable *saprolegniaceae* spores on the epidemis of the salmonid fish *Salmotrutta* and *Salvelinus alpinus*. *Trans Br Mycol Soc*, 68:91-95, 1977.
55. Willoughby LG, McGrory CB, Pickering AD : Zoospore germination of *Saprolegnia* pathogenic to fish. *Trans Br Mycol Soc*, 80:421-435, 1983.
56. Youatt JB : Selective production of resistant sporangia in suspensions of *Allomyces*. *Trans Br Mycol Soc*, 75:334-336, 1980.
57. Yung C, Stenton H : A study of the phycomycetes in the soils of Hong Kong. *Trans Br Mycol Soc*, 47:127-139, 1964.
58. Zebrowska E : Mycoflora of several containers of the kampions Forest, Poland. *Acta Mycol*, 12:77-89, 1976.
59. Zucha H : *Mucorinea (kryptogamenfyora der Mark brandenburg) Band Via : Pilze II*, p 265, Printed in Germany, 1963.

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