

Prevalence of Extended-Spectrum Beta-Lactamase Producing *Klebsiella*: Cross-Sectional Study, 2000-2021

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ABSTRACT

Increasing resistance due to extended-spectrum β -lactamases (ESBLs) and multiple resistance mechanisms in gram-negative hospital isolates restrict the role of β -lactam antibiotics in empirical treatment of serious infections. As the prevalence of ESBL producing strains and resistance rates to antimicrobial agents can vary in each center, local surveillance studies are required to guide therapy. In this study, the prevalence of ESBL-K hospitalized population and the change of prevalence through years from 2000 to 2021.

Klebsiella strains isolated between 2000 and 2021 years, were included. 2000 data was collected from two hospitals; one state and private group hospitals. The other whole data were collected only from private group hospitals. ESBL tests were performed according to CLSI and EUCAST guidelines.

ESBL positive *Klebsiella* strains were mostly commonly isolated from intensive care units and from sputum + tracheal aspirate (%41). Total prevalence of ESBL positive *Klebsiella* strains were 51.29%. The prevalence of 2000 years was high probably due to the different hospital /patient profile. By excluding the 2000 data, the prevalence were increasing by years; 15.38% at 2001 to 61.50% at 2021.

High prevalence of ESBL in Turkey was increasing by years. Our private hospitals data was lower than the other state hospitals in the Turkey. Different hospital /patient profile could be the reason of low prevalence through precarity level, unnecessarily broad-spectrum antibiotic treatment, environmental contamination, kitchen hygiene and European health tourism. More research must be done to clarify the reason of this differences.

Keywords: Extended-spectrum beta-lactamase producing *Klebsiella*, Prevalence of ESBL-K, ESBL

Introduction

Extended-spectrum β -lactamase (ESBL) producing Enterobacteriaceae are an important reason for β -lactam antibiotics treatment failure (1). Asymptomatic carriage often precede infections due to ESBL-producing Enterobacteriaceae (ESBL-E) (2). Potential routes of transmission of ESBL-E to humans are the human-to-human transmission, food chain, direct contact with animals or indirectly via the environment (3).

ESBL-E are emerging worldwide and have been rated by the WHO as high priority pathogens among resistant bacteria (4).

Enterobacteriaceae group bacteria that exhibit resistance to beta-lactams, broad-spectrum beta-lactams, and third-generation cephalosporins are named as ESBL-E. ESBL-E such as TEM-1, TEM-2, SHV-1, and OXA-10 has been predominantly

associated with nosocomial outbreaks because of the resistance caused by point mutations and transferred by plasmid. The CTX-M-15 type ESBL-E is the most commonly identified and common in many countries in Europe, Asia, Africa and the United States (5-7). TEM and SHV-type ESBLs are most often found in *Escherichia coli* and *Klebsiella pneumoniae* TEM -lactamases have been found mainly in clinical isolates of *E. coli* (8, 9).

The majority of SHV-type ESBLs are found in strains of *K. pneumoniae*. The SHV-1 β -lactamase is most commonly found in *K. pneumoniae* and is responsible for up to 20% of the plasmid-mediated ampicillin resistance in this species (10, 11).

This study aimed to determine the prevalence for ESBL-producing *Klebsiella* (ESBL-K) in the Turkey hospitalized population and the change of prevalence by years between 2000-2021.

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Materials and Methods

Study Design and Participants: This was a cross-sectional study.

In our study, 35526 *Klebsiella* strains isolated from various clinical samples as an infectious agent between 2000 and 2021 years, were used.

In 2000, *Klebsiella* strains were collected from a government hospital, Ankara Training And Research Hospital and a private hospital groups, Acibadem Hospitals. After 2000, *Klebsiella* strains were collected from only Acibadem Hospitals.

Procedures: Initially, susceptibility tests were performed with the VITEK 2 (BioMérieux, France) system and ESBL results were evaluated by expert system of VITEK 2 according to manufacturer's recommendations. When the ESBL results were suspicious, the Double Disc Synergy Test (DDST) or Combine Disc Test (CDT) were applied as ESBL confirmations tests according to CLSI and EUCAST guidelines (12,13).

To perform these manual tests, bacterial suspension prepared in 0.5 McFarland turbidity was spread to Mueller-Hinton Agar medium by using sterile swab.

In DDST; after placing an amoxicillin clavulanic acid (AMC) (20/10 µg) disc in the centre of the petri dish and placing ceftazidime (CAZ) (30 µg), ceftriaxone (CRO) (30 µg), cefepim (FEP) (30 µg), cefotaxime (CTX) (30 µg), aztreonam (ATM) (30 µg) radially at a distance of 25 mm from AMC's disc circumference, an expansion towards the AMC disk in the inhibition zones around the CAZ, CRO, FEP, CTX or ATM discs or the presence of a non-bacterial synergy area evaluated as ESBL production. In CDT; ceftazidime (CAZ) (30 µg), ceftazidime-clavulanic acid (CCA) (30/10 µg), cefotaxime (CTX) (30 µg), cefotaxime-clavulanic acid (CCT) (30/10 µg) were used. CAZ and CCA discs were placed in the petri dish with 30 mm distance and same procedure was applied to CTX and CCT discs.

Petri dishes were incubated at 35°C for 18 hours and results were evaluated according to EUCAST criteria. 5 mm or more difference between cephalosporin disc and cephalosporin -clavulanate disc was evaluated as ESBL production (12,13).

Results

Demographic informations: Patients included in the study were 0-94 years old and 72% patients were male, 28% female.

ESBL K strains were commonly isolated from intensive care units (30,0%), then respectively; surgical departments (22,0%), hematology-oncology departments (16%), internal medical departments (14%), emergency departments (12,0%) and department of pediatrics (6,0%) (Table1).

The patient sample types that most ESBL-K isolated that ESBL-K strains isolated were sputum and/or tracheal aspirate (%41) , blood (37%)and urine (%22) (Table2).

The number of *Klebsiella* isolated was increasing by years. It was 328 at 2000 and 3978 at 2021.

The number of *Klebsiella* isolated from in-patients was also progressively increasing by years. It was 172 at 2000 and 2678 at 2021

Total prevalence of ESBL positive *Klebsiella* strains were 51.29%.

Year 2000 data was collected from two hospitals groups; one state and private group hospitals. The other whole data were collected only from private group hospitals. The prevalence of ESBL-K (isolated ESBL-*Klebsiella* number from in-patient/ isolated *Klebsiella* number from in-patient %) were also increasing by years. But at year 2000, the ratio was similar as 2014. This could be due to difference at hospital/patient population. By excluding the 2000 data, the ratio was increased from 15.38% at 2001 to 61.50% at 2021 (Table 3).

Discussion

Since possessing bactericidal effect and low side effects, broad-spectrum beta-lactam antibiotics are often preferred. On the other hand, due to increased clinical use of these drugs the resistance to beta-lactamases has been appeared. ESBL-producing strains cause mortality and serious economic losses (14-16).

Since 2000, the prevalence of ESBL-K has been increasing worldwide with large geographical variations (17). Turkey is one of the countries that possess the increased ESBL-K prevalence due to the redundant use of antibiotics [18-20]. In a study published in 2001, it was reported that prevalence of ESBL positivity was reported higher among *K. pneumoniae* than among *E. coli* strains (19).

Our study was set out to obtain a representative sample of the country's general population. In all, 35526 strains were included and the total prevalence was 51.29%, which is comparable to lower than the other studies in the Turkey. But although the ESBL positivity ratio was lower than

Table 1. Distribution of ESBL Positive *Klebsiella* According To Clinical Departments

Departments	ESBL positive ratio %
Intensive care units	30,0
Surgical departments	22,0
Hematology-oncology departments	16,0
Internal medical departments	14,0
Emergency departments	12,0
Department of pediatrics	6,0

Table 2. Distribution of ESBL Positive *Klebsiella* According To The Most Common Patient Sample Types

Sample type	ESBL positive ratio %
Sputum + Tracheal aspirate	41,0
Blood	37,0
Urine	22,0

average of the country, it was increasing by every year as expected from 15,0 to 61,0.

Studies have shown that these rates can be variable depending on the country and region. At a multi-center study (MYSTIC) Gur *et al.* found that hospital acquired ESBL-K strains ratio were 40.5% in 2004-2005. In mystic study, already results of the two centers were higher than the other centers. All the hospitals occupied in the study were state hospitals. In our study, 2000 data collected from state and private hospitals were higher than other year's data collected from just private hospitals; similar as MYSTIC study (21).

In the other study published in 2007, Guducuoglu *et al.* reported that ESBL production was 63% in hospitalized patients while this ratio was 30% in outpatients (23). Studies also published at 2000, 2001 and 2009 yr in Turkey present different prevalence ratio 22%, 33% and 44%. When the studies conducted in our country are examined, it is seen that there has been an increase in ESBL production rates over the years (14, 22, 23, 24).

In the European population, the prevalence of ESBL-producing Enterobacterales colonization in the community ranges from 6% to 11%.4, in hospitalized patients has been reported to be as high as 13% in some regions (25, 26). In the study conducted in Germany at 2007-2011 yr, the prevalence of ESBL-K was reported as 13,8%, 15% and 11,7% in two-year periods (2007-2008, 2009-2010 and 2011-2012, respectively) (27).

Known specific risk factors for ESBL-producing bacterial infections are ESBL-E carriage, age >55 years, male sex (for urinary tract infections) (28-30), precarity level (31,32), antibiotic treatment in the past 1-3 months (33-36), particularly broad-

spectrum antibiotics (36-40), recent hospitalization (33), other healthcare activities (e.g. urinary catheter), environmental contamination, kitchen hygiene, ESBL-E carriage in poultry, pig and cattle farms and travelling to endemic areas as Africa and Asia (37-43). Other studies reported that the risk of ESBL-E is higher after swimming in freshwater (44, 45). Moreover, agricultural land may be contaminated through the practice of spreading livestock manure (46, 47). ESBL has also been detected in aquatic environments close to healthcare centers. Altogether, these results suggest that the characteristics (agricultural, environmental or healthcare related) of the area where people live can play a major role in the risk of ESBL-producing *Klebsiella* infections (44-48).

High precarity level, low unnecessary take of antibiotic treatment, high kitchen hygiene, low environmental contamination and high number of foreign patients especially European and Balkan states could be the reason of the low ESBL positivity ratio in our study.

Globally, ESBL-E prevalence varies from 2 to 46% between communities from different subregions (49). Every year ESBL-E carriage rates increase worldwide with more than 5% among healthy individuals (49). Also in Europe, an increase in ESBL-E community carriage rates has been documented over the past years (50). Three studies found an ESBL-E prevalence of 4.5-8.6% among healthy individuals. Two of these identified travel to Asia or Africa in the previous 12 months and the use of proton pump inhibitors (PPI's) to be associated with a higher risk for ESBL-E carriage in the community. Other risk factors were the use of antimicrobials, travel to North and

Table 3: The Total Number of Isolated *Klebsiella*, The Number of *Klebsiella* Isolated From In-Patients, The Number and The Prevalence of ESBL-K In-Patients

Year	Isolated <i>Klebsiella</i> number	Isolated <i>Klebsiella</i> number from in-patient	Isolated ESBL-K number from in-patient	Prevalence of ESBL-K %, in-patients
2000	328	172	74	43.02
2001	179	78	12	15.38
2002	229	84	15	17.78
2003	297	89	18	20.22
2004	361	123	27	21.89
2005	461	131	30	23.12
2006	497	138	32	23.18
2007	521	154	38	24.67
2008	721	190	43	22.63
2009	565	202	74	36.63
2010	904	309	75	24.27
2011	1936	590	183	31.01
2012	1642	836	95	11.36
2013	1617	835	265	31.73
2014	2415	1165	478	41.03
2015	2493	1527	852	55.79
2016	2837	1699	986	58.03
2017	2991	1782	968	54.32
2018	3237	1979	1184	59.82
2019	3727	2467	1509	61.17
2020	3590	2380	1451	60.97
2021	3978	2678	1647	61.50
Total	35526	19608	10056	51.29

Latin America, keeping cows, living in the proximity of a mink farm, and owning or having contact with a horse (51-53). In countries with similar ESBL-E community carriage rates as the Netherlands, previous antibiotic use was identified as a predictor in Japan, Germany and France (49, 53, 54). Travel to Asia or Africa and travel to Africa or Greece were identified as predictors for ESBL-E carriage in Swedish and German communities, respectively (54). Overall, studies found a variety of risk factors. Therefore, elucidation of risk factors is needed to identify definitive sources for ESBL-E carriage in the community and to foresee possible public health risks and interventions.

In conclusion, ESBL production rates of ESBL-producing *K. pneumoniae* that cause infections which are expensive and difficult to treat, should be monitored by each center. Furthermore, broad-spectrum beta-lactam antibiotics preferred in the treatment of infections should be used carefully and hospitalized patients should be isolated.

Finally we suggest the surveillance work should be done in hospital departments at risk.

References

1. Pitout JDD and Laupland KB (2008) Extended-spectrum beta-lactamase-producing Enterobacteriaceae: an emerging public-health concern. *The Lancet Infectious Diseases* 8, 159–166.
2. Freeman JT et al. (2012) Bloodstream infection with extended-spectrum beta-lactamase-producing Enterobacteriaceae at a tertiary care hospital in New Zealand: risk factors and outcomes. *International Journal of Infectious Diseases* 16, e371–e374.
3. Huijbers PM et al. (2015) Role of the environment in the transmission of antimicrobial resistance to humans: a review. *Environmental Science & Technology* 49, 11993–12004.
4. Tacconelli E, Carrara E, Savoldi A et al. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-

- resistant bacteria and tuberculosis. *Lancet Infect Dis* 2018; 18: 318–27.
5. Paterson DL, Bonomo RA, Extended-spectrum b-lactamases: a clinical update. *Clin Microbiol Rev* 2005; 18(4):1657–686.
 6. European Centre for Disease Prevention and Control. Antimicrobial Resistance Surveillance in Europe 2015. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). Stockholm: European Centre for Disease Prevention and Control; 2017.
 7. Lee MY, Ko KS, Kang CI, Chung DR, Peck KR et al. High prevalence of CTX-M-15-producing *Klebsiella pneumoniae* isolates in Asian countries: diverse clones and clonal dissemination. *Int J Antimicrob Agents* 2011;38:160–163.
 8. Bret, L., C. Chanel, D. Sirot, R. Labia, and J. Sirot. Characterization of an inhibitor-resistant enzyme IRT-2 derived from TEM-2 β -lactamase produced by *Proteus mirabilis* strains. *J. Antimicrob. Chemother.* 1996. 38:183– 191.
 9. Lemozy, J., D. Sirot, C. Chanal, C. Hue, R. Labia, H. Dabernat, and J. Sirot. First characterization of inhibitor-resistant TEM (IRT) B-lactamases in *Klebsiella pneumoniae* strains. *Antimicrob. Agents Chemother.* 1995; 33:2580–2582.
 10. Tzouveleki, L. S., and R. A. Bonomo. SHV-type β -lactamases. *Curr. Pharm. Des.*1999; 5:847–864.
 11. Bradford PA, Extended-Spectrum β -lactamases in the 21st Century: Characterization, Epidemiology, and Detection of This Important Resistance Threat. *Clin Microbiol Rev.* 2001;14(4):933-51.
 12. Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing. 24th Informational Supplement. CLSI Document M100-S24, CLSI, Wayne,
 13. European Committee on antimicrobial Susceptibility Testing (EUCAST) (2012). Guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance.
 14. Mumcuoğlu İ. *Klebsiella* ve *Proteus* suşlarında genişlemiş spektrumlu beta-laktamaz varlığı ve çeşitli antibiyotiklere direnç durumu. *ANKEM Dergisi* 2004;18:9-11.
 15. Esen Ş, Eroğlu C, Sünbül M ve ark. *Escherichia coli* ve *Klebsiella pneumoniae* suşlarında TEM ve SHV türü beta laktamazların sıklığı. *Mikrobiyol Bülteni* 2001;35:37-43.
 16. Koçoğlu E, Karabay O, Koç İnce N ve ark. Toplum kaynaklı üriner sistem enfeksiyonlarından izole edilen *Escherichia coli* suşlarında genişlemiş spektrumlu beta laktamaz ve bazı antibiyotiklere direnç sıklığının araştırılması. *ANKEM Dergisi* 2007;21:5-9.
 17. Kahlmeter G, Poulsen HO. Antimicrobial susceptibility of *Escherichia coli* from community-acquired urinary tract infections in Europe: the ECOSENS study revisited. *Int J Antimicrob Agent* 2012; 39: 45–51.
 18. Philippon A, Labia R, Jacoby G. Extended spectrum beta-lactamases. *Antimicrob Agents Chemother* 1989;33: 1131-1136.
 19. Öksüz L, Gürler N, Akıncı N, Şirin A. İki aylık bir dönemde pediatrik poliklinik hastalarının idrar örneklerinden izole edilen GSBL oluşturan *Escherichia coli* ve *Klebsiella pneumoniae* suşları. *ANKEM Dergisi* 2008;22: 14-19.
 20. Del Carmen Rodriguez M, Vera DE, Ramirez-Ronda CH, et al. Phenotypic confirmation of extended spectrum beta lactamases (ESBL) in *Escherichia coli* and *Klebsiella pneumoniae* at the San Juan Veterans Affairs Medical Center. *P R Health Sci J* 2004;23:207-215.
 21. Gur D, Gülay Z, Arıkan Akan Ö, et al. Resistance to newer beta-lactams and related ESBL types in gram negative nosocomial isolates in Turkish hospitals: results of the multicenter Hitit study. *Mikrobiyol Bul* 2008; 42: 537-544.
 22. Albayrak N, Kaya Ş. Çeşitli klinik örneklerden izole edilen *Escherichia coli* ve *Klebsiella pneumoniae* suşlarının genişlemiş spektrumlu beta laktamaz üretimleri ve antibiyotik direnç oranları. *Türk Mikrobiyol Cem Dergisi* 2009;39:16-21.
 23. Güdücüoğlu H, Baykal S, İzci H ve ark. Genişlemiş spektrumlu beta-laktamaz (GSBL) üreten *Escherichia coli* ve *Klebsiella pneumoniae* suşlarının antibiyotiklere direnci. *ANKEM Dergisi* 2007;21:155-160.
 24. Yurttutan N, Aksaray S. Hastane kaynaklı *Klebsiella* suşlarında genişlemiş spektrumlu β -laktamaz araştırılması. *Tıpta Uzmanlık Tezi, T.C. Sağlık Bakanlığı Ankara Numune Eğitim ve Araştırma Hastanesi, Mikrobiyoloji ve Klinik Mikrobiyoloji Bölümü*; 2000.
 25. Hamprecht A, Rohde AM, Behnke M et al. Colonization with thirdgeneration cephalosporin-resistant Enterobacteriaceae on hospital admission: prevalence and risk factors. *J Antimicrob Chemother* 2016; 71: 2957–63.
 26. Hagel S, Makarewicz O, Hartung A et al. ESBL colonization and acquisition in a hospital population: the molecular epidemiology and transmission of resistance genes. *PLoS One* 2019; 14: e0208505.
 27. Leistner R, Schröder C, Geffers C, Breier AC, Gastmeier P, Behnke M. Regional distribution of nosocomial infections due to ESBL-positive Enterobacteriaceae in Germany: data from the German National Reference Center for the Surveillance of Nosocomial Infections (KISS). *Clin Microbiol Infect* 2015;21:255.e1–5.
 28. Park SH, Choi S-M, Lee D-G et al. Impact of extended-spectrum b-lactamase production on treatment outcomes of acute pyelonephritis caused by *Escherichia coli* in patients without health care-associated risk factors. *Antimicrob Agents Chemother* 2015; 59: 1962–8.

29. Søgaard M, Heide-Jørgensen U, Vandenbroucke JP et al. Risk factors for extended-spectrum b-lactamase-producing *Escherichia coli* urinary tract infection in the community in Denmark: a case-control study. *Clin Microbiol Infect* 2017; 23: 952–60.
30. Pe´rez Heras I, Sanchez-Gomez JC, Beneyto-Martin P et al. Community-onset extended-spectrum b-lactamase producing *Escherichia coli* in urinary tract infections in children from 2015 to 2016: prevalence, risk factors, and resistances. *Medicine (Baltimore)* 2017; 96: e8571.
31. Alividza V, Mariano V, Ahmad R et al. Investigating the impact of poverty on colonization and infection with drug-resistant organisms in humans: a systematic review. *Infect Dis Poverty* 2018; 7: 76.
32. Nomamiukor BO, Horner C, Kirby A et al. Living conditions are associated with increased antibiotic resistance in community isolates of *Escherichia coli*. *J Antimicrob Chemother* 2015; 70: 3154–8.
33. Hertz FB, Schønning K, Rasmussen SC et al. Epidemiological factors associated with ESBL- and non ESBL-producing *E. coli* causing urinary tract infection in general practice. *Infect Dis (Lond)* 2016; 48: 241–5.
34. Toumi A, Hafsa M, Kadri Y. Risk factors for community-acquired acute pyelonephritis caused by extended-spectrum b-lactamase-producing *Escherichia coli* in a university hospital in Tunisia. Twenty-Fifth European Congress of Clinical Microbiology and Infectious Diseases, Copenhagen, Denmark, 2015. Abstract P2143.
35. Castillo-Tokumori F, Irey-Salgado C, Ma´laga G. Worrysome high frequency of extended-spectrum b-lactamase-producing *Escherichia coli* in community-acquired urinary tract infections: a case-control study. *Int J Infect Dis* 2017; 55: 16–9.
36. Kang C-I, Wi YM, Lee MY et al. Epidemiology and risk factors of community onset infections caused by extended-spectrum b-lactamase-producing *Escherichia coli* strains. *J Clin Microbiol* 2012; 50: 312–7.
37. Søraas A, Sundsfjord A, Sandven I et al. Risk factors for community-acquired urinary tract infections caused by ESBL-producing Enterobacteriaceae—a case-control study in a low prevalence country. *PLoS One* 2013; 8: e69581.
38. Calbo E, Romani´ V, Xercavins M et al. Risk factors for community-onset urinary tract infections due to *Escherichia coli* harbouring extended-spectrum b-lactamases. *J Antimicrob Chemother* 2006; 57: 780–3.
39. Azap O`K, Arslan H, S, erefhanoglu K et al. Risk factors for extended-spectrum b-lactamase positivity in uropathogenic *Escherichia coli* isolated from community-acquired urinary tract infections. *Clin Microbiol Infect* 2010; 16: 147–51.
40. Ozdogan FN, Demirdal T, Nemli SA, Risk factors for community acquired urinary tract infections caused by extended spectrum b-lactamase (ESBL) producing *Escherichia coli*. Twenty-Fifth European Congress of Clinical Microbiology and Infectious Diseases, Copenhagen, Denmark, 2015. Abstract P0794.
41. Tnsawai U, Walsh TR, Niumsup PR. Extended spectrum b-lactamase-producing *Escherichia coli* among backyard poultry farms, farmers, and environments in Thailand. *Poult Sci* 2019; 98: 2622–31.
32. Liu Z, Wang Y, Walsh TR et al. Plasmid-mediated novel blaNDM-17 gene encoding a carbapenemase with enhanced activity in a sequence type 48 *Escherichia coli* strain. *Antimicrob Agents Chemother* 2017; 61: e02233–16.
42. Pruthivishree BS, Vinodh Kumar OR, Sinha DK et al. Spatial molecular epidemiology of carbapenem-resistant and New Delhi metallo b-lactamase (blaNDM)-producing *Escherichia coli* in the piglets of organized farms in India. *J Appl Microbiol* 2017; 122: 1537–46.
43. Yaici L, Haenni M, Saras E et al. blaNDM-5-carrying IncX3 plasmid in *Escherichia coli* ST1284 isolated from raw milk collected in a dairy farm in Algeria. *J Antimicrob Chemother* 2016; 71: 2671–2.
44. Machado E, Coque TM, Canto´n R et al. Leakage into Portuguese aquatic environments of extended-spectrum-b-lactamase-producing Enterobacteriaceae. *J Antimicrob Chemother* 2009; 63: 616–8.
45. Dhanji H, Murphy NM, Akhigbe C et al. Isolation of fluoroquinolone-resistant O25b: h 4-ST131 *Escherichia coli* with CTX-M-14 extended-spectrum b-lactamase from UK river water. *J Antimicrob Chemother* 2011; 66: 512–6.
46. Hartmann A, Locatelli A, Amoureux L et al. Occurrence of CTX-M Producing *Escherichia coli* in soils, cattle, and farm environment in France (Burgundy region). *Front Microbiol* 2012; 3: 83.
47. Touati M, Hadjadj L, Berrazeg M et al. Emergence of *Escherichia coli* harbouring mcr-1 and mcr-3 gene in North West Algerian farmlands. *J Glob Antimicrob Resist* 2020; 21: 132–7.
48. Galvin S, Boyle F, Hickey P et al. Enumeration and characterization of antimicrobial-resistant *Escherichia coli* bacteria in effluent from municipal, hospital, and secondary treatment facility sources. *Appl Environ Microbiol* 2010; 76: 4772–9.
49. Karanika S, Karantanos T, Arvanitid M, Grigoras C, Mylonakis E. Fecal colonization with extended-spectrum beta-lactamase-producing Enterobacteriaceae and risk factors among healthy individuals: a systemic review and meta-analysis. *Clin Infect Dis: Off Publ Infect Dis Soc Am* 2016; 63(3):310-8.

50. Woerther PL, Burdet C, Cahchaty E, Andremont A. Trends in human fecal carriage of extended-spectrum beta-lactamase in the community: towards the globalization of CTX-M. *Clin Microbiol Rev* 2013;26(4):744-58.
51. Huibers PM, de Kraker M, Grat EA, van Hoek AH, van Santen MG, de Jong MC, et al. Prevalence of extended-spectrum beta-lactamase-producing Enterobacteriaceae in humans living in municipalities with high and low broiler density. *Clin Microbiol Infect: Off Publ Eur Soc Clin Microbiol Infect Dis* 2013;19(6):E256-9.
52. Reulan EA, Al Naime N, Kaiser AM, Heck M, Kluytmans JA, Savelkoul PH, et al. Prevalence and risk factors for carriage of extended-spectrum beta-lactamase-producing Enterobacteriaceae in Amsterdam. *J Antimicrob Chemother* 2016;71(4):1076-82.
53. Wienders CCH, van Hoek A, Hengeveld PD, Veenman C, Dierikx CM, Zomer TP, et al. Extended-spectrum beta-lactamase- and pAmpC-producing Enterobacteriaceae among the general population in a livestock-dense area. *Clin Microbiol Infect: Off Publ Eur Soc Clin Microbiol Infect Dis* 2017;23(2):120e1-e8.
54. Luvsansharav UO, Hirai I, Niki M, Nakata A, Yoshinaga A, Moriyama T, et al. Prevalence of fecal carriage of extended-spectrum beta-lactamase-producing Enterobacteriaceae among healthy adult people in Japan. *J Infect Chemother: Off J Jpn Soc Chemother* 2011;17(5):722-5.
55. Meyer E, Gastmeier P, Kola A, Schwab F. Pet animals and foreign travel are risk factors for colonization extended-spectrum beta-lactamase-producing E. Coli. *Infect* 2012;40(6):685-7.